

Synthesis and Bioactivity of β -(1 \rightarrow 4)-Linked Oligomannoses and Partially Acetylated Derivatives

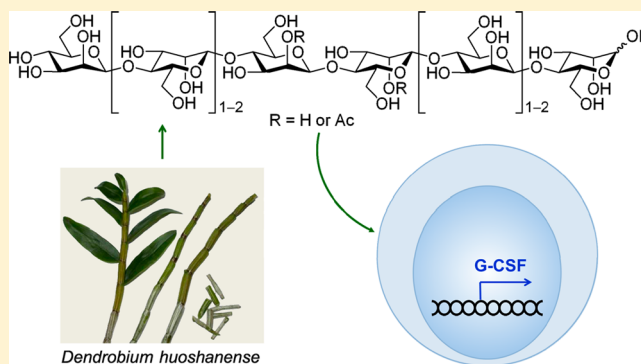
Keiichiro Ohara,[†] Chih-Chien Lin,[†] Pei-Jung Yang,[†] Wei-Ting Hung,[†] Wen-Bin Yang,[†] Ting-Jen Rachel Cheng,[†] Jim-Min Fang,^{*,†,‡} and Chi-Huey Wong^{*,†}

[†]The Genomics Research Center, Academia Sinica, Taipei 115, Taiwan

[‡]Department of Chemistry, National Taiwan University, Taipei 106, Taiwan

S Supporting Information

ABSTRACT: The synthesis of β -(1 \rightarrow 4)-linked hexa- to octamannoses and their partially acetylated derivatives was efficiently carried out by assembly of appropriate oligomeric fragments using β -selective glucosylation followed by gluco to manno epimerization at a late stage of the synthetic pathway. In the course of this study, we also observed that 2-*O*-acetylated oligomannoses coexisted in equilibrium with the 3-*O*-acetylated isomers due to intramolecular migration of the acetyl group. Bioactivity of the synthetic oligomannoses and partially acetylated derivatives was investigated in order to identify the possible smallest oligomer for induction of cytokines as that shown in the polysaccharides extracted from *Dendrobium huoshanense*.



INTRODUCTION

The Chinese herb plant *Dendrobium huoshanense* is used as traditional medicine in China for nourishment of the stomach, improvement in circulation of body fluids, and enhancement of the immune system.¹ It was found that *D. huoshanense* exerts important bioactivity such as antiaging, anticancer, and antidiabetes effects.² For this reason, many research efforts have been devoted to find bioactive compounds from *D. huoshanense*.³ We previously reported a bioactive polysaccharide extracted from the aerial parts of *D. huoshanense*.⁴ The polysaccharide is composed of β -(1 \rightarrow 4)-glucomanan having partial acetylation (~35%) at the 2- and 3-positions of mannosides in an approximate ratio of 1:10 glucoside to mannoside. Our study revealed that the polysaccharide exhibits specific functions in stimulating murine splenocytes to produce several cytokines including IFN- γ , IL-10, IL-6, IL-1 α , GM-CSF, and G-CSF. Among the cytokines examined, a relatively high increase of G-CSF was observed at transcriptional and translational levels.

G-CSF is a hematopoietic growth factor of myeloid lineage.⁵ Because of its many important roles in granulopoiesis,⁶ G-CSF has currently been used for overcoming granulocytopenia, which is one of the most serious side effects for cancer patients treated with chemotherapies or radiotherapies. In addition, G-CSF exerts profound immunoregulatory effects in adaptive immunity^{6b} and has neuroprotective activities in cerebral ischemia and neurodegeneration.⁷ Therefore, agents inducing G-CSF production have potential therapeutic applications in autoimmune disease and neurological disorders, which prompted us to investigate a minimal structural requirement

responsible for the effect of the polysaccharide on induction of G-CSF.

In our preliminary study,⁸ it was found that a mixture of oligomers generated by digestion of the natural polysaccharides using hemicellulase could induce expression of G-CSF. Our study also showed that a mixture of β -(1 \rightarrow 4)-linked oligomannoses with the number of mannose unit greater than 6 had G-CSF induction activity. However further study was hampered due to the difficulties in separating and isolating these oligosaccharides. For this reason, we chose to synthesize hexa-, hepta-, and octamannoses 1, 4, and 7 (Figure 1) for detailed structure–activity relationship studies. We also envisioned a synthesis of partially acetylated oligomannosides 2, 3, 5, 6, and 8 (Figure 1) to assess the effect of the acetyl group on the bioactivity. In the natural polysaccharides of *D. huoshanense*, the C-2 hydroxyl groups in the mannose residues are acetylated more abundantly than the C-3 hydroxyl groups. Thus, diacetylated oligomannoses 3 and 6 were designed to mimic the natural oligosaccharides. Monoacetylated oligomannoses 2, 5, and 8 were synthesized to evaluate the influence of the acetyl groups on the bioactivity.

RESULTS AND DISCUSSION

Synthesis of Hexasaccharides 1–3. Although synthesis of the related nonacetylated β -(1 \rightarrow 4)-linked hexamannoside has been reported,⁹ the method involving a procedure to remove the ester-type protecting groups at the final stage

Received: March 12, 2013

Published: June 10, 2013

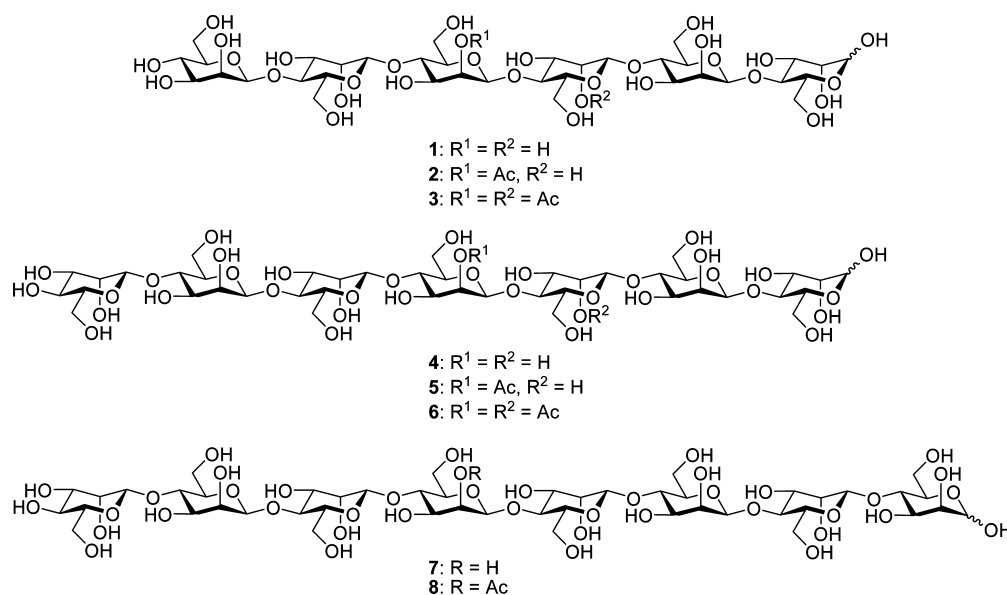
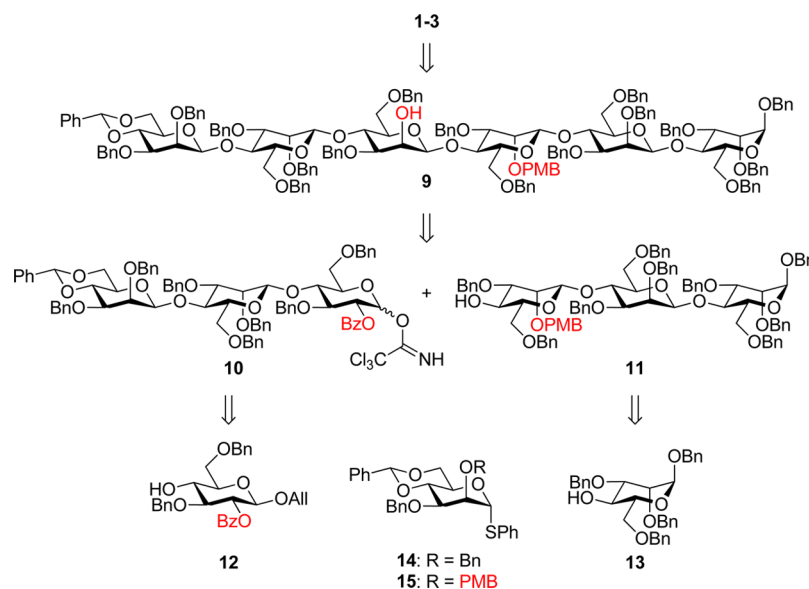


Figure 1. Oligomannoses 1–8 as the target molecules for inducing the expression of granulocyte colony-stimulating factor (G-CSF).

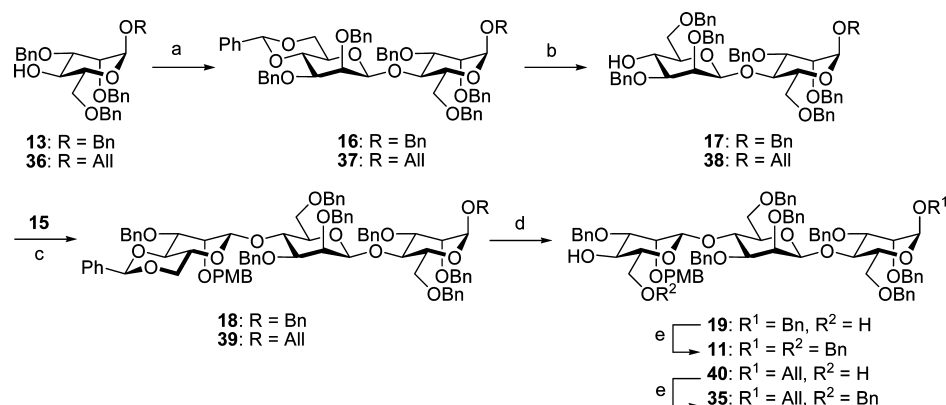
Scheme 1. Retrosynthetic Analysis of Hexasaccharides 1–3



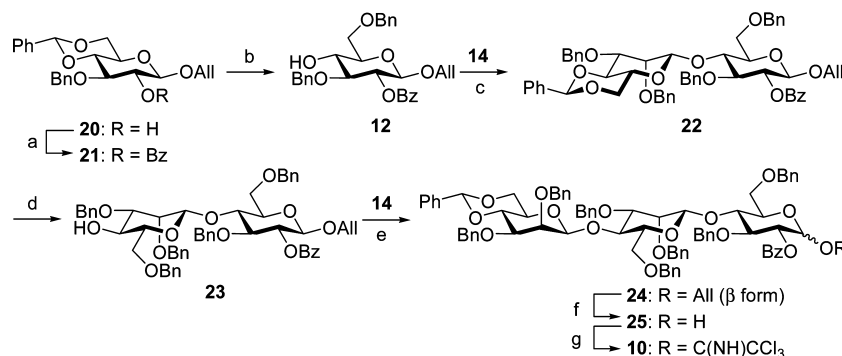
cannot be applied to the synthesis of acetylated hexasaccharides 2 and 3. We thus developed an alternative synthetic route for hexamannosides 1–3. The retrosynthetic analysis (Scheme 1) shows that hexasaccharides 1–3 can be derived from a common intermediate 9. Considering the overall efficiency, it is desirable to synthesize the key intermediate 9 by a [3 + 3] glycosylation. Even though several methods for direct β -selective mannosylation have been developed,¹⁰ direct formation of β -oligomannosides¹¹ is still difficult because properly protected mannosyl donors are often required for the coupling reactions with oligosaccharide fragments.¹² We chose to synthesize the hexamannoside intermediate 9 by an indirect method involving β -selective glucosylation of trimannoside acceptor 11 with a trisaccharide donor 10. The subsequent epimerization of the C-2 hydroxyl group in the glucoside moiety would be performed via an oxidation–reduction sequence to afford the desired configuration of β -mannoside. Furthermore, this methodology allows installation of an acetyl

group at the designated position. The trisaccharide fragments 10 and 11 can be respectively obtained from monosaccharides 12 and 13¹³ by repeating the β -selective mannosylations using 14¹⁴ or 15¹⁴ as the donors. To facilitate the [3 + 3] glycosylation for the assembly of oligosaccharide fragments, the trichloroacetimidate method is preferable to attain high efficiency.¹⁵ Although gluco to manno epimerization in large oligosaccharides seems to be difficult, some successful examples for synthesis of complex oligosaccharides incorporating such epimerization at a late stage have been demonstrated.¹⁶

We first undertook the synthesis of trisaccharide acceptor 11 (Scheme 2), which proceeded with a β -selective mannosylation of 13 with donor 14 using 1-benzenesulfonyl piperidine (BSP) and trifluoromethanesulfonic anhydride (Tf₂O) as the combined promoters^{10c} in the presence of a bulky base 2,4,6-tri-*tert*-butylpyrimidine (TTBP)¹⁷ to afford dimannoside 16 in 73% yield. The reductive ring-opening of benzylidene acetal occurred regioselectively by using Et₃SiH and BF₃·OEt₂¹⁸ to

Scheme 2. Synthesis of Trisaccharide Acceptors 11 and 35^a

^aReagents and conditions: (a) 14, BSP, Tf₂O, TTBP, CH₂Cl₂, MS3A, -78 °C to rt, 5–6 h; 73% for 16, 79% for 37; (b) Et₃SiH, BF₃·OEt₂, CH₂Cl₂, 0 °C, 7 h; 62% for 17, 60% for 38; (c) 15, BSP, Tf₂O, TTBP, CH₂Cl₂, MS3A, -78 to -60 °C, 2–5 h; Et₃N, (EtO)₃P, -60 °C to rt, 15–20 min; 65% for 18, 53% for 39; (d) CSA, neopentyl glycol, CH₂Cl₂, rt, 24–35 h; 74% for 19 from 18, 72% for 40 from 39; (e) Bu₂SnO, toluene, reflux, 3 h; BnBr, CsF, toluene, reflux, 6 h; 67% for 11, 80% for 35.

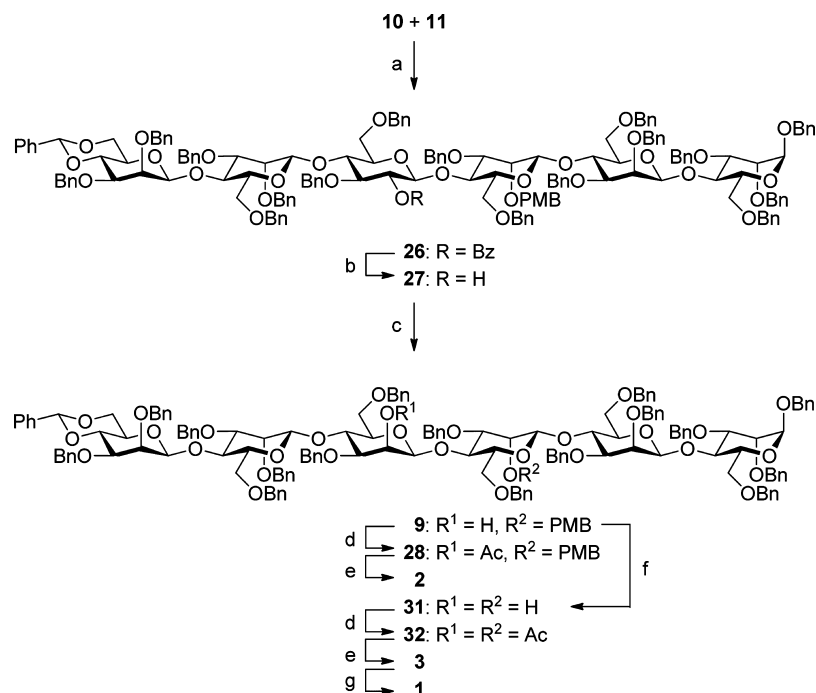
Scheme 3. Synthesis of Trisaccharide Donor 10^a

^aReagents and conditions: (a) BzCl, pyr, 0 °C to rt, 10 h; 99%; (b) Et₃SiH, TFA, CH₂Cl₂, 0 °C, 1 h; 88%; (c) 14, BSP, Tf₂O, TTBP, CH₂Cl₂, MS3A, -78 to -60 °C, 5 h; Et₃N, (EtO)₃P, -60 °C to rt, 15 min; 71%; (d) Et₃SiH, BF₃·OEt₂, CH₂Cl₂, 0 °C, 1.5 h; 74%; (e) 14, BSP, Tf₂O, TTBP, CH₂Cl₂, MS3A, -78 to -60 °C, 2 h; Et₃N, (EtO)₃P, -60 °C to rt, 20 min; 53%; (f) PdCl₂, MeOH/CH₂Cl₂ (1:1, v/v), rt, 6 h; 77%; (g) CCl₃CN, DBU, CH₂Cl₂, 0 °C, 1 h; 72%.

give the desired alcohol 17 in 62% yield. Mannosylation of 17 with donor 15 by activation of BSP/Tf₂O furnished trimannoside 18. It was noted that the addition of triethylamine and triethyl phosphite¹⁹ for quenching the transiently formed electrophilic species, (piperidino)phenyl(*S*-thiophenyl)sulfide triflate, was required to avoid the decomposition of compound 18. Due to the instability of the PMB ether against BF₃·OEt₂,²⁰ conversion of 18 to 11 could not be realized by using Et₃SiH/BF₃·OEt₂ to effect a direct reductive opening of the benzylidene acetal. Alternatively, we carried out this conversion through two steps: transacetalation^{9,21} using neopentyl glycol in the presence of camphorsulfonic acid (CSA) and selective monobenylation of the resulting diol 19 via formation of a stannylene acetal.²²

The synthesis of trisaccharide donor 10 began with benzylation of allyl glucoside 20²³ to give compound 21 (Scheme 3). Opening of the benzylidene acetal using Et₃SiH and TFA²⁴ provided the desired 4-hydroxy derivative 12. Preparation of trisaccharide 24 from monosaccharide 12 was achieved in a three-step sequence similar to that for conversion of 13 to 18. After removal of the allyl group with PdCl₂,²⁵ hemiacetal 25 was subject to trichloroacetimidation to afford the trisaccharide donor 10.

Having trisaccharide donor 10 and trisaccharide acceptor 11 in hand, assembly of these fragments was accomplished by treatment with a catalytic amount of TMSOTf to afford hexasaccharide 26 (Scheme 4). The benzoyl group was removed using LiAlH₄^{16a,b} to obtain hexasaccharide 27 containing a free hydroxyl group at C-2 of the central glucoside residue. To our delight, Albright–Goldman oxidation²⁶ of alcohol 27 and the subsequent reduction of the resulting ketone using NaBH₄²⁷ afforded the key intermediate 9 having an all-manno configuration. As shown by TLC analysis, the reduction product also contained a small amount of 27 having a gluco configuration at the central saccharide moiety. To verify the all-β-manno pyranoside structure of 9, we determined the ¹J_{CH} values between the anomeric carbons and protons. The anomeric C–H coupling constants of the mannoside residues, except for that at the reducing end, are around 160 Hz, in agreement with the β-manno pyranoside backbone of hexasaccharide 9.²⁸ Our original attempt to prepare non-acetylated hexamer 1 by hydrogenation of 9 encountered difficulties in workup and purification. Alternatively, we considered preparation of 1 by saponification of its acetylated derivatives 2 and 3. Thus, compound 9 was acetylated to afford hexamannoside 28, which was subjected to hydrogenation over

Scheme 4. Synthesis of Hexamannoses 1–3^a

^aReagents and conditions: (a) TMSOTf, CH₂Cl₂, MS4A, -75 to 0 °C, 6.5 h; 80%; (b) LiAlH₄, Et₂O, -40 to 0 °C, 30 min; 71%; (c) Ac₂O, DMSO, rt, 16 h; NaBH₄, MeOH/CH₂Cl₂ (1:1, v/v), 0 °C to rt, 3 h; 64%; (d) Ac₂O, DMAP, pyr, rt, 3.5–4.0 h; 88% for **28**, 66% for **32**; (e) H₂, Pd/C, MeOH/THF/AcOH (10:20:3, v/v/v), rt, 23 h; 65% for **2**, 74% for **3**; (f) CAN, CH₃CN/toluene/H₂O (18:1:1, v/v/v), 0 °C to rt, 1.5 h; 58%; (g) NaOH, H₂O, rt, 2 h; 84%.

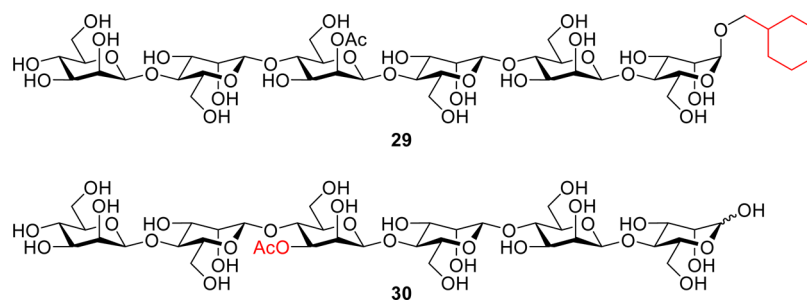


Figure 2. Structures of side products **29** and **30**, which were formed by saturation of the phenyl group and migration of the acetyl group, respectively.

Pd/C in a mixed solvent of MeOH/THF/AcOH. Addition of ~10% acetic acid in the media was found to suppress migration of the acetyl group during the hydrogenation process. The MALDI-TOF MS analysis of the reaction after 23 h showed the desired product of hexasaccharide **2** (m/z 1055.3482 for the $[M + Na]^+$ ion) that was formed by global deprotection of all benzyl, benzylidene and *p*-methoxybenzyl groups. However, the MALDI-TOF MS spectrum also showed a small signal at m/z 1151.4009, which might be from the sodiated molecular ion of mannoside **29** (calcd for C₄₅H₇₆NaO₃₂ 1151.4217) having a cyclohexylmethyl substituent at the anomeric position (Figure 2). The formation of cyclohexylmethyl glycoside on hydrogenation, instead of cleaving the anomeric benzyl group, has been previously reported.²⁹ The side product **29** could be separated from compound **2** by chromatography on a C18 column.

The ¹H NMR analysis (Figure 3) of the crude compound **2**, obtained by removal of the catalyst followed by concentration in vacuo, further revealed the possible migration of the 2-*O*-

acetyl group to the neighboring C-3 position, giving a 3-*O*-acetylated isomer **30** (Figure 2). Figure 3a shows the selected areas of ¹H NMR spectrum of the crude compound **2** recorded immediately after dissolving the crude sample in D₂O. The presence of 2-*O*-acetylated mannoside residue was confirmed by showing the H-2 signal at δ 5.50 as a broad doublet ($J = 3.5$ Hz). The singlet signal at δ 2.17 was attributable to the acetyl group. A trace amount of **30** was observed from a weak signal at δ 2.18 (s) corresponding to its acetyl group. After incubation of the mixture in D₂O at 25 °C for 2 h, new signals appearing at δ 4.18 (d, $J = 3.1$ Hz) and δ 5.10 (dd, $J = 9.8, 3.3$ Hz) were ascribed to H-2 and H-3, respectively, in compound **30**. The protons in **2** and **30** were assigned according to the reported chemical shifts of an *O*-acetylated galactoglucomannan,²⁹ and further confirmed by the HMBC and COSY experiments. To our knowledge, this case represents the first detailed NMR study of intramolecular acetyl migration on an oligomannan, though transesterifications of naturally occurring polymeric mannan and synthetic mannopyranose derivatives have been

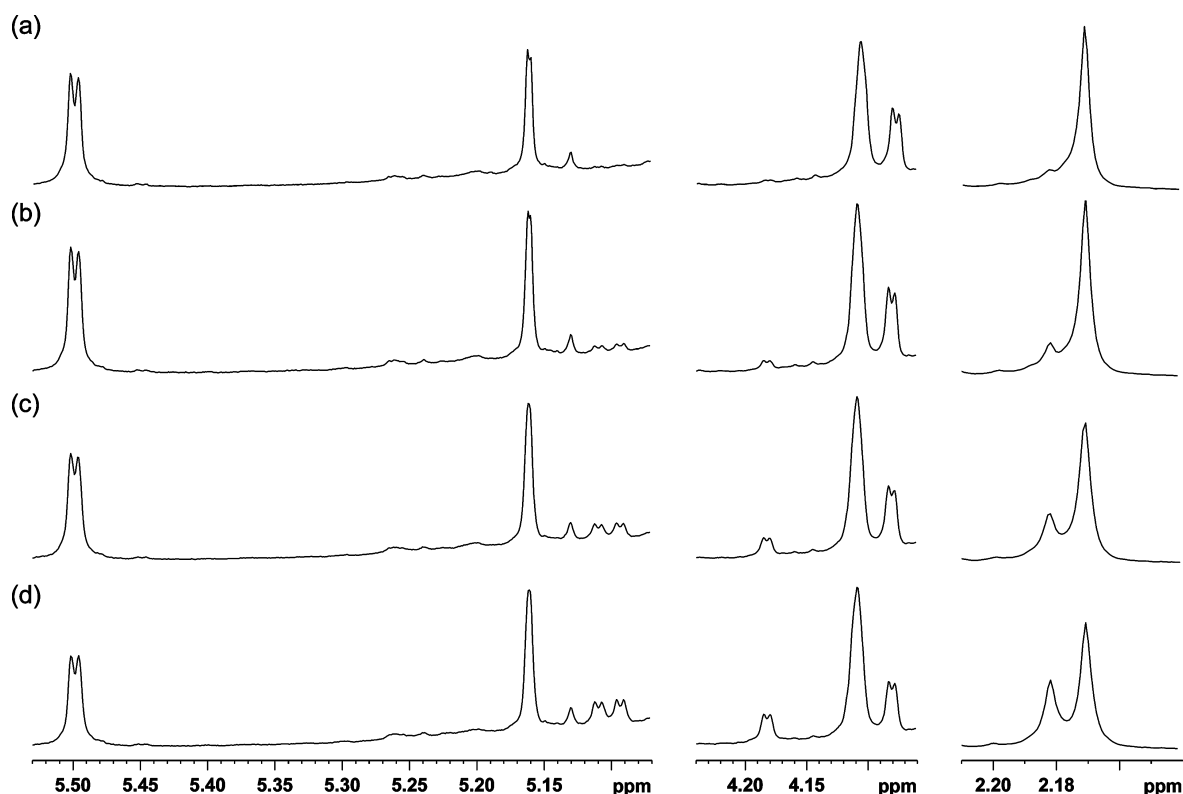
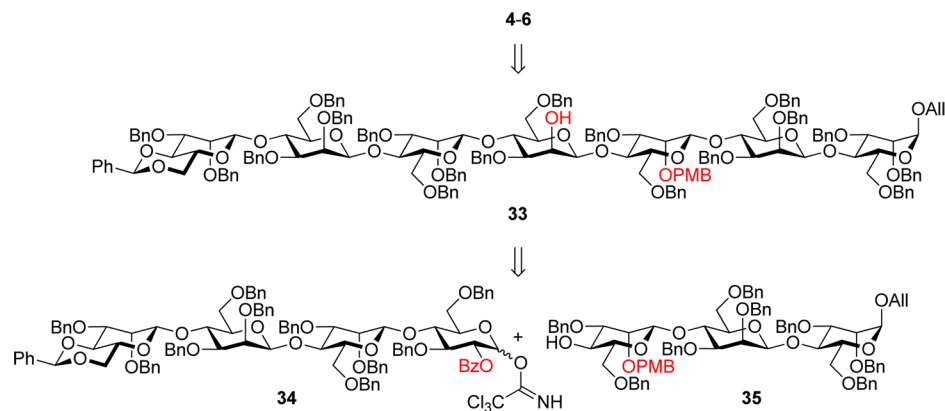


Figure 3. Selected areas of the ^1H NMR spectra of the crude product **2** recorded in D_2O : immediately after dissolving the crude sample in D_2O (a) and after incubation at $25\text{ }^\circ\text{C}$ for 2 h (b), 8 h (c), and 24 h (d). Compound **30** generated by acetyl migration showed signals at δ 2.18 (s), 4.18 (d), and 5.10 (dd).

Scheme 5. Retrosynthetic Analysis of Heptasaccharides 4–6



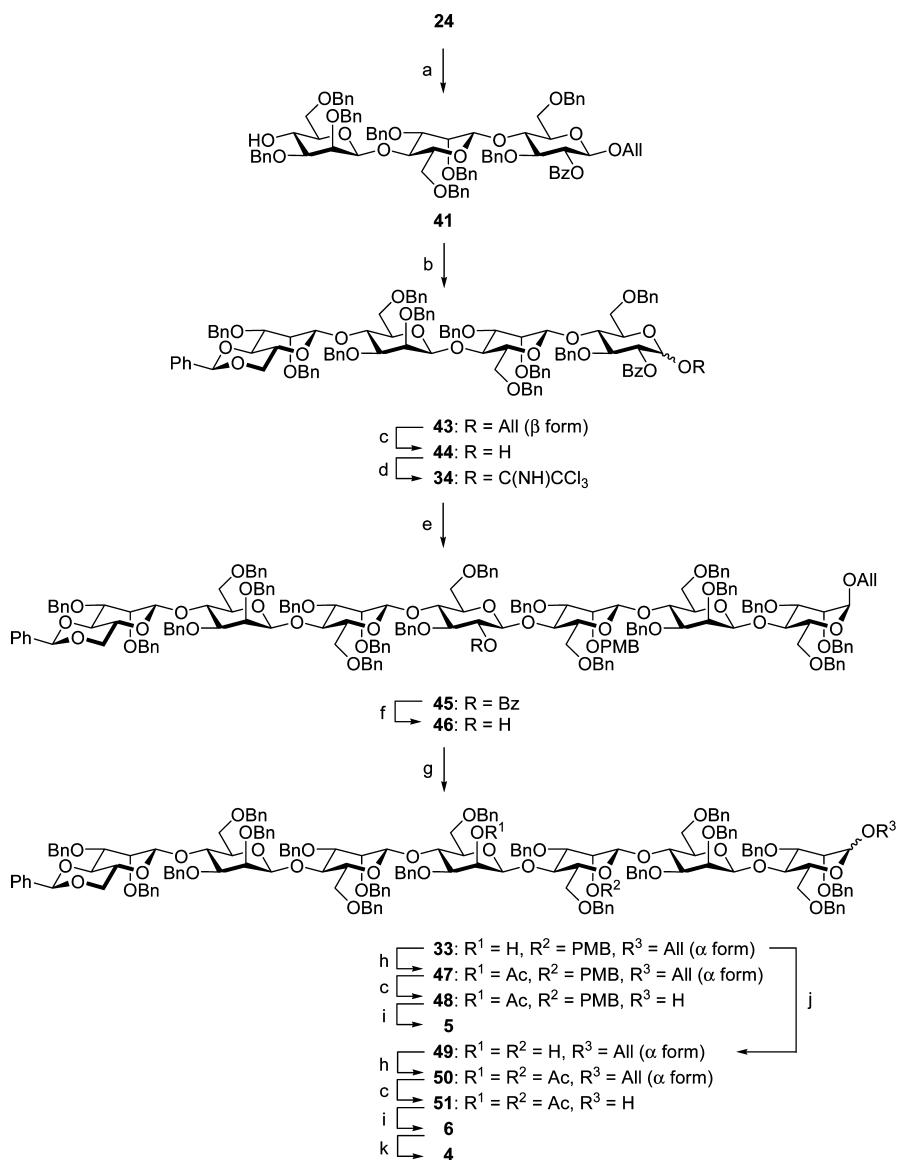
reported.³⁰ Isomerization between **2** and **30** reached an equilibrium ($2/30 \approx 5:3$) after incubation at room temperature for 24 h (Figure 3d). We attempted to suppress the acetyl migration by reversed-phase chromatography under mild acidic conditions (1% AcOH in H_2O); however, the isolated hexamannose **2** was still contaminated with its isomer **30**. In contrast to the previous report,^{30a} there was no acetyl migration from O2 to O6 position in compound **2**.

The synthesis of diacetylated hexasaccharide **3** commenced with removal of the PMB ether group in **9** using CAN in a solution of $\text{CH}_3\text{CN}/\text{PhMe}/\text{H}_2\text{O}$ (18:1:1, v/v/v)³¹ to give alcohol **31**. After acetylation, the diacetylated compound **32** was subjected to hydrogenation to furnish diacetylated hexamannose **3**, which also existed as a mixture of regioisomers due to migration of the acetyl groups. Saponification of **3**

followed by neutralization allowed us to obtain nonacetylated hexamannose **1** in high purity without further purification.

Synthesis of Heptasaccharides 4–6. By a strategy similar to the synthesis of hexasaccharides **1–3**, a [4 + 3] glycosylation of **34** with **35** was applied (Scheme 5). The subsequent epimerization would provide heptasaccharide **33** as the pivotal intermediate leading to heptamannoses **4–6**. The reducing end of trisaccharide acceptor **35** was protected with an allyl group instead of using benzyl group that might form a cyclohexylmethyl glycoside as the side product on hydrogenation. Trisaccharide **35** was prepared by sequential β -selective mannosylations of allyl mannoside **36**¹³ (Scheme 2) in a way similar to that for **11**.

Trisaccharide **41** having a free 4-OH group was designed to synthesize the tetrasaccharide donor **34** via a route incorporat-

Scheme 6. Synthesis of Heptamannoses 4–6^a

^aReagents and conditions: (a) Et₃SiH, TfOH, CH₂Cl₂, MS4A, -78 °C, 15 h; TBAF, CH₂Cl₂, -78 °C to rt, 2 h; 58%; (b) 14, BSP, Tf₂O, TTBP, CH₂Cl₂, MS3A, -78 to -60 °C, 1.5 h; Et₃N, (EtO)₃P, -60 to -10 °C, 15 min; 68%; (c) PdCl₂, MeOH/CH₂Cl₂ (1:1, v/v), rt, 3 h; 79% for 44, 60% for 48, 61% for 51; (d) CCl₃CN, DBU, CH₂Cl₂, 0 °C, 1 h; 56%. (e) 35, TMSOTf, CH₂Cl₂, MS4A, -71 to 0 °C, 3 h; 65%; (f) LiAlH₄, -40 to 0 °C, 30 min; 80%; (g) (i) Ac₂O, DMSO, rt, 16 h; (ii) NaBH₄, MeOH/CH₂Cl₂ (1:1, v/v), 0 °C to rt, 3 h; 60%; (h) Ac₂O, DMAP, pyr, rt, 3 h; 92% for 47, 91% for 50; (i) H₂, Pd/C, THF/MeOH/AcOH (2.0:1.0:0.3, v/v/v), rt, 23 h; H₂, Pd/C, H₂O/MeOH/AcOH (3:6:1, v/v/v), rt, 17 h; 69% for 5, 60% for 6; (j) CAN, CH₃CN/toluene/H₂O (18:1:1, v/v/v), 0 °C to rt, 2.5 h; 75%; (k) NaOH, H₂O, rt, 2 h; 95%.

ing the coupling reaction with mannoside donor 14 under BSP/Tf₂O conditions^{10c} (Scheme 6). Though reductive opening of the benzylidene acetal in trisaccharide 24 using Et₃SiH/BF₃·OEt₂ gave the desired alcohol 41 in 42% crude yield, a considerable amount (26%) of hydrolysis product was also obtained. This problem was not circumvented by addition of molecular sieves. We thus treated 24 with Et₃SiH/TfOH³² at -78 °C in the presence of molecular sieves to yield a major product 41 according to the TLC analysis. Upon workup by addition of Et₃N, this product disappeared along with occurrence of a less polar compound as shown by TLC analysis. This new compound was tentatively assigned to structure 42 (Figure 4) according to the NMR and MALDI-TOF MS analyses (found *m/z* 1505.6879 for [M + Na]⁺, calcd for C₉₀H₁₀₂NaO₁₇Si 1505.6784). We speculated that alcohol 41

might be silylated with the in situ generated TESOTf, a process conceivably promoted by addition of Et₃N base. To avoid forming the silylated compound 42, Et₃N was replaced by a THF solution of TBAF on workup to afford the desired product 41 in 58% yield. Trisaccharide 41 was then coupled with monomer 14 to afford tetrasaccharide 43, which was converted to 34 by deallylation and trichloroacetimidation.

The [4 + 3] coupling reaction of 34 and 35 gave heptasaccharide 45, which was converted to heptamannoside 33 by epimerization of the central glucoside moiety via a sequence comprising removal of the benzoyl group, oxidation, and reduction (Scheme 6). After acetylation, the anomeric allyl group in 47 was removed using PdCl₂, and global reduction of the benzyl and PMB groups by hydrogenation provided monoacetylated heptamannoside 5 (*m/z* 1217.3980 for [M +

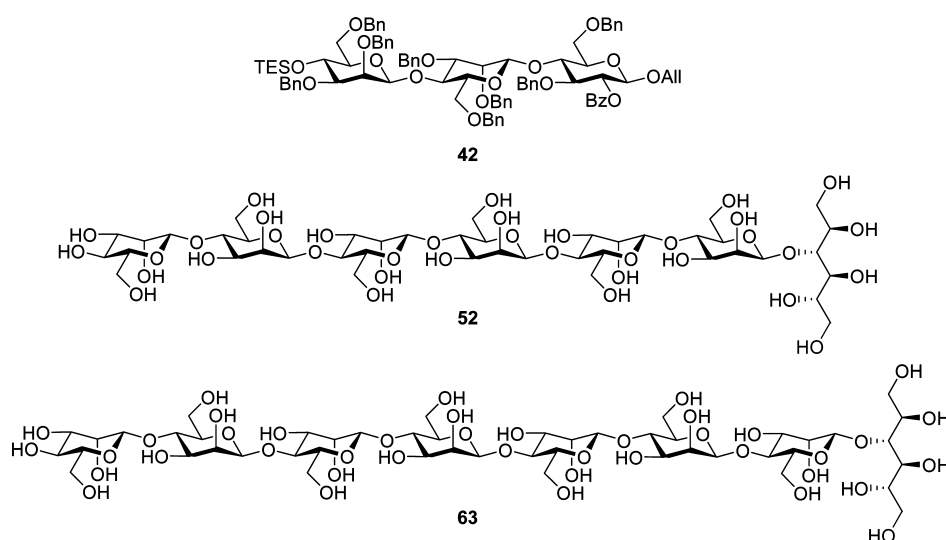
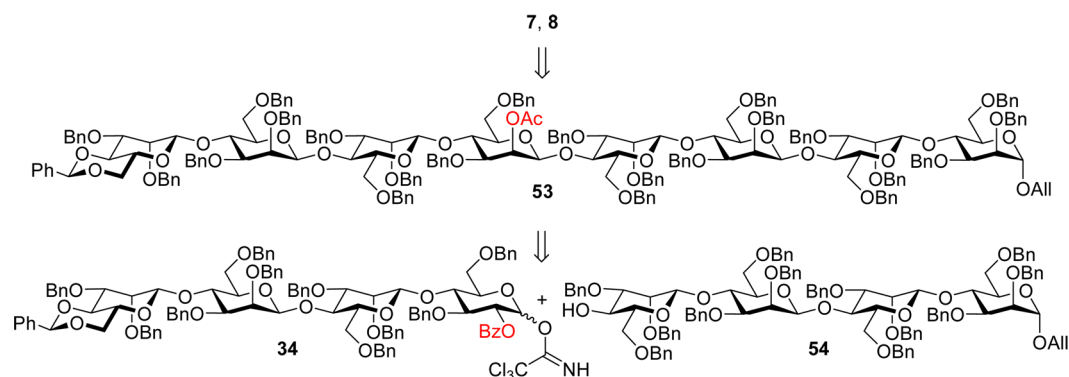


Figure 4. Structures of side products 42, 52, and 63, which were formed by silylation of the hydroxyl group or by over-reduction of the terminal mannose.

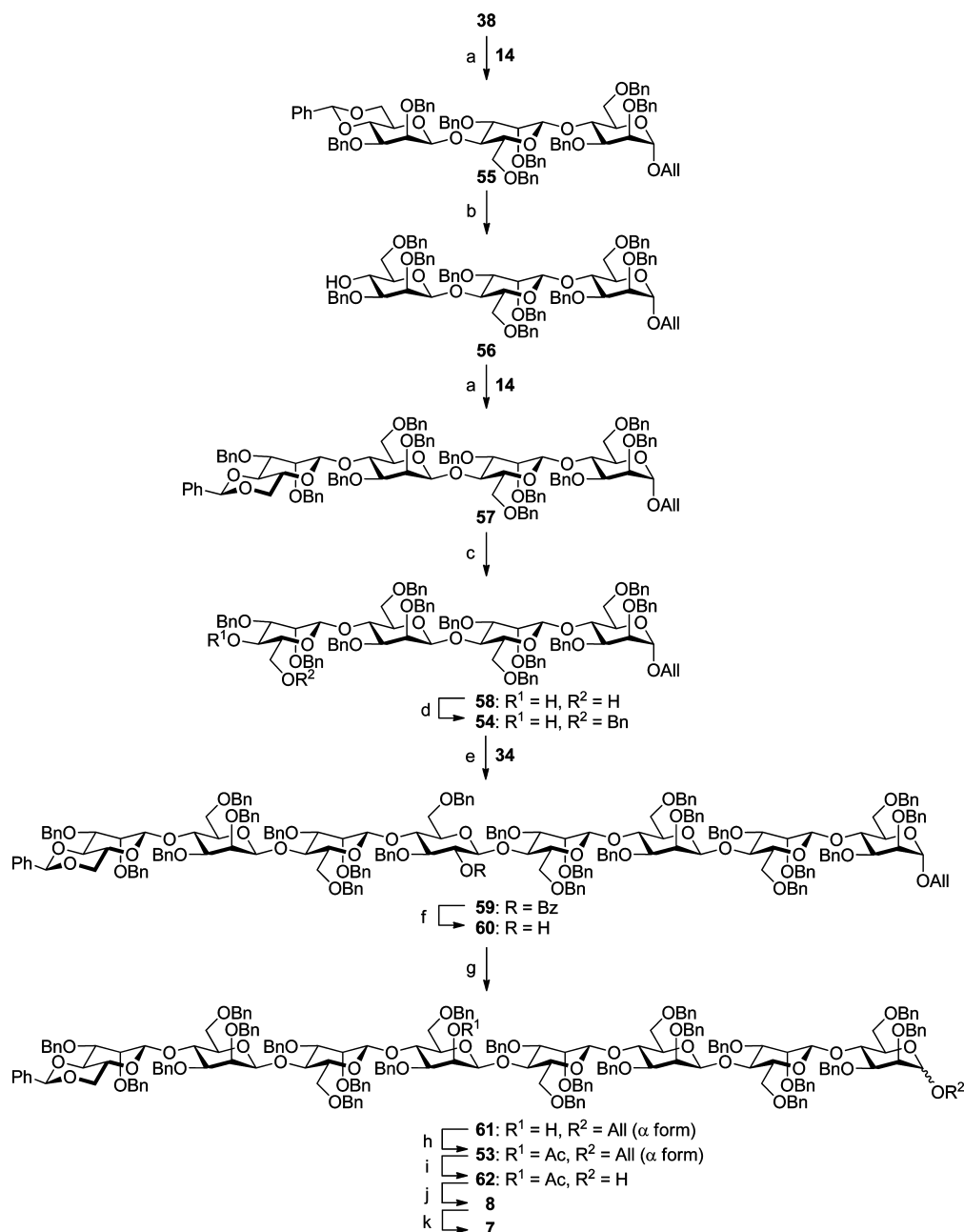
Scheme 7. Retrosynthetic Analysis of Octasaccharides 7 and 8



Na⁺ ion) in 60% yield along with its 3-*O*-acetylated isomer. Heptasaccharide 4 and the diacetylated derivative 6 were similarly prepared from the common intermediate 33. By meticulous examination of the ¹³C NMR and DEPT-135 spectra, we found the product 4 was contaminated with a small amount of 52 bearing a mannitol moiety (Figure 4). Besides the signal at δ_C 69.4 for the C4 at nonreducing end of compound 4, the corresponding signal for the side product 52 occurred at δ_C 65.1 with ~8% intensity. More accurate percentage of the side product was not determined, partly due to overlapping of the characteristic proton signals in the ¹H NMR spectrum. The mannitol moiety in 52 was presumably formed by reduction of the terminal mannose during the process of hydrogenation. The structure of 52 was confirmed by an independent synthesis comprising reduction of the terminal mannose in 51 with NaBH₄, debenzoylation by catalytic hydrogenation, and removal of the acetyl groups by saponification.

Synthesis of Octasaccharides 7 and 8. Scheme 7 shows our retrosynthetic analysis for octamannoses 7 and 8. Starting from glycosylation of disaccharide 38 with mannoside donor 14, the coupling product 55 was then subject to reductive opening of the benzylidene acetal and a second glycosylation with 14 to give tetrasaccharide 57 (Scheme 8). Reduction of the benzylidene acetal in 57 was found inefficient using the combined reagents of Et₃SiH/TfOH/TBAF. We thus con-

verted 57 to alcohol 54 in two steps comprising trans-acetalization with neopentyl glycol^{9,21} and monobenylation via the stannylene acetal.²² Octasaccharide 60 was obtained by assembly of oligosaccharide fragments 34 and 54, followed by removal of the benzoyl group. However, epimerization of the central glucoside moiety in 60 turned out to be problematic. Though oxidation of 60 proceeded smoothly to afford the corresponding ketone as judged by the NMR analysis, the subsequent reduction with NaBH₄ did not give the desired axial alcohol in a reproducible manner. This problem was finally overcome by using L-Selectride³³ as the reducing agent to obtain compound 61 in all-manno configuration. After acetylation, the allyl, benzyl, and benzylidene groups were removed to give 2-*O*-acetylated octamannose 8, which also coexisted with its 3-*O*-acetylated isomer. Saponification of 8 yielded octamannose 7. A small amount (~8%) of the heptamannose–mannitol compound 63 (Figure 4) was also detected in the ¹³C NMR and DEPT-135 spectra, similar to that observed in 52. The structure of 63 was confirmed by an independent synthesis in a five-step sequence: (i) palladium-catalyzed deallylation, (ii) reduction of the terminal mannose by NaBH₄, (iii) acetylation, (iv) debenzoylation by catalytic hydrogenation, and (v) removal of the acetyl group by saponification. Interestingly, the monoacetylated derivative 8 is more soluble in water than its parental octamannose 7; even these compounds differ only in one acetyl group. This

Scheme 8. Synthesis of Octasaccharides 7 and 8^a

^aReagents and conditions: (a) 14, BSP, Tf₂O, TTBP, CH₂Cl₂, MS3A, -78 to -60 °C, 1.5 h; Et₃N, (EtO)₃P, -60 °C to rt, 20 min; 68% for 55, 65% for 57; (b) Et₃SiH, TfOH, CH₂Cl₂, MS4A, -78 °C, 15 h; TBAF -78 °C to rt, 2 h; 44%; (c) CSA, neopentyl glycol, CH₂Cl₂, rt, 24 h; 78%; (d) Bu₂SnO, toluene, reflux, 3 h; BnBr, CsF, toluene, reflux, 6 h; 81%; (e) 34, TMSOTf, CH₂Cl₂, MS4A, -70 to 0 °C, 4 h; 78%; (f) LiAlH₄, -40 to 0 °C, 30 min; 67%; (g) Ac₂O, DMSO, rt, 16 h; L-Selectride, THF, -78 °C to rt, 1.5 h; 58%; (h) Ac₂O, DMAP, pyr, rt, 3 h; 91%; (i) PdCl₂, MeOH/CH₂Cl₂ (1:1, v/v), rt, 3 h; 59%; (j) H₂, Pd/C, THF/MeOH/AcOH (2:1:0.3, v/v/v), rt, 23 h; H₂, Pd/C, H₂O/MeOH/AcOH (3:6:1, v/v/v), rt, 17 h; 63%; (k) NaOH, H₂O, rt, 1.5 h; 85%.

phenomenon implies that acetylation of β -(1 \rightarrow 4)-linked oligomannosides in varied degrees may cause the disturbance of intra- and intermolecular hydrogen bond interactions.

In summary, we have synthesized β -(1 \rightarrow 4)-linked hexa-, hepta-, and octamannoses as well as their mono- and diacetylated derivatives. Case by case, various reaction reagents and conditions were tuned to optimize the yields of the desired products and to minimize the side reactions. The efficiency of our strategy relied on the combined use of direct and indirect mannosylation methods to construct β -mannoside backbone

and to install acetyl groups at late stage of the synthetic pathways. The acetyl group migrated reversibly from O2 to O3 positions in the central mannose residues, but no acetyl migration from O2 to O6 was observed. The oligomannose compounds 1–8 existed as anomers of hemiacetal forms, of which ratio could not be ambiguously determined because the α/β anomeric proton/carbon resonances either overlapped or interfered with HOD in solvent.

Biological Activity. The synthesized β -(1 \rightarrow 4)-linked hexa-, hepta-, and octamannoses as well as their mono- and

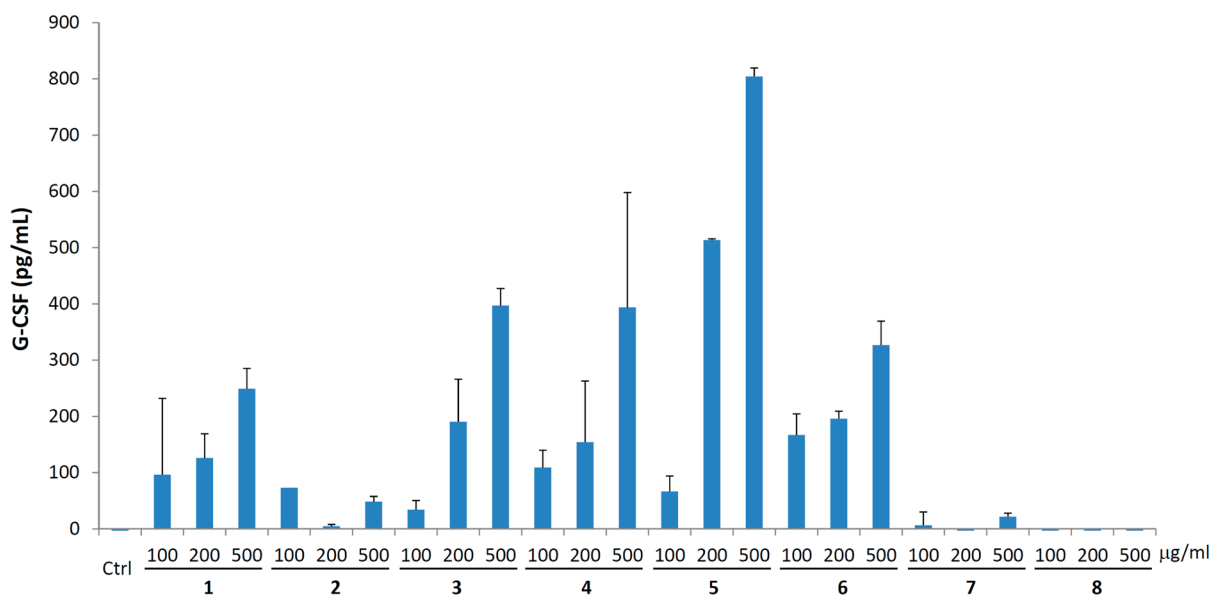


Figure 5. Induced expression of G-CSF by oligomannoses. RAW264.7 cells were treated with oligomannoses 1–8 at indicated concentrations for 6 h, and the media were collected for ELISA analysis. Ctrl: nontreated cells.

diacetylated derivatives were evaluated for their activities in inducing G-CSF expression. The oligosaccharide was dissolved in a solution buffered at neutral pH and the dissolved material was directly used for activity evaluation with an incubation time of 6 h. The possible acetyl isomerization of samples during incubation, presumably less than 25% in the case of compound 2 according to the results shown in Figure 3b (for 2 h incubation in D₂O) and Figure 3c (for 8 h incubation), was not rigorously determined. It was shown that both hexa- and heptamannoses (1 and 4, respectively) can significantly induce G-CSF expression (Figure 5). Surprisingly, octamannose (7) did not significantly induce G-CSF expression. It was also unexpected to observe that the expression of G-CSF induced by oligomannose is similar to the level induced by their acetylated derivatives. We also tested the bioactivity of oligomannose–mannitol compounds 52 and 63, but no G-CSF induction was found (data not shown). More systematic studies on the location as well as the degree of acetylation may be needed for further investigation.

EXPERIMENTAL SECTION

General Methods. All chemicals were of commercial grade and were used without further purification. Anhydrous solvents were purchased and used without additional purification. Thin-layer chromatography (TLC) was performed on silica gel 60 F254 glass plate. Detection was effected by exposure to UV light (254 nm) and/or by charring with sulfuric acid (5% in ethanol solution). Column chromatography was performed on silica gel 60 (40–60 µm). Melting points were determined with a melting point apparatus and were uncorrected. All reactions were carried out under an argon atmosphere. ¹H and ¹³C NMR spectra were recorded on a 600 MHz spectrometer at 25 °C. In ¹H NMR spectra, chemical shifts were referenced to internal tetramethylsilane (δ 0.00 ppm in CDCl₃), C₆D₅H (δ 7.16 ppm in C₆D₆), or HOD (δ 4.79 ppm in D₂O or in CD₃CO₂D/D₂O (1:99, v/v)). In ¹³C NMR spectra, chemical shifts were referenced to the carbon signals in CDCl₃ (δ 77.00 ppm) and C₆D₆ (δ 128.39 ppm). In ¹³C NMR spectra acquired in D₂O and in CD₃CO₂D/D₂O (1:99, v/v), chemical shifts were corrected to 4,4-dimethyl-4-silapentane-1-sulfonic acid (DSS) as a reference. Coupling constants are given in hertz. Signal splitting patterns are described as singlet (s), doublet (d), triplet (t), double of doublet (dd), double of

triplet (dt), triple of doublet (td), double of doublet doublet (ddd), broad signal (br), and multiplet (m). Mass spectra (MS) were recorded on an ESI-TOF mass spectrometer or a MALDI-TOF/TOF mass spectrometer using 2,5-dihydroxybenzoic acid (DHB) or 6-aza-2-thiothymine (ATT) as matrices. High-resolution mass spectra (HRMS) were recorded on an ESI-FTICR mass spectrometer.

Cell Treatment and ELISA Analysis. The mouse macrophage cell line RAW264.7 (ATCC TIB71) was maintained at 37 °C with 5% CO₂ in supplemented RPMI 1640 (10% fetal bovine serum, 0.2 mM L-glutamine, 100 U of penicillin/mL, and 100 µg of streptomycin/mL). RAW cells were plated at 6 × 10⁵ cells in wells of 24-well plates (Greiner Bio-One) in supplemented RPMI 1640 one day prior to treatment. The cells were treated with synthetic oligomannoses at indicated concentrations for 6 h. Following the treatment, the medium were collected for G-CSF quantitation using the Quantikine mouse ELISA kits (R&D Systems) according to the manufacturer's protocol. Shown data are a representative data set from three independent experiments.

Benzyl 2,3,6-Tri-O-benzyl-4-O-(2,3-di-O-benzyl-4,6-O-benzylidene-β-D-mannopyranosyl)-α-D-mannopyranoside (16). Representative Procedure for β-Selective Mannosylation. To a stirred solution containing compound 14 (948 mg, 1.75 mmol, 1.25 equiv), 1-benzenesulfonylpiperidine (BSP) (404 mg, 1.93 mmol, 1.4 equiv), tritert-butylpyrimidine (TTBP) (871 mg, 3.51 mmol, 2.5 equiv) and activated 3 Å powdered molecular sieves (5 g) in CH₂Cl₂ (70 mL) at –60 °C under an argon atmosphere was added Tf₂O (350 µL, 2.08 mmol, 1.5 equiv). After 5 min, a solution of benzyl 2,3,6-tri-O-benzyl-α-D-mannopyranoside 13 (0.758 g, 1.40 mmol, 1.0 equiv) in CH₂Cl₂ (8 mL) was added. The mixture was cooled to –78 °C, stirred for 6 h, warmed to room temperature, and filtered. The filtrate washed with saturated aqueous NaHCO₃ and brine, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was subjected to column chromatography on silica gel (hexane/EtOAc, 88:12) to give compound 16 (994 mg, 1.02 mmol, 73%): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 3.06 (td, J = 9.6, 4.7 Hz, 1H), 3.40 (dd, J = 10.0, 3.0 Hz, 1H), 3.54–3.65 (m, 3H), 3.69 (d, J = 2.8 Hz, 1H), 3.75 (dt, J = 9.5, 3.2 Hz, 1H), 3.79 (t, J = 2.7 Hz, 1H), 3.93 (dd, J = 8.4, 3.1 Hz, 1H), 3.99 (dd, J = 10.4, 4.8 Hz, 1H), 4.06 (t, J = 9.6 Hz, 1H), 4.22 (t, J = 9.0 Hz, 1H), 4.40 (d, J = 12.1, Hz, 1H), 4.46–4.59 (d overlapped s, 2H), 4.53–4.59 (2 overlapping d, 2H), 4.62–4.86 (m, 8H), 4.96 (d, J = 2.1 Hz, 1H), 5.50 (s, 1H), 7.15–7.41 (m, integration not determined), 7.44–7.48 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz) δ 67.3, 68.5, 69.1, 71.4, 72.45, 72.49, 72.8, 73.4, 74.9, 75.6, 76.2, 77.0, 78.0, 78.4, 78.7, 97.6, 101.3, 102.0, 126.1, 127.2, 127.3, 127.4, 127.5,

127.7, 127.75, 127.80, 128.0, 128.1, 128.17, 128.20, 128.22, 128.3, 128.37, 128.44, 128.8, 137.4, 137.7, 138.2, 138.3, 138.6, 138.7, 139.1; HRMS (ESI) found m/z 993.4164 $[M + Na]^+$, calcd for $C_{61}H_{62}NaO_{11}$ 993.4190.

Benzyl 2,3,6-Tri-O-benzyl-4-O-(2,3,6-tri-O-benzyl- β -D-mannopyranosyl)- α -D-mannopyranoside (17). *Representative Procedure for Reductive Ring-Opening of Benzylidene Acetal.* To a stirred solution of compound 16 (837 mg, 0.86 mmol, 1.0 equiv) in CH_2Cl_2 (8.7 mL) at 0 °C were added Et_3SiH (1.7 mL, 10.5 mmol, 12.2 equiv) and $BF_3 \cdot OEt_2$ (0.17 mL, 1.34 mmol, 1.6 equiv). After the solution was stirred for 9 h at 0 °C, saturated aqueous $NaHCO_3$ was added. The organic layer was separated, dried over Na_2SO_4 , and concentrated in vacuo. Chromatography of the residue on a silica gel column (hexane/EtOAc, 85:15) gave compound 17 (523 mg, 0.54 mmol, 62%): colorless oil; 1H NMR ($CDCl_3$, 600 MHz) δ 2.69 (br s, 1H), 3.14 (dd, $J = 9.3, 2.7$ Hz, 1H), 3.21 (dt, $J = 9.6, 4.9$ Hz, 1H), 3.56 (dd, $J = 10.4, 5.4$ Hz, 1H), 3.59–3.66 (2 overlapping dd, 2H), 3.67–3.73 (dd overlapped with d, 2H), 3.78 (t, $J = 2.8$ Hz, 1H), 3.83 (ddd, $J = 9.5, 4.3, 2.5$ Hz, 1H), 3.93 (t, $J = 9.5$ Hz, 1H), 3.97 (dd, $J = 8.3, 2.9$ Hz, 1H), 4.26 (t, $J = 8.9$ Hz, 1H), 4.37–4.50 (m, 6H), 4.51 (s, 1H), 4.56 (d, $J = 12.0$ Hz, 1H), 4.61–4.74 (m, 5H), 4.78 (d, $J = 12.1$ Hz, 1H), 4.83 (d, $J = 12.0$ Hz, 1H), 4.95 (d, $J = 2.3$ Hz, 1H), 7.17–7.46 (m, integration not determined); ^{13}C NMR ($CDCl_3$, 151 MHz) δ 68.8, 69.1, 69.4, 71.1, 71.46, 71.51, 72.5, 72.7, 73.4, 73.7, 74.2, 74.6, 75.6, 75.7, 78.1, 81.8, 97.7, 101.4, 127.2, 127.3, 127.39, 127.44, 127.5, 127.57, 127.60, 127.70, 127.73, 127.8, 128.05, 128.13, 128.2, 128.3, 128.37, 128.41, 128.42, 137.5, 138.0, 138.1, 138.3, 138.4, 138.9, 139.2; MS (ESI) found m/z 995.4 $[M + Na]^+$, calcd for $C_{61}H_{64}NaO_{11}$ 995.4.

Benzyl 3-O-Benzyl-4,6-O-benzylidene-2-O-p-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (18). According to the representative procedure for β -selective mannosylation, the reaction compound of 17 (254 mg, 0.26 mmol) and phenyl 2-O-p-methoxybenzyl-3-O-benzyl-4,6-O-benzylidene-1-deoxy-1-thio- α -D-mannopyranoside (15, 182 mg, 0.32 mmol) was performed, followed by quench with Et_3N (220 μ L, 1.58 mmol) and $(EtO)_3P$ (109 μ L, 0.64 mmol), to give compound 18 (244 mg, 0.17 mmol, 65%) after chromatography on silica gel (hexane/EtOAc, 80:20): colorless oil; 1H NMR ($CDCl_3$, 600 MHz) δ 3.04 (td, $J = 9.7, 4.8$ Hz, 1H), 3.15–3.25 (m, 1H), 3.31 (dd, $J = 9.2, 2.8$ Hz, 1H), 3.38 (dd, $J = 9.8, 2.9$ Hz, 1H), 3.64–3.74 (m, 4H), 3.75–3.84 (m, 5H), 3.92–3.99 (m, 2H), 4.02 (t, $J = 9.6$ Hz, 1H), 4.14 (t, $J = 9.4$ Hz, 1H), 4.21–4.29 (t overlapped with d, 2H), 4.40–4.51 (m, 5H), 4.53 (2 overlapping s, 2H), 4.58 (d, $J = 12.4$ Hz, 1H), 4.61–4.79 (m, 10H), 4.87 (d, $J = 12.4$ Hz, 1H), 4.95 (d, $J = 2.1$ Hz, 1H), 5.47 (s, 1H), 6.78–6.84 (m, 2H), 7.07–7.40 (m, integration not determined), 7.41–7.49 (m, 2H); ^{13}C NMR ($CDCl_3$, 151 MHz, selected signals) δ 55.2, 67.2, 68.5, 69.0, 69.29, 69.34, 71.5, 72.2, 72.3, 72.6, 72.7, 73.4, 73.5, 74.2, 74.6, 75.5, 75.7, 75.9, 76.1, 76.7, 78.35, 78.38, 78.6, 80.7, 97.6, 101.3, 101.5, 102.2, 113.5, 126.1, 126.9, 127.1, 127.2, 127.2, 127.35, 127.42, 127.5, 127.59, 127.64, 127.8, 128.0, 128.11, 128.14, 128.2, 128.3, 128.4, 128.7, 137.4, 137.7, 138.3, 138.36, 138.37, 138.6, 139.05, 139.10, 139.3, 159.1; MS (MALDI, DHB) found m/z 1455.6 $[M + Na]^+$, calcd for $C_{89}H_{92}NaO_{17}$: 1455.6.

Benzyl 3-O-Benzyl-2-O-p-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (19). *Representative Procedure for Transacetalation.* To a stirred solution of compound 18 (562 mg, 0.39 mmol, 1.0 equiv) and neopentyl glycol (122 mg, 1.17 mmol, 3.0 equiv) in CH_2Cl_2 (3.9 mL) was added camphorsulfonic acid (18 mg, 0.08 mmol, 0.2 equiv). After being stirred for 1 day at room temperature, saturated aqueous $NaHCO_3$ was added. The organic layer was separated, washed with water, dried over Na_2SO_4 , and concentrated in vacuo. Chromatography of the residue on a silica gel column (hexane/EtOAc, 55:45) gave compound 19 (388 mg, 0.29 mmol, 74%): colorless oil; 1H NMR ($CDCl_3$, 600 MHz) δ 3.01–3.14 (m, 2H), 3.18–3.29 (m, 2H), 3.40 (dd, $J = 11.8, 6.7$ Hz, 1H), 3.53–3.58 (2 overlapping dd, 2H), 3.62–3.70 (m, 3H), 3.71–3.84 (m, 8H), 3.96 (dd, $J = 8.7, 3.0$ Hz, 1H), 4.11–4.17 (m, 2H), 4.25–4.34 (m, 3H), 4.41 (d, $J = 12.1$ Hz, 1H), 4.44–4.53 (m, 5H), 4.59 (d, $J = 12.3$ Hz, 1H), 4.61–4.80 (m, 9H), 4.90 (d, $J = 12.5$

Hz, 1H), 4.95 (d, $J = 2.0$ Hz, 1H), 6.75–6.84 (m, 2H), 7.14–7.41 (m, integration not determined); ^{13}C NMR ($CDCl_3$, 151 MHz, selected signals) δ 55.3, 62.8, 67.3, 69.1, 69.3, 69.5, 70.9, 71.6, 72.6, 72.68, 72.73, 73.2, 73.4, 73.6, 73.7, 74.4, 75.46, 75.51, 75.56, 75.62, 75.8, 76.5, 78.3, 80.3, 82.0, 97.7, 101.37, 101.39, 113.6, 127.1, 127.2, 127.26, 127.28, 127.4, 127.47, 127.52, 127.6, 127.72, 127.74, 127.77, 127.83, 127.88, 127.89, 128.06, 128.14, 128.2, 128.3, 128.35, 128.43, 128.5, 129.6, 130.7, 137.4, 137.8, 138.32, 138.33, 138.4, 138.8, 138.9, 139.3, 159.13; MS (MALDI, DHB) found m/z 1367.5951 $[M + Na]^+$, calcd for $C_{82}H_{88}NaO_{17}$ 1367.5919.

Benzyl 3,6-Di-O-benzyl-2-O-p-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (11). *Representative Procedure for Regioselective Benzylation.* To a stirred solution of diol 19 (367 mg, 0.27 mmol, 1.0 equiv) in toluene (8.0 mL) was added Bu_2SnO (136 mg, 0.55 mmol, 2.0 equiv) at room temperature. The mixture was stirred under reflux for 3 h and cooled to room temperature. CsF (191 mg, 1.26 mmol, 4.7 equiv) and $BnBr$ (130 μ L, 1.09 mmol, 4.0 equiv) were added to the above-prepared stannylene acetal. The mixture was stirred under reflux for another 6 h. After the mixture was cooled to 0 °C, saturated aqueous $NaHCO_3$ was added, and the mixture was diluted with EtOAc. The organic phase was separated, washed with water and brine, dried over Na_2SO_4 , and concentrated in vacuo. The crude product was purified by column chromatography on silica gel (hexane/EtOAc, 80:20) to afford compound 11 (263 mg, 0.18 mmol, 67%): colorless oil; 1H NMR ($CDCl_3$, 600 MHz) δ 2.67 (br s, 1H), 3.12 (dd, $J = 9.4, 2.9$ Hz, 1H), 3.19 (dt, $J = 9.6, 4.8$ Hz, 1H), 3.23–3.28 (m, 1H), 3.34 (dd, $J = 9.3, 2.8$ Hz, 1H), 3.50–3.56 (m, 2H), 3.56–3.66 (m, 4H), 3.69 (d, $J = 2.7$ Hz, 1H), 3.73 (d, $J = 2.7$ Hz, 1H), 3.75 (s, 3H), 3.77–3.83 (m, 2H), 3.89 (t, $J = 9.5$ Hz, 1H), 3.96 (dd, $J = 8.4, 3.1$ Hz, 1H), 4.17 (t, $J = 9.3$ Hz, 1H), 4.22 (t, $J = 9.1$ Hz, 1H), 4.27–4.33 (2 overlapping d, 2H), 4.35–4.54 (m, 9H), 4.57 (d, $J = 12.2$ Hz, 1H), 4.59–4.75 (m, 8H), 4.77 (d, $J = 12.3$ Hz, 1H), 4.87 (d, $J = 12.4$ Hz, 1H), 4.94 (d, $J = 2.3$ Hz, 1H), 6.73–6.80 (m, 2H), 7.08–7.40 (m, integration not determined); ^{13}C NMR ($CDCl_3$, 151 MHz, selected signals) δ 55.2, 68.7, 69.0, 69.4, 69.5, 71.1, 71.3, 71.5, 72.3, 72.6, 72.7, 73.3, 73.5, 73.7, 73.8, 74.0, 74.1, 74.6, 75.3, 75.5, 75.7, 75.8, 76.0, 78.3, 80.7, 81.8, 97.7, 101.5, 101.6, 113.5, 127.1, 127.2, 127.3, 127.4, 127.46, 127.51, 127.56, 127.64, 127.71, 127.74, 127.8, 127.9, 128.1, 128.2, 128.25, 128.33, 128.37, 128.41, 129.5, 131.1, 137.5, 138.12, 138.14, 138.3, 138.4, 138.5, 139.1, 139.2, 139.3, 159.0; MS (MALDI, DHB) found m/z 1457.6 $[M + Na]^+$, calcd for $C_{89}H_{94}NaO_{17}$ 1457.6.

Allyl 2-O-Benzoyl-3-O-benzyl-4,6-O-benzylidene- β -D-glucopyranoside (21). Allyl 3-O-benzyl-4,6-O-benzylidene- β -D-glucopyranoside (20, 21.3 g, 53.5 mmol, 1.0 equiv) was dissolved in pyridine (200 mL), and benzoyl chloride (9.4 mL, 80.9 mmol, 1.5 equiv) was added at 0 °C. The mixture was stirred at room temperature for 10 h and quenched with water at 0 °C. After being stirred for another 2 h, the mixture was diluted with EtOAc and washed successively with saturated aqueous $NaHCO_3$, water and brine. The organic layer was dried over Na_2SO_4 and concentrated in vacuo. The residue was purified by silica gel column chromatography (hexane/EtOAc, 70:30) to give compound 21 (26.7 g, 53.1 mmol, 99%): white solid; mp 118–119 °C; 1H NMR (C_6D_6 , 600 MHz) δ 3.21 (td, $J = 9.7, 4.9$ Hz, 1H), 3.49 (t, $J = 10.2$ Hz, 1H), 3.57 (t, $J = 9.4$ Hz, 1H), 3.78–3.89 (m, 2H), 4.08–4.23 (m, 2H), 4.43 (d, $J = 8.0$ Hz, 1H), 4.77 (d, $J = 12.4$ Hz, 1H), 4.82–4.93 (m, 2H), 5.10–5.17 (m, 1H), 5.22 (s, 1H), 5.56–5.65 (m, 1H), 5.77 (t, $J = 8.6$ Hz, 1H), 6.96–7.02 (m, 3H), 7.07 (t, $J = 7.5$ Hz, 2H), 7.12–7.18 (m, integration not determined), 7.20–7.27 (m, 4H), 7.63 (d, $J = 7.8$ Hz, 2H), 8.16 (d, $J = 7.8$ Hz, 2H); ^{13}C NMR (C_6D_6 , 151 MHz) δ 66.9, 69.1, 70.1, 74.1, 74.2, 78.6, 82.3, 101.4, 101.7, 117.1, 126.9, 128.0, 128.7, 128.8, 128.8, 128.9, 129.4, 130.5, 131.2, 133.4, 134.5, 138.7, 139.2, 165.4; HRMS (ESI) found m/z 525.1869 $[M + Na]^+$, calcd for $C_{30}H_{30}NaO_7$ 525.1889.

Allyl 2-O-Benzoyl-3,6-di-O-benzyl- β -D-glucopyranoside (12). According to the representative procedure for reductive ring-opening of benzylidene acetal, compound 21 (2.69 g, 5.35 mmol) was treated with Et_3SiH (4.3 mL, 26.6 mmol) and CF_3CO_2H (2.0 mL, 26.9 mmol) at 0 °C for 1 h to give compound 12 (2.38 g, 4.72 mmol, 88%) after

chromatography on a silica gel column (hexane/EtOAc, 70:30): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 2.84 (d, $J = 2.5$ Hz, 1H), 3.54 (dt, $J = 9.5, 4.8$ Hz, 1H), 3.67 (t, $J = 9.1$ Hz, 1H), 3.74–3.83 (m, 3H), 4.00–4.10 (m, 1H), 4.25–4.33 (m, 1H), 4.56–4.60 (2 overlapping d, 2H), 4.63 (d, $J = 12.1$ Hz, 1H), 4.68 (d, $J = 11.6$ Hz, 1H), 4.72 (d, $J = 11.6$ Hz, 1H), 5.02–5.10 (m, 1H), 5.16–5.22 (m, 1H), 5.28 (t, $J = 8.7$ Hz, 1H), 5.70–5.80 (m, 1H), 7.12–7.22 (m, 5H), 7.27–7.32 (m, 1H), 7.33–7.38 (m, 4H), 7.44 (t, $J = 7.7$ Hz, 2H), 7.53–7.60 (m, 1H), 8.00–8.06 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 69.6, 70.3, 72.1, 73.3, 73.7, 74.1, 74.3, 82.1, 99.9, 117.2, 127.7, 127.8, 128.0, 128.3, 128.4, 129.7, 129.9, 133.0, 133.6, 137.7, 137.9, 165.2; MS (ESI) found m/z 527.2 [$\text{M} + \text{Na}$] $^+$, calcd for $\text{C}_{30}\text{H}_{32}\text{NaO}_7$ 527.2; HRMS found m/z 527.2046 [$\text{M} + \text{Na}$] $^+$, calcd for $\text{C}_{30}\text{H}_{32}\text{NaO}_7$ 527.2052.

Allyl 2-O-Benzoyl-3,6-di-O-benzyl-4-O-(2,3-di-O-benzyl-4,6-O-benzylidene- β -D-mannopyranosyl)- β -D-glucopyranoside (22). According to the representative procedure for β -selective mannosylation, the reaction of compounds 12 (3.01 g, 5.97 mmol) and 14 (4.06 g, 5.51 mmol) was performed, followed by quench with Et_3N and $(\text{EtO})_3\text{P}$, to give compound 22 (3.98 g, 4.26 mmol, 71%) after chromatography on silica gel (hexane/EtOAc, 90:10): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 3.10 (td, $J = 9.7, 4.8$ Hz, 1H), 3.41 (dd, $J = 9.9, 3.0$ Hz, 1H), 3.43–3.48 (m, 1H), 3.52 (t, $J = 10.3$ Hz, 1H), 3.59 (dd, $J = 11.0, 3.8$ Hz, 1H), 3.68 (dd, $J = 2.3, 11.0$ Hz, 1H), 3.72–3.78 (m, 2H), 4.00–4.12 (m, 4H), 4.28–4.35 (m, 1H), 4.43 (d, $J = 12.1$ Hz, 1H), 4.54 (s, 1H), 4.55–4.62 (m, 3H), 4.65 (d, $J = 12.1$ Hz, 1H), 4.73 (d, $J = 12.4$ Hz, 1H), 4.84 (d, $J = 12.0$ Hz, 1H), 4.88 (d, $J = 12.0$ Hz, 1H), 4.92 (d, $J = 11.5$ Hz, 1H), 5.07–5.11 (m, 1H), 5.16–5.23 (m, 1H), 5.26 (dd, $J = 8.1, 9.0$ Hz, 1H), 5.50 (s, 1H), 5.69–5.83 (m, 1H), 7.03–7.13 (m, 5H), 7.20–7.49 (m, integration not determined), 7.51–7.58 (m, 1H), 7.96–8.02 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 67.3, 68.5, 68.6, 69.5, 72.6, 73.2, 73.6, 74.4, 74.98, 75.02, 78.1, 78.3, 78.7, 80.6, 99.9, 101.3, 101.8, 117.2, 126.1, 127.2, 127.4, 127.51, 127.53, 127.7, 127.8, 128.0, 128.1, 128.2, 128.26, 128.32, 128.5, 128.8, 129.8, 130.1, 132.9, 133.8, 137.6, 137.8, 138.5, 138.6, 165.1; MS (ESI) found m/z 957.5 [$\text{M} + \text{Na}$] $^+$, calcd for $\text{C}_{57}\text{H}_{58}\text{NaO}_{12}$ 957.4.

Allyl 2-O-Benzoyl-3,6-di-O-benzyl-4-O-(2,3,6-tri-O-benzyl- β -D-mannopyranosyl)- β -D-glucopyranoside (23). According to the representative procedure for reductive ring-opening of benzylidene acetal, compound 22 (862 mg, 0.92 mmol) was treated with Et_3SiH (1.8 mL, 11.1 mmol) and $\text{BF}_3 \cdot \text{OEt}_2$ (187 μL , 1.48 mmol) at 0 $^\circ\text{C}$ for 1.5 h to give compound 23 (637 mg, 0.68 mmol, 74%) after chromatography on a silica gel column (hexane/EtOAc, 70:30): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 2.78 (br s, 1H), 3.16 (dd, $J = 9.5, 3.0$ Hz, 1H), 3.26 (dt, $J = 9.7, 4.9$ Hz, 1H), 3.51–3.57 (m, 2H), 3.61 (dd, $J = 10.2, 4.9$ Hz, 1H), 3.66 (dd, $J = 11.0, 4.1$ Hz, 1H), 3.72 (dd, $J = 10.9, 2.6$ Hz, 1H), 3.75 (d, $J = 2.9$ Hz, 1H), 3.80 (t, $J = 8.9$ Hz, 1H), 3.95 (t, $J = 9.5$ Hz, 1H), 4.02–4.10 (m, 2H), 4.31 (ddt, $J = 13.4, 4.7, 1.6$ Hz, 1H), 4.35–4.51 (m, 5H), 4.54 (s, 1H), 4.59 (d, $J = 7.9$ Hz, 1H), 4.60–4.68 (m, 2H), 4.77 (d, $J = 11.9$ Hz, 1H), 4.85 (d, $J = 11.9$ Hz, 1H), 4.93 (d, $J = 11.7$ Hz, 1H), 5.06–5.10 (m, 1H), 5.17–5.23 (m, 1H), 5.28 (dd, $J = 9.0, 8.0$ Hz, 1H), 5.76 (dddd, $J = 17.1, 10.7, 6.2, 4.7$ Hz, 1H), 6.96–7.00 (m, 2H), 7.01–7.08 (m, 3H), 7.20–7.35 (m, integration not determined), 7.37–7.43 (m, 4H), 7.51–7.58 (m, 1H), 7.91–8.04 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 68.8, 68.9, 69.5, 71.1, 71.6, 73.1, 73.6, 73.6, 74.0, 74.2, 74.5, 74.6, 75.0, 77.4, 80.4, 81.7, 99.8, 101.1, 117.2, 127.0, 127.4, 127.5, 127.6, 127.60, 127.67, 127.73, 127.75, 127.82, 127.9, 128.1, 128.2, 128.3, 128.4, 128.5, 129.8, 130.0, 132.9, 133.8, 137.86, 137.93, 138.1, 138.6, 138.7, 165.6; MS (ESI) found m/z 959.4 [$\text{M} + \text{Na}$] $^+$, calcd for $\text{C}_{57}\text{H}_{60}\text{NaO}_{12}$ 959.4.

Allyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-benzoyl-3,6-di-O-benzyl- β -D-glucopyranoside (24). According to the representative procedure for β -selective mannosylation, the reaction of compounds 23 (2.83 g, 3.02 mmol) and 14 (2.04 g, 3.77 mmol) was performed, followed by quench with Et_3N and $(\text{EtO})_3\text{P}$, to give compound 24 (2.19 g, 1.60 mmol, 53%) after chromatography on silica gel (toluene/EtOAc, 95:5): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 3.06 (td, $J = 9.7, 4.8$ Hz, 1H), 3.18–3.25 (m, 1H), 3.30 (dd, $J = 9.2, 2.9$ Hz, 1H), 3.40 (dd, $J = 9.8, 3.0$ Hz, 1H), 3.45–3.52 (m, 2H), 3.52–3.59 (m, 2H), 3.63 (dd, $J = 11.1, 3.9$ Hz, 1H), 3.70–3.75 (m, 3H), 3.77 (t, $J = 9.0$ Hz, 1H), 3.98 (dd, $J = 4.8, 10.4$ Hz, 1H), 4.03–4.10 (m, 3H), 4.17 (t, $J = 9.3$ Hz, 1H), 4.23 (d, $J = 11.8$ Hz, 1H), 4.27–4.34 (m, 1H), 4.41–4.49 (m, 3H), 4.50–4.59 (m, 4H), 4.61–4.69 (m, 3H), 4.74–4.83 (m, 4H), 4.85 (d, $J = 12.0$ Hz, 1H), 4.98 (d, $J = 12.1$ Hz, 1H), 5.05–5.09 (m, 1H), 5.16–5.22 (m, 1H), 5.29 (t, $J = 8.6$ Hz, 1H), 5.72–5.82 (m, 1H), 6.92–6.96 (m, 1H), 6.98–7.02 (m, 1H), 7.04 (d, $J = 7.5$ Hz, 1H), 7.13–7.42 (m), 7.43–7.47 (m, 2H), 7.51–7.57 (m, 1H), 7.96 (d, $J = 8.1$ Hz, 1H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 67.2, 68.5, 68.7, 69.3, 69.5, 72.4, 73.1, 73.4, 73.5, 74.1, 74.2, 74.9, 75.0, 75.4, 76.0, 76.1, 77.1, 77.8, 78.4, 78.6, 80.55, 80.61, 99.9, 101.2, 101.3, 102.2, 117.2, 126.1, 126.9, 127.0, 127.2, 127.3, 127.35, 127.43, 127.46, 127.53, 127.66, 127.67, 127.8, 127.9, 128.0, 128.08, 128.10, 128.13, 128.2, 128.26, 128.29, 128.5, 128.8, 129.8, 130.1, 132.9, 133.8, 137.7, 137.9, 138.2, 138.5, 138.7, 138.9, 139.1, 165.2; MS (ESI) found m/z 1389.7 [$\text{M} + \text{Na}$] $^+$, calcd for $\text{C}_{84}\text{H}_{86}\text{NaO}_{17}$ 1389.6.

2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-benzoyl-3,6-di-O-benzyl- β -D-glucopyranoside (25). Representative Procedure for Removal of Allyl Protecting Group. To a solution of compound 24 (220 mg, 0.16 mmol, 1.0 equiv) in $\text{MeOH}/\text{CH}_2\text{Cl}_2$ (1:1, v/v, 2.6 mL) was added a suspension of PdCl_2 (6 mg, 0.03 mmol, 0.2 equiv) in CH_2Cl_2 . After being stirred at room temperature for 6 h, additional PdCl_2 (18 mg, 0.10 mmol, 0.6 equiv) was added. The mixture was stirred for another 2 h and then diluted with EtOAc. The mixture was filtered through a small pad of silica gel. The filtrate was washed with saturated aqueous NaHCO_3 , dried over MgSO_4 , and concentrated in vacuo. The residue was purified by column chromatography on silica gel (hexane/EtOAc, 70:30) to give compound 25 (164 mg, 0.12 mmol, 77%): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 3.07 (td, $J = 4.9, 9.7$ Hz, integration not determined), 3.20–3.26 (m, integration not determined), 3.26–3.31 (m, integration not determined), 3.40 (dd, $J = 3.0, 9.8$ Hz, integration not determined), 3.44–3.52 (m, integration not determined), 3.52–3.66 (m, integration not determined), 3.81 (t, $J = 9.1$ Hz, integration not determined), 3.95–4.21 (m, integration not determined), 4.23 (d, $J = 12.0$ Hz, integration not determined), 4.39–4.57 (m, integration not determined), 4.59–4.73 (m, integration not determined), 4.75–4.87 (m, integration not determined), 5.03–5.15 (m, integration not determined), 5.49 (s, integration not determined), 5.55 (d, $J = 3.6$ Hz, integration not determined), 6.93–7.49 (m, integration not determined), 7.51–7.58 (m, integration not determined), 7.90–8.01 (m, integration not determined); ^{13}C NMR (CDCl_3 , 151 MHz, selected signals) δ 67.3, 68.49, 68.54, 68.7, 69.3, 69.4, 70.1, 72.4, 72.5, 73.4, 73.5, 73.6, 73.7, 74.2, 74.3, 74.5, 74.8, 74.9, 75.36, 75.40, 75.9, 76.06, 76.10, 76.13, 76.2, 77.5, 77.6, 77.8, 78.4, 78.6, 80.2, 80.50, 80.52, 90.5, 96.1, 101.1, 101.2, 101.3, 102.2, 126.1, 127.0, 127.1, 127.2, 127.3, 127.36, 127.43, 127.47, 127.51, 127.6, 127.65, 127.68, 127.8, 127.90, 127.94, 128.0, 128.05, 128.09, 128.12, 128.2, 128.28, 128.30, 128.4, 128.51, 128.53, 128.8, 129.3, 129.6, 129.8, 129.9, 133.2, 133.4, 137.66, 137.69, 138.2, 138.3, 138.5, 138.66, 138.72, 138.9, 139.1, 165.9, 167.1; MS (MALDI, DHB) found m/z 1349.5 [$\text{M} + \text{Na}$] $^+$, calcd for $\text{C}_{81}\text{H}_{82}\text{NaO}_{17}$ 1349.5.

2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-benzoyl-3,6-di-O-benzyl- α/β -D-glucopyranosyl Trichloroacetimidate (10). Representative Procedure for Trichloroacetimidation. To a solution of compound 25 (517 mg, 0.39 mmol, 1.0 equiv) and trichloroacetonitrile (391 μL , 3.90 mmol, 10 equiv) in CH_2Cl_2 (6.7 mL) was added a solution of DBU (5.8 μL , 0.04 mmol, 0.1 equiv) in CH_2Cl_2 (0.1 mL) at 0 $^\circ\text{C}$. After being stirred for 1 h, the mixture was concentrated under reduced pressure. Chromatography of the residue on a silica gel column (hexane/EtOAc/ Et_3N , 70:30:1) gave compound 10 (411 mg, 0.28 mmol, 72%): colorless oil; ^1H NMR (C_6D_6 , 600 MHz) δ 3.03–3.14 (m, integration not determined), 3.23–3.42 (m, integration not determined), 3.45–3.52 (m, integration not determined), 3.53–3.71 (m, integration not determined), 3.73–3.82 (m, integration not determined), 3.87 (d, $J = 2.8$ Hz, integration not determined), 3.96 (t, $J = 8.7$ Hz, integration not determined), 4.00–

4.07 (m, integration not determined), 4.16–4.35 (m, integration not determined), 4.36–4.43 (m, integration not determined), 4.43–4.65 (m, integration not determined), 4.71 (s, integration not determined), 4.75–4.82 (m, integration not determined), 4.90–5.15 (m, integration not determined), 5.21 (s, integration not determined), 5.22 (s, integration not determined), 5.33 (d, $J = 12.3$ Hz, integration not determined), 5.44 (d, $J = 12.4$ Hz, integration not determined), 5.66–5.75 (m), 6.02 (t, $J = 8.4$ Hz), 6.27 (d, $J = 7.9$ Hz), 6.92–7.30 (m), 7.32–7.40 (m), 7.44–7.52 (m, integration not determined), 7.54–7.62 (m, integration not determined), 8.08–8.18 (m, integration not determined), 8.37 (s, integration not determined), 8.58 (s, integration not determined); ^{13}C NMR (C_6D_6 , 151 MHz, selected signals) δ 68.1, 69.0, 69.1, 69.2, 69.86, 69.92, 72.7, 72.9, 73.3, 73.5, 73.6, 74.0, 74.1, 74.18, 74.22, 74.4, 74.7, 75.3, 75.39, 75.43, 76.1, 76.5, 76.7, 76.77, 76.83, 77.0, 77.6, 77.8, 78.3, 78.9, 79.6, 79.7, 81.0, 81.3, 81.6, 91.5, 92.0, 95.0, 97.3, 101.7, 101.8, 102.0, 102.7, 127.0, 127.59, 127.64, 127.8, 127.9, 129.2, 130.4, 130.5, 130.6, 133.4, 133.5, 138.70, 138.73, 139.1, 139.17, 139.24, 139.8, 139.90, 139.94, 140.05, 140.09, 140.5, 161.0, 162.2, 165.5, 166.0; MS (MALDI, ATT) found m/z 1492.5 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{83}\text{H}_{88}\text{Cl}_3\text{NNaO}_{17}$ 1492.5.

Benzyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-benzoyl-3,6-di-O-benzyl- β -D-glucopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl-2-O-p-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (26). *Representative Procedure for Glycosylation.* Acceptor 11 (72 mg, 50.2 μmol , 1.0 equiv) and trichloroacetimidate 10 (111 mg, 75.4 μmol , 1.5 equiv) were combined and coevaporated three times with toluene. The mixture was dissolved in CH_2Cl_2 (1.5 mL), and molecular sieves 4 Å (160 mg) were added. The mixture was stirred for 30 min and cooled to -75 °C, and a solution of TMSOTf (1.35 μL , 7.5 μmol , 0.15 equiv) in CH_2Cl_2 (10 μL) was added. The mixture was stirred for 3.5 h, during which the temperature increased to 0 °C. After being stirred at 0 °C for another 3 h, the reaction was quenched by addition of Et_3N (5.2 μL , 37.4 μmol , 0.75 equiv). Saturated aqueous NaHCO_3 was added. The mixture was diluted with CH_2Cl_2 and washed with water. The organic layer was dried over MgSO_4 and concentrated in vacuo. The residue was purified by column chromatography on silica gel (hexane/EtOAc, 80:20) to give compound 26 (111 mg, 40.4 μmol , 80%): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 2.86–2.93 (m, 1H), 3.04 (td, $J = 9.6$, 4.9 Hz, 1H), 3.14–3.22 (m, 4H), 3.24–3.31 (m, 3H), 3.37–3.51 (m, 7H), 3.52–3.63 (m, 5H), 3.63–3.67 (2 overlapping d, 2H), 3.68–3.74 (m, 5H), 3.74–3.81 (m, 2H), 3.91–4.10 (m, 6H), 4.11–4.23 (m, 5H), 4.28 (d, $J = 12.2$ Hz, 1H), 4.33 (s, 1H), 4.36–4.85 (m, 33H), 4.92 (d, $J = 2.3$ Hz, 1H), 4.99 (d, $J = 12.2$ Hz, 1H), 5.19 (dd, $J = 9.4$, 8.3 Hz, 1H), 5.48 (s, 1H), 6.61–6.72 (m, 2H), 6.84–6.92 (m, 2H), 6.93–7.00 (m, 3H), 7.02–7.42 (m), 7.42–7.48 (m, 2H), 7.51–7.59 (m, 1H), 7.78–7.97 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 55.2, 67.2, 68.3, 68.5, 69.0, 69.26, 69.32, 69.34, 71.5, 72.1, 72.29, 72.31, 72.4, 72.6, 72.7, 73.27, 73.29, 73.33, 73.4, 73.5, 73.7, 74.0, 74.16, 74.23, 74.3, 74.9, 75.1, 75.3, 75.4, 75.5, 75.66, 75.69, 75.8, 76.0, 76.1, 77.1, 78.1, 78.2, 78.5, 78.6, 80.46, 80.52, 80.6, 80.9, 97.6, 100.5, 101.27, 101.33, 101.40, 102.2, 113.4, 126.1, 126.8, 126.9, 126.97, 126.99, 127.02, 127.06, 127.09, 127.2, 127.27, 127.30, 127.35, 127.41, 127.44, 127.47, 127.52, 127.60, 127.63, 127.64, 127.7, 127.8, 127.9, 128.0, 128.05, 128.07, 128.11, 128.17, 128.20, 128.26, 128.29, 128.34, 128.4, 128.8, 129.2, 129.8, 129.9, 131.3, 132.9, 137.4, 137.7, 138.1, 138.3, 138.4, 138.5, 138.7, 138.8, 138.9, 139.0, 139.09, 139.11, 139.2, 139.4, 158.8, 165.0; MS (MALDI, DHB) found m/z 2766.0 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{170}\text{H}_{174}\text{NaO}_{33}$ 2766.2.

Benzyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl- β -D-glucopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl-2-O-p-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (27). *Representative Procedure for Removal of the Benzoyl Group.* To a solution of compound 26 (110 mg, 40.1 μmol , 1.0 equiv) in Et_2O (4.2 mL) was added a suspension of LiAlH_4 (6.1 mg, 160.1 μmol , 4.0 equiv) in Et_2O (320 μL) at -40 °C. After being stirred at -40 °C for 30 min, the mixture was warmed to 0 °C.

Saturated aqueous NaHCO_3 was added. The mixture was diluted with EtOAc and washed with water. The organic layer was dried over MgSO_4 and concentrated in vacuo. Chromatography of the residue on a silica gel column (hexane/EtOAc, 85:15) gave compound 27 (75 mg, 28.4 μmol , 71%): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 3.03 (td, $J = 9.6$, 4.9 Hz, 1H), 3.08–3.16 (m, 2H), 3.18–3.24 (m, 4H), 3.27 (dd, $J = 11.1$, 3.2 Hz, 1H), 3.30–3.43 (m, 5H), 3.44–3.76 (m, 17H), 3.77–3.83 (m, 3H), 3.91–3.98 (m, 2H), 4.02 (t, $J = 9.6$ Hz, 1H), 4.12 (t, $J = 9.3$ Hz, 1H), 4.14–4.26 (m, 5H), 4.28 (d, $J = 11.9$ Hz, 1H), 4.32–4.42 (m, 5H), 4.43–4.54 (m, 12H), 4.57 (d, $J = 12.3$ Hz, 1H), 4.60–4.80 (m, 16H), 4.86 (d, $J = 12.3$ Hz, 1H), 4.94 (d, $J = 2.1$ Hz, 1H), 4.99 (d, $J = 11.9$ Hz, 1H), 5.47 (s, 1H), 6.69–6.74 (m, 2H), 7.11–7.41 (m, integration not determined), 7.42–7.45 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 55.2, 67.2, 68.5, 68.6, 69.0, 69.3, 69.4, 69.6, 71.2, 71.5, 72.2, 72.3, 72.4, 72.6, 72.7, 73.25, 73.34, 73.39, 73.41, 73.6, 73.8, 74.1, 74.2, 74.6, 74.7, 74.8, 74.9, 75.0, 75.19, 75.23, 75.45, 75.48, 75.7, 75.8, 76.0, 76.1, 77.07, 77.10, 78.2, 78.4, 78.6, 80.6, 80.7, 81.8, 82.5, 97.6, 101.1, 101.3, 101.4, 101.9, 102.2, 103.7, 113.5, 126.1, 126.6, 126.8, 126.9, 127.0, 127.1, 127.15, 127.19, 127.24, 127.3, 127.38, 127.41, 127.43, 127.46, 127.50, 127.56, 127.60, 127.63, 127.7, 128.0, 128.07, 128.10, 128.13, 128.16, 128.21, 128.25, 128.29, 128.32, 128.4, 128.8, 129.3, 131.0, 137.4, 137.7, 137.9, 138.0, 138.29, 138.31, 138.35, 138.42, 138.5, 138.7, 138.97, 139.03, 139.1, 139.2, 139.6, 158.9; MS (MALDI, DHB) found m/z 2662.2 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{163}\text{H}_{170}\text{NaO}_{32}$ 2662.2.

Benzyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl-2-O-p-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (9). *Representative Procedure for Gluco to Manno Epimerization.* A solution of compound 27 (66 mg, 25 μmol , 1 equiv) in DMSO (0.82 mL) and acetic anhydride (0.42 mL) was stirred for 16 h at room temperature. After the solution was cooled to 0 °C, saturated aqueous NaHCO_3 was added. The mixture was diluted with Et_2O , washed with saturated aqueous NaHCO_3 , water, and brine, dried over Na_2SO_4 , filtered, and concentrated in vacuo. The residue was coevaporated with toluene three times and dried in vacuo. The residue was dissolved in CH_2Cl_2 (1.0 mL) and MeOH (1.0 mL) and cooled to 0 °C. NaBH_4 (12 mg, 317 μmol , 13 equiv) was added in one portion. The mixture was stirred for 3 h, during which time the solution was allowed to warm to room temperature. Water was added. The mixture was diluted with CH_2Cl_2 and washed with water. The organic layer was dried over MgSO_4 and concentrated in vacuo. The residue was purified by column chromatography on silica gel (hexane/EtOAc, 65:35) to give compound 9 (42 mg, 15.9 μmol , 64%): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 2.70 (br s, 1H), 3.03 (td, $J = 9.6$, 4.8 Hz, 1H), 3.12–3.29 (m, 5H), 3.29–3.41 (m, 4H), 3.43–3.58 (m, 8H), 3.59–3.76 (m, 10H), 3.76–3.82 (m, 2H), 3.90 (br d, 1H), 3.91–3.98 (m, 2H), 4.00–4.09 (2 overlapping d, 2H), 4.11–4.19 (3 overlapping t, 3H), 4.20–4.42 (m, 8H), 4.44–4.54 (m, 13H), 4.57 (d, $J = 12.3$ Hz, 1H), 4.59–4.80 (m, 17H), 4.87 (d, $J = 12.3$ Hz, 1H), 4.94 (d, $J = 2.1$ Hz, 1H), 5.48 (s, 1H), 6.71–6.76 (m, 2H), 7.09–7.39 (m, integration not determined), 7.41–7.47 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 55.2, 67.2, 68.1, 68.5, 69.0, 69.1, 69.31, 69.34, 69.4, 69.5, 71.5, 71.6, 71.8, 72.2, 72.3, 72.4, 72.6, 72.7, 73.2, 73.3, 73.37, 73.43, 73.7, 74.0, 74.1, 74.7, 74.87, 74.90, 75.0, 75.16, 75.23, 75.51, 75.53, 75.7, 75.8, 75.86, 75.93, 76.1, 77.1, 78.2, 78.5, 78.6, 79.5, 80.7, 81.4, 97.6 ($^1J_{\text{CH}} = 168.5$ Hz), 100.2 ($^1J_{\text{CH}} = 158.8$ Hz), 101.3, 101.4 ($^1J_{\text{CH}} = 159.2$ Hz), 101.6 ($^1J_{\text{CH}} = 155.7$ Hz), 101.8 ($^1J_{\text{CH}} = 155.1$ Hz), 102.1 ($^1J_{\text{CH}} = 156.4$ Hz), 113.5, 126.1, 126.9, 127.1, 127.15, 127.24, 127.25, 127.30, 127.35, 127.39, 127.41, 127.44, 127.46, 127.50, 127.56, 127.62, 127.68, 127.73, 127.8, 127.96, 127.98, 128.06, 128.10, 128.13, 128.16, 128.18, 128.21, 128.25, 128.32, 128.4, 128.8, 129.3, 131.1, 137.4, 137.7, 138.30, 138.32, 138.36, 138.42, 138.5, 138.71, 138.74, 139.03, 139.05, 139.07, 139.14, 139.3, 158.9; MS (MALDI, DHB) found m/z 2662.0 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{163}\text{H}_{170}\text{NaO}_{32}$ 2662.2.

Benzyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl-3,6-di-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl-2-O-p-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-

2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (28). *Representative Procedure for Acetylation.* To a solution of compound 9 (42 mg, 15.9 μ mol, 1 equiv) and DMAP (5 mg, 40.9 μ mol, 3 equiv) in pyridine (1.8 mL) was added acetic anhydride (0.1 mL). The mixture was stirred for 3.5 h at room temperature and then quenched by addition of MeOH at 0 °C. After the mixture was stirred for 10 min, saturated NaHCO₃ was added. The mixture was diluted with EtOAc, washed with saturated aqueous NaHCO₃ and water, dried over MgSO₄, filtrated, and concentrated in vacuo. Chromatography of the residue on a silica gel column (hexane/EtOAc, 70:30) gave compound 28 (38 mg, 14.2 μ mol, 88%): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 2.01 (s, 3H), 3.03 (td, *J* = 9.7, 4.9 Hz, 1H), 3.12–3.19 (m, 3H), 3.19–3.35 (m, 5H), 3.37–3.43 (m, 3H), 3.43–3.74 (m, 16H), 3.76–3.83 (m, 2H), 3.90–3.99 (m, 3H), 4.02 (t, *J* = 9.6 Hz, 1H), 4.07 (t, *J* = 9.4 Hz, 1H), 4.12–4.24 (m, 4H), 4.25–4.32 (2 overlapping d, 2H), 4.32–4.42 (m, 4H), 4.43–4.82 (m, 31H), 4.86 (d, *J* = 12.3 Hz, 1H), 4.94 (d, *J* = 1.8 Hz, 1H), 5.39 (d, *J* = 2.9 Hz, 1H), 5.47 (s, 1H), 6.65–6.72 (m, 2H), 7.09–7.41 (m, integration not determined), 7.41–7.48 (m, 2H); ¹³C NMR (CDCl₃, 600 MHz) δ 21.0, 55.2, 67.2, 68.5, 68.8, 68.95, 69.0, 69.1, 69.36, 69.42, 71.4, 71.5, 71.8, 72.1, 72.3, 72.4, 72.6, 72.7, 73.1, 73.3, 73.39, 73.42, 73.7, 74.1, 74.2, 74.6, 74.90, 74.94, 75.0, 75.2, 75.30, 75.34, 75.5, 75.7, 75.8, 75.9, 76.1, 78.20, 78.23, 78.4, 78.6, 80.6, 80.7, 81.3, 97.7, 99.8, 101.2, 101.3, 101.4, 101.7, 102.2, 113.4, 126.1, 126.77, 126.84, 127.0, 127.10, 127.14, 127.2, 127.26, 127.32, 127.4, 127.45, 127.50, 127.6, 127.66, 127.73, 127.8, 127.96, 128.00, 128.03, 128.08, 128.12, 128.14, 128.18, 128.20, 128.25, 128.32, 128.4, 128.8, 129.3, 131.2, 137.4, 137.7, 138.3, 138.36, 138.42, 138.46, 138.52, 138.76, 138.79, 139.05, 139.14, 139.2, 139.3, 158.8, 170.3; MS (MALDI, DHB) found *m/z* 2704.2 [M + Na]⁺, calcd for C₁₆₅H₁₇₂NaO₃₃ 2704.2.

β -D-Mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranose (2). *Representative Procedure for Global Deprotection of Benzyl, Benzylidene, and p-Methoxybenzyl Groups.* A mixture of compound 28 (8 mg, 2.98 μ mol) and 10% Pd/C (8 mg) in THF/MeOH/AcOH (2:1:0.3, v/v/v, 1.0 mL) was stirred under an atmosphere of hydrogen at room temperature for 23 h. The mixture was passed through a pad of Celite to remove the catalyst and then concentrated under reduced pressure. The residue was subject to reversed-phase chromatography on a C18 column (1% AcOH in H₂O), followed by freezing drying, and gave compound 2 (2.0 mg, 1.94 μ mol, 65%): white hygroscopic solid; ¹H NMR (1% CD₃CO₂D in D₂O, 600 MHz) δ 2.19 (s, integration not determined), 2.20 (s, integration not determined), 3.41–3.60 (m, integration not determined), 3.61–3.68 (m, integration not determined), 3.70–4.08 (m, integration not determined), 4.09–4.15 (m, integration not determined), 4.20 (d, *J* = 3.3 Hz, integration not determined), 4.72 (s, integration not determined), 4.73 (s, integration not determined), 4.76 (s, integration not determined), 4.83 (s, integration not determined), 4.91 (s, integration not determined), 4.95 (s, integration not determined), 5.12 (dd, *J* = 9.8, 3.1 Hz, integration not determined), 5.18 (s, integration not determined), 5.52 (d, *J* = 3.5 Hz, integration not determined); ¹³C NMR (1% CD₃CO₂D in D₂O, 151 MHz, selected signals) δ 63.2, 63.4, 63.7, 69.4, 71.4, 71.7, 72.67, 72.70, 72.8, 72.85, 72.94, 73.20, 73.23, 73.4, 73.6, 74.1, 74.17, 74.21, 74.27, 74.29, 74.4, 75.48, 75.49, 76.0, 76.1, 77.51, 77.54, 77.71, 77.74, 77.9, 78.0, 79.12, 79.14, 79.19, 79.22, 79.29, 79.31, 79.5, 80.1, 96.4, 96.6, 101.8, 102.3, 102.8, 102.87, 102.92, 102.93, 103.0, 175.7, 176.1; MS found *m/z* 1055.3, calcd for C₃₈H₆₄NaO₃₂ 1055.3; HRMS found *m/z* 1033.3425 [M + H]⁺, calcd for C₃₈H₆₅O₃₂ 1033.3459.

Benzyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (31). *Representative Procedure for Selective Deprotection of p-Methoxybenzyl Group.* Compound 9 (12 mg, 4.54 μ mol, 1.0 equiv) was dissolved in a mixture of CH₃CN/toluene/H₂O (90:5:5, v/v/v, 1.0 mL) and cooled to 0 °C. CAN (12 mg, 21.9 mmol, 4.8 equiv) was added in one portion. The mixture was stirred at 0 °C for 30 min and at room temperature for another 1 h. The mixture was diluted with EtOAc and

quenched with saturated aqueous NaHCO₃. The organic layer was washed with saturated aqueous NaHCO₃ and water, dried over MgSO₄, and concentrated in vacuo. The residue was purified by column chromatography on silica gel (hexane/EtOAc, 62:38) to give compound 31 (7 mg, 2.78 μ mol, 58%): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 2.60 (br s, 1H), 2.78 (br s, 1H), 3.03 (td, *J* = 9.7, 4.9 Hz, 1H), 3.15–3.21 (m, 3H), 3.21–3.28 (m, 2H), 3.30 (dd, *J* = 9.1, 3.2 Hz, 1H), 3.35–3.41 (m, 3H), 3.47–3.59 (m, 7H), 3.61–3.69 (m, 5H), 3.72 (d, *J* = 2.9 Hz, 1H), 3.73 (d, *J* = 2.8 Hz, 1H), 3.78 (t, *J* = 2.7 Hz, 1H), 3.79–3.83 (m, 1H), 3.89 (d, *J* = 2.7 Hz, 1H), 3.92–3.98 (m, 3H), 4.03 (t, *J* = 9.6 Hz, 1H), 4.05–4.10 (2 overlapping t, 2H), 4.12–4.20 (2 overlapping t, 2H), 4.21–4.30 (m, 3H), 4.32–4.37 (2 overlapping d, 2H), 4.39 (d, *J* = 11.8 Hz, 1H), 4.41–4.54 (m, 15H), 4.57 (d, *J* = 12.3 Hz, 1H), 4.60–4.80 (m, 15H), 4.83 (d, *J* = 12.3 Hz, 1H), 4.94 (d, *J* = 2.2 Hz, 1H), 5.48 (s, 1H), 7.11–7.39 (m, integration not determined), 7.42–7.46 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz, selected signals) δ 67.2, 67.4, 68.1, 68.5, 69.0, 69.06, 69.14, 69.3, 69.40, 69.42, 71.2, 71.5, 71.8, 71.9, 72.2, 72.4, 72.5, 72.65, 72.70, 72.9, 73.36, 73.38, 73.42, 73.5, 74.1, 74.2, 74.7, 74.8, 74.9, 75.1, 75.17, 75.23, 75.3, 75.7, 75.8, 75.9, 76.1, 77.1, 78.1, 78.5, 78.6, 79.5, 78.0, 80.7, 81.5, 97.6, 100.32, 100.34, 101.3, 101.6, 102.1, 126.1, 126.9, 127.16, 127.19, 127.3, 127.40, 127.43, 127.60, 127.63, 127.72, 127.74, 128.0, 128.05, 128.07, 129.1, 128.2, 128.26, 128.28, 128.30, 128.36, 128.41, 128.8, 137.4, 137.7, 138.09, 138.14, 138.2, 138.26, 138.29, 138.5, 138.67, 138.74, 138.9, 139.0, 139.05, 139.09; MS (MALDI, DHB) found *m/z* 2542.1 [M + Na]⁺, calcd for C₁₅₅H₁₆₂NaO₃₁ 2542.1.

Benzyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl-3,6-di-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl-3,6-di-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (32). According to the representative procedure for acetylation, alcohol 31 (76 mg, 30 μ mol) was treated with acetic anhydride (0.55 mL) in pyridine (5.5 mL) and DMAP (22 mg, 180 μ mol) for 4 h at room temperature to give acetate 32 (52 mg, 20.0 μ mol, 66%) after chromatography on a silica gel column (hexane/EtOAc, 65:35): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 2.00–2.05 (2 overlapping s, 6H) 3.03 (td, *J* = 9.7, 4.9 Hz, 1H), 3.10–3.20 (m, 4H), 3.27 (dd, *J* = 9.1, 2.8 Hz, 1H), 3.30 (dd, *J* = 9.3, 2.8 Hz, 1H), 3.33–3.42 (m, 6H), 3.44–3.67 (m, 10H), 3.71 (d, *J* = 3.0 Hz, 1H), 3.76 (t, *J* = 2.7 Hz, 1H), 3.77–3.87 (m, 1H), 3.90–3.98 (m, 3H), 3.98–4.05 (2 overlapping t, 2H), 4.07 (t, *J* = 9.2 Hz, 1H), 4.16–4.23 (m, 2H), 4.25–4.23 (2 overlapping d, 2H), 4.36–4.77 (m, 34H), 4.83 (d, *J* = 12.2 Hz, 1H), 4.93 (d, *J* = 2.3 Hz, 1H), 5.35 (d, *J* = 3.1 Hz, 1H), 5.38 (d, *J* = 3.4 Hz, 1H), 5.47 (s, 1H), 7.09–7.40 (m), 7.42–7.46 (m); ¹³C NMR (CDCl₃, 151 MHz, selected signals) δ 21.0, 67.2, 68.5, 68.6, 68.7, 68.8, 68.9, 69.0, 69.1, 69.4, 71.3, 71.38, 71.44, 72.10, 72.11, 72.4, 72.5, 72.7, 73.0, 73.3, 73.39, 73.40, 74.1, 74.2, 74.57, 74.58, 74.7, 74.9, 74.95, 75.00, 75.3, 75.66, 75.71, 75.73, 75.9, 76.1, 78.10, 78.13, 78.4, 78.6, 78.9, 80.5, 81.3, 97.7, 99.5, 99.7, 101.2, 101.3, 101.4, 102.2, 126.1, 126.8, 126.9, 127.06, 127.11, 127.15, 127.20, 127.3, 127.4, 127.5, 127.62, 127.64, 127.67, 127.70, 127.72, 127.95, 128.02, 128.07, 128.10, 128.12, 128.17, 128.20, 128.23, 128.29, 128.33, 128.4, 128.8, 137.4, 137.6, 138.2, 138.29, 138.31, 138.33, 138.38, 138.39, 138.5, 138.66, 138.71, 138.74, 138.98, 139.01, 139.1, 170.25, 170.26; MS (MALDI, DHB) found *m/z* 2625.9 [M + Na]⁺, calcd for C₁₅₉H₁₆₆NaO₃₃ 2626.1.

β -D-Mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranose (3). According to the representative procedure for global deprotection of benzyl, benzylidene, and *p*-methoxybenzyl groups, compound 32 (21 mg, 8.06 μ mol) was treated with 10% Pd/C (21 mg) in THF/MeOH/AcOH (2.0:1.0:0.3, v/v/v) (3.52 mL) under an atmosphere of H₂ for 23 h at room temperature to give compound 3 (6.4 mg, 5.95 μ mol, 74%) after reversed-phase chromatography on a C18 column (1% AcOH in H₂O). Hexasaccharide 3 existed as a mixture of regioisomers resulting from migration of the acetyl groups: white hygroscopic solid; ¹H NMR (1% CD₃CO₂D in D₂O, 600 MHz) δ 2.17 (s, integration not determined), 2.187 (s, integration not determined), 2.190 (s, integration not determined), 2.202 (s,

integration not determined), 2.211 (s, integration not determined), 3.40–3.68 (m, integration not determined), 3.71–4.08 (m, integration not determined), 4.08–4.15 (m, integration not determined), 4.17–4.21 (m, integration not determined), 4.73 (s, integration not determined), 4.75 (s, integration not determined), 4.83 (s, integration not determined), 4.90 (s, integration not determined), 4.91 (s, integration not determined), 4.92 (s, integration not determined), 4.93 (s, integration not determined), 4.94 (s, integration not determined), 5.05 (dd, $J = 9.7, 3.2$ Hz, integration not determined), 5.07–5.14 (m, integration not determined), 5.16–5.19 (m, integration not determined), 5.41 (d, $J = 3.6$ Hz, integration not determined), 5.48 (d, $J = 3.5$ Hz, integration not determined), 5.50–5.53 (m, integration not determined); ^{13}C NMR (1% $\text{CD}_3\text{CO}_2\text{D}$ in D_2O , 125 MHz, selected signals) δ 63.08, 63.11, 63.2, 63.7, 69.4, 71.3, 71.4, 71.6, 72.66, 72.69, 72.74, 72.76, 72.81, 72.89, 72.94, 73.20, 73.22, 73.4, 73.6, 74.1, 74.16, 74.21, 74.30, 74.32, 74.34, 74.4, 74.5, 75.5, 75.7, 75.9, 76.09, 76.12, 77.5, 77.66, 77.70, 77.8, 77.87, 77.92, 78.0, 79.1, 79.19, 79.24, 79.3, 79.5, 79.9, 80.1, 96.4, 96.6, 101.3, 101.8, 101.9, 102.3, 102.8, 102.85, 102.93, 103.1, 175.5, 175.7, 176.10, 176.14, 176.2; HRMS found m/z 1097.3382 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{40}\text{H}_{66}\text{NaO}_{33}$ 1097.3384.

β -D-Mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- α -D-mannopyranose (1). Representative Procedure for Saponification of Acetate. NaOH (1.0 M, 80 μL) was added to a solution of hexasaccharide **3** (5.5 mg, 5.12 μmol) in water (720 μL), and the mixture was stirred at room temperature for 2 h. The solution was passed through a short cation-exchange column (Dowex 50W-X8 H^+ form) and washed with water. The eluent was concentrated under reduced pressure and lyophilized to give compound **1** (4.6 mg, 4.34 μmol , 84%): white hygroscopic solid; ^1H NMR (D_2O , 600 MHz) δ 3.45 (ddd, $J = 9.6, 7.1, 2.4$ Hz, integration not determined), 3.51 (ddd, $J = 8.6, 5.8, 2.4$ Hz, integration not determined), 3.54–3.61 (m, integration not determined), 3.67 (dd, $J = 9.6, 3.2$ Hz, integration not determined), 3.71–4.03 (m, integration not determined), 4.08 (d, $J = 3.4$ Hz, integration not determined), 4.12–4.17 (m, integration not determined), 4.74 (s, integration not determined), 4.76 (s, integration not determined), 4.78 (s, integration not determined), 4.92 (s, integration not determined), 5.19 (d, $J = 1.5$ Hz, integration not determined); ^{13}C NMR (D_2O , 151 MHz, selected signals) δ 63.2, 63.7, 69.4, 71.7, 72.6, 72.67, 72.71, 73.0, 73.2, 73.4, 73.6, 74.15, 74.18, 74.4, 75.5, 77.5, 77.8, 79.15, 79.21, 79.27, 79.30, 79.4, 79.5, 96.4, 96.6, 102.85, 102.88, 102.93; HRMS found m/z 991.3341 $[\text{M} + \text{H}]^+$, calcd for $\text{C}_{36}\text{H}_{63}\text{O}_{31}$ 991.3353.

Allyl 2,3,6-Tri-O-benzyl-4-O-(2,3-di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl)- α -D-mannopyranoside (37). According to the representative procedure for selective β -mannosylation, the reaction of allyl 2,3,6-tri-O-benzyl- α -D-mannopyranoside (**36**, 3.62 g, 7.38 mmol) and **14** (5.03 g, 9.30 mmol) was performed, followed by quench with Et_3N and $(\text{EtO})_3\text{P}$, to give compound **37** (5.39 g, 5.85 mmol, 79%) after chromatography on silica gel (hexane/EtOAc, 83:17): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 3.05 (td, $J = 9.7, 4.8$ Hz, 1H), 3.40 (dd, $J = 9.8, 3.1$ Hz, 1H), 3.57–3.65 (m, 3H), 3.69 (d, $J = 2.9$ Hz, 1H), 3.72 (dt, $J = 9.6, 3.3$ Hz, 1H), 3.78 (t, $J = 2.8$ Hz, 1H), 3.91 (dd, $J = 8.6, 3.1$ Hz, 1H), 3.94–3.99 (m, 1H), 4.02 (dd, $J = 10.4, 5.0$ Hz, 1H), 4.06 (t, $J = 9.6$ Hz, 1H), 4.15–4.24 (m, 2H), 4.39 (d, $J = 12.2$ Hz, 1H), 4.50 (s, 1H), 4.54–4.59 (2 overlapping d, 2H), 4.63–4.84 (m, 7H), 4.90 (d, $J = 2.3$ Hz, 1H), 5.16–5.21 (m, 1H), 5.23–5.29 (m, 1H), 5.51 (s, 1H), 5.84–5.93 (m, 1H), 7.18–7.41 (m, integration not determined), 7.44–7.48 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 67.2, 68.1, 68.6, 69.1, 71.3, 72.4, 72.5, 72.8, 73.4, 74.9, 75.6, 76.1, 77.1, 77.9, 78.4, 78.7, 97.5, 101.3, 101.9, 117.2, 127.2, 127.3, 127.4, 127.5, 127.69, 127.74, 127.8, 128.0, 128.1, 128.17, 128.20, 128.23, 128.3, 128.4, 128.8, 133.8, 137.7, 138.2, 138.4, 138.6, 138.7, 139.1; MS (MALDI, DHB) found m/z 943.5 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{57}\text{H}_{60}\text{NaO}_{11}$ 943.4.

Allyl 2,3,6-Tri-O-benzyl-4-O-(2,3,6-tri-O-benzyl- β -D-mannopyranosyl)- α -D-mannopyranoside (38). According to the representative procedure for reductive ring-opening of benzylidene acetal, compound **37** (187 mg, 0.20 mmol, 1.0 equiv) was treated with Et_3SiH (395 μL , 2.45 mmol) and $\text{BF}_3 \cdot \text{OEt}_2$ (41 μL , 0.32 mmol) in CH_2Cl_2 for

7 h at 0°C to give compound **38** (112 mg, 0.12 mmol, 60%) after chromatography on silica gel (hexane/EtOAc, 80:20): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 2.71 (br s, 1H), 3.14 (dd, $J = 9.4, 2.9$ Hz, 1H), 3.22 (dt, $J = 9.6, 4.8$ Hz, 1H), 3.58 (dd, $J = 10.8, 5.4$ Hz, 1H), 3.61–3.70 (m, 2H), 3.67–3.73 (m, 2H), 3.76–3.81 (m, 2H), 3.90–3.99 (m, 3H), 4.14–4.19 (m, 1H), 4.24 (t, $J = 9.0$ Hz, 1H), 4.37–4.49 (m, 5H), 4.51 (s, 1H), 4.56 (d, $J = 12.2$ Hz, 1H), 4.63–4.72 (m, 4H), 4.78 (d, $J = 12.0$ Hz, 1H), 4.84 (d, $J = 12.1$ Hz, 1H), 4.89 (d, $J = 2.2$ Hz, 1H), 5.15–5.19 (m, 1H), 5.21–5.27 (m, 1H), 5.82–5.92 (m, 7.16–7.37 (m, integration not determined); ^{13}C NMR (CDCl_3 , 151 MHz) δ 68.0, 68.8, 69.4, 71.1, 71.4, 71.5, 72.5, 72.7, 73.4, 73.7, 74.2, 74.65, 74.67, 75.5, 75.7, 78.2, 81.8, 97.6, 101.4, 117.2, 127.2, 127.3, 127.4, 127.45, 127.50, 127.58, 127.60, 127.71, 127.73, 127.8, 128.06, 128.13, 128.2, 128.3, 128.37, 128.43, 133.9, 138.05, 138.10, 138.3, 138.4, 138.9, 139.2; MS (MALDI, DHB) found m/z 945.4 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{57}\text{H}_{62}\text{NaO}_{11}$ 945.4.

Allyl 3-O-Benzyl-4,6-benzylidene-2-O-*p*-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (39). According to the representative procedure for selective β -mannosylation, the reaction of compounds **38** (250 mg, 0.27 mmol) and **15** (193 mg, 0.34 mmol) was performed, followed by quench with Et_3N and $(\text{EtO})_3\text{P}$, to give compound **39** (200 mg, 0.14 mmol, 53%) after chromatography on silica gel (toluene/EtOAc, 95:5): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 3.04 (td, $J = 9.7, 4.9$ Hz, 1H), 3.17–3.24 (m, 1H), 3.31 (dd, $J = 9.2, 3.0$ Hz, 1H), 3.38 (dd, $J = 9.9, 3.0$ Hz, 1H), 3.48–3.57 (m, 3H), 3.62–3.71 (m, 3H), 3.73 (d, $J = 3.1$ Hz, 1H), 3.75–3.80 (m, 5H), 3.92–3.98 (m, 3H), 4.02 (t, $J = 9.6$ Hz, 1H), 4.11–4.19 (m, 2H), 4.23 (t, $J = 9.1$ Hz, 1H), 4.29 (d, $J = 11.9$ Hz, 1H), 4.41 (d, $J = 12.2$ Hz, 1H), 4.43–4.52 (m, 3H), 4.52–4.54 (2 overlapping s, 2H), 4.59 (d, $J = 12.4$ Hz, 1H), 4.61–4.79 (m, 9H), 4.86–4.91 (2 overlapping d, 2H), 5.15–5.19 (m, 1H), 5.21–5.27 (m, 1H), 5.47 (s, 1H), 5.81–5.94 (m, 1H), 6.78–6.82 (m, 2H), 7.10–7.39 (m, integration not determined), 7.41–7.47 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 55.2, 67.2, 67.9, 68.5, 69.29, 69.33, 71.3, 72.2, 72.3, 72.6, 72.7, 73.4, 73.5, 74.2, 74.5, 75.4, 75.6, 75.7, 75.9, 76.1, 76.7, 78.4, 78.6, 80.7, 97.5, 101.2, 101.5, 102.2, 113.5, 117.2, 126.1, 126.9, 127.1, 127.16, 127.19, 127.3, 127.4, 127.5, 127.59, 127.64, 127.7, 127.75, 127.76, 128.0, 128.10, 128.13, 128.2, 128.3, 128.7, 129.7, 130.9, 133.8, 137.7, 138.3, 138.37, 138.41, 138.6, 139.06, 139.10, 139.3, 159.1; MS (MALDI, DHB) found m/z 1405.6 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{85}\text{H}_{90}\text{NaO}_{17}$ 1405.6.

Allyl 3-O-Benzyl-2-O-*p*-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (40). According to the representative procedure for transacetalation, compound **39** (969 mg, 0.70 mmol) was treated with neopentyl glycol in the presence of camphorsulfonic acid to give compound **40** (657 mg, 0.51 mmol, 72%) after chromatography on silica gel (hexane/EtOAc, 60:40): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 3.04–3.10 (m, 2H), 3.23 (dt, $J = 9.7, 3.4$ Hz, 1H), 3.26 (dd, $J = 9.4, 2.9$ Hz, 1H), 3.41 (dd, $J = 11.6, 6.5$ Hz, 1H), 3.55–3.60 (2 overlapping dd, 2H), 3.64–3.71 (m, 3H), 3.72–3.74 (2 overlapping d, 2H), 3.75–3.80 (m, 6H), 3.92–3.99 (m, 2H), 4.11–4.19 (m, 3H), 4.27 (t, $J = 9.2$ Hz, 1H), 4.30–4.35 (2 overlapping d, 2H), 4.41 (d, $J = 12.0$ Hz, 1H), 4.46–4.54 (m, 4H), 4.60 (d, $J = 12.4$ Hz, 1H), 4.62–4.79 (m, 8H), 4.88–4.93 (2 overlapping d, 2H), 5.15–5.21 (m, 1H), 5.21–5.28 (m, 1H), 5.83–5.92 (m, 1H), 6.77–6.83 (m, 2H), 7.14–7.34 (m, integration not determined), 7.36–7.39 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz, selected signals) δ 55.2, 62.7, 67.2, 67.9, 69.3, 69.5, 70.9, 71.4, 72.6, 72.65, 72.74, 73.3, 73.4, 73.6, 73.7, 74.4, 75.46, 75.51, 75.54, 75.6, 75.8, 76.5, 78.3, 80.2, 81.9, 97.5, 101.33, 101.34, 113.6, 117.2, 127.1, 127.16, 127.18, 127.3, 127.4, 127.45, 127.49, 127.6, 127.7, 127.8, 127.9, 128.0, 128.1, 128.2, 128.26, 128.32, 128.4, 129.6, 130.7, 133.8, 137.8, 138.27, 138.30, 138.4, 138.7, 138.9, 139.3, 159.1; MS (MALDI, DHB) found m/z 1317.6 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{78}\text{H}_{86}\text{NaO}_{17}$ 1317.6.

Allyl 3,6-Di-O-benzyl-2-O-*p*-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (35). According to the representative procedure for selective monobenzoylation, diol **40** (765

mg, 0.59 mmol) was heated with Bu₂SnO (294 mg, 1.18 mmol), followed by treatment with BnBr (280 μL, 2.34 mmol) in the presence of CsF (404 mg, 2.66 mmol), to give compound 35 (652 mg, 0.47 mmol, 80%) after chromatography on silica gel (hexane/EtOAc, 70:30): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 2.67 (br s, 1H), 3.12 (dd, *J* = 9.4, 2.9 Hz, 1H), 3.19 (dt, *J* = 9.7, 4.8 Hz, 1H), 3.26 (ddd, *J* = 9.5, 4.1, 2.4 Hz, 1H), 3.34 (d, *J* = 9.2, 2.9 Hz, 1H), 3.53 (dd, *J* = 10.5, 5.4 Hz, 1H), 3.55–3.66 (m, 5H), 3.69 (d, *J* = 2.9 Hz, 1H), 3.73 (d, *J* = 2.9 Hz, 1H), 3.74–3.78 (m, 5H), 3.89 (t, *J* = 9.4 Hz, 1H), 3.92–3.98 (m, 2H), 4.14–4.20 (m, 2H), 4.22 (t, *J* = 9.1 Hz, 1H), 4.29–4.34 (2 overlapping d, 2H), 4.36–4.43 (m, 4H), 4.47 (d, *J* = 12.2 Hz, 1H), 4.50–4.54 (d overlapped with 2 s, 3H), 4.56–4.67 (m, 4H), 4.68–4.80 (m, 5H), 4.87–4.91 (2 overlapping d, 2H), 5.14–5.19 (m, 1H), 5.21–5.27 (m, 1H), 5.83–5.91 (m, 1H), 6.75–6.80 (m, 2H), 7.11–7.39 (m, integration not determined); ¹³C NMR (CDCl₃, 151 MHz, selected signals) δ 55.2, 67.9, 68.7, 69.4, 69.5, 71.1, 71.3, 71.4, 72.3, 72.6, 72.7, 73.3, 73.5, 73.6, 73.8, 74.07, 74.12, 74.6, 75.3, 75.5, 75.66, 75.73, 76.1, 78.3, 80.7, 81.8, 97.5, 101.4, 101.6, 113.5, 117.2, 127.1, 127.15, 127.21, 127.4, 127.46, 127.51, 127.56, 127.61, 127.64, 127.7, 127.8, 127.9, 128.13, 128.14, 128.2, 128.25, 128.32, 128.4, 129.5, 131.0, 133.9, 138.11, 138.14, 138.3, 138.4, 138.5, 139.1, 139.2, 139.3, 159.0; MS (MALDI, DHB) found 1407.6256 [M + Na]⁺, calcd for C₈₈H₉₂NaO₁₇ 1407.6232.

Allyl 2,3,6-Tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2-O-benzoyl-3,6-di-O-benzyl-β-D-glucopyranoside (41). According to the representative procedure for reductive ring-opening of benzylidene acetal, trisaccharide 24 (1.031 g, 0.75 mmol) was treated with Et₃SiH (365 μL, 2.26 mmol) and TfOH (225 μL, 2.54 mmol) at –78 °C for 15 h, followed by stirring with TBAF (7.0 mL, 1 M solution in THF) for 2 h at room temperature, to give compound 41 (597 mg, 0.44 mmol, 58%) after chromatography on silica gel (toluene/EtOAc, 92:8): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 2.69 (br s, 1H), 3.14 (dd, *J* = 9.4, 2.9 Hz, 1H), 3.22 (dt, *J* = 9.6, 4.8 Hz, 1H), 3.29 (ddd, *J* = 9.4, 4.3, 2.3 Hz, 1H), 3.33 (dd, *J* = 9.3, 2.9 Hz, 1H), 3.48 (ddd, *J* = 9.6, 3.9, 2.4 Hz, 1H), 3.53–3.57 (m, 2H), 3.58–3.64 (m, 3H), 3.68–3.78 (m, 4H), 3.92 (t, *J* = 9.4 Hz, 1H), 4.02–4.10 (m, 2H), 4.21 (t, *J* = 9.3 Hz, 1H), 4.26–4.32 (m, 2H), 4.34 (d, *J* = 11.8 Hz, 1H), 4.40–4.49 (m, 6H), 4.53–4.57 (s overlapped with s and d, 3H), 4.60 (d, *J* = 12.1 Hz, 1H), 4.66 (d, *J* = 12.2 Hz, 1H), 4.71 (d, *J* = 12.0 Hz, 1H), 4.78–4.85 (m, 4H), 4.99 (d, *J* = 12.2 Hz, 1H), 5.05–5.09 (m, 1H), 5.16–5.22 (m, 1H), 5.28 (dd, *J* = 9.3, 7.7 Hz, 1H), 5.70–5.81 (m, 1H), 6.91–6.96 (m, 2H), 6.97–7.02 (m, 1H), 7.03–7.07 (m, 2H), 7.14–7.32 (m, integration not determined), 7.33–7.42 (m, 6H), 7.51–7.55 (m, 1H), 7.94–7.97 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz) δ 68.7, 68.8, 69.5, 71.0, 71.4, 72.6, 73.1, 73.4, 73.5, 73.6, 74.06, 74.10, 74.2, 74.6, 74.7, 75.0, 75.4, 76.1, 77.8, 80.5, 80.6, 81.8, 99.9, 101.2, 101.5, 117.1, 126.9, 127.1, 127.18, 127.23, 127.3, 127.4, 127.5, 127.55, 127.56, 127.59, 127.66, 127.67, 127.77, 127.82, 127.9, 128.0, 128.09, 128.12, 128.19, 128.24, 128.3, 128.38, 128.42, 129.8, 130.1, 132.9, 133.8, 137.9, 138.06, 138.10, 138.3, 138.7, 138.86, 138.91; MS (MALDI, DHB) found *m/z* 1391.6 [M + Na]⁺, calcd for C₆₄H₈₈NaO₁₇ 1391.6.

Allyl 2,3-Di-O-benzyl-4,6-O-benzylidene-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2-O-benzoyl-3,6-di-O-benzyl-β-D-glucopyranoside (43). According to the representative procedure for β-selective mannosylation, the reaction of compounds 41 (112 mg, 82 μmol) and 14 (58 mg, 107 μmol) was performed, followed by quench with Et₃N and (EtO)₃P, to give compound 43 (100 mg, 56 μmol, 68%) after chromatography on silica gel (toluene/EtOAc, 92:8): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 3.04 (td, *J* = 9.7, 4.9 Hz, 1H), 3.16–3.21 (m, 1H), 3.25 (ddd, *J* = 9.5, 4.2, 2.3 Hz, 1H), 3.28–3.35 (2 overlapping dd, 2H), 3.39 (dd, *J* = 9.9, 3.0 Hz, 1H), 3.45–3.64 (m, 7H), 3.68–3.74 (m, 4H), 3.77 (t, *J* = 9.0 Hz, 1H), 3.94 (dd, *J* = 10.4, 4.9 Hz, 1H), 4.01–4.09 (m, 3H), 4.14 (t, *J* = 9.4 Hz, 1H), 4.21 (t, *J* = 9.4 Hz, 1H), 4.23–4.28 (m, 2H), 4.28–4.33 (m, 1H), 4.38–4.46 (3 overlapping d, 3H), 4.47–4.57 (m, 7H), 4.60 (d, *J* = 12.2 Hz, 1H), 4.62–4.88 (m, 10H), 4.99 (d, *J* = 12.2 Hz, 1H), 5.02–5.10 (m, 1H), 5.15–5.22 (m, 1H), 5.29 (dd, *J* = 9.3, 7.8 Hz, 1H), 5.48 (s, 1H), 5.75 (dddd, *J* = 4.7, 6.2, 10.7, 17.0 Hz, 1H),

6.91–6.95 (m, 1H), 6.97–7.02 (m, 1H), 7.02–7.06 (m, 1H), 7.11–7.46 (m, integration not determined), 7.51–7.56 (m, 1H), 7.94–7.97 (m, 1H); ¹³C NMR (CDCl₃, 600 MHz, selected signals) δ 67.2, 68.5, 68.7, 69.2, 69.4, 69.5, 72.1, 72.3, 72.5, 73.1, 73.3, 73.4, 73.5, 74.0, 74.1, 74.2, 74.9, 75.0, 75.3, 75.4, 75.5, 75.8, 76.0, 76.1, 77.1, 77.7, 78.4, 78.6, 80.5, 80.7, 99.8, 101.15, 101.22, 101.6, 102.1, 117.1, 126.0, 126.9, 127.0, 127.1, 127.16, 127.18, 127.24, 127.3, 127.4, 127.5, 127.6, 127.66, 127.69, 127.75, 127.78, 127.83, 127.99, 128.02, 128.03, 128.05, 128.07, 128.09, 128.13, 128.18, 128.22, 128.3, 128.4, 128.7, 129.7, 130.0, 132.8, 133.7, 137.6, 137.9, 138.3, 138.4, 138.5, 138.65, 138.72, 138.9, 139.0, 139.2, 165.2; MS (MALDI, DHB) found *m/z* 1821.8 [M + Na]⁺, calcd for C₁₁₁H₁₁₄NaO₂₂ 1821.8.

2,3-Di-O-benzyl-4,6-O-benzylidene-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2-O-benzoyl-3,6-di-O-benzyl-β-D-glucopyranoside (44). According to the representative procedure for deallylation, compound 43 (109 mg, 60.6 μmol) was treated with a catalytic amount of PdCl₂ in MeOH/CH₂Cl₂ at room temperature for 4 h to give compound 44 (84 mg, 47.7 μmol, 79%) after chromatography on silica gel (hexane/EtOAc, 70:30): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 2.99 (br s, integration not determined), 3.04 (td, *J* = 9.7, 5.0 Hz, integration not determined), 3.16–3.21 (m, integration not determined), 3.22–3.27 (m, integration not determined), 3.27–3.32 (m, integration not determined), 3.39 (dd, *J* = 9.8, 2.6 Hz, integration not determined), 3.46–3.64 (m, integration not determined), 3.66–3.75 (m, integration not determined), 3.80 (t, *J* = 9.0 Hz, integration not determined), 3.94 (dd, *J* = 10.4, 4.8 Hz, integration not determined), 3.98–4.16 (m, integration not determined), 4.18–4.28 (m, integration not determined), 4.34–4.45 (m, integration not determined), 4.45–4.55 (m, integration not determined), 4.56–4.90 (m, integration not determined), 5.03–5.13 (m, integration not determined), 5.48 (s, integration not determined), 5.54 (d, *J* = 3.5 Hz, integration not determined), 6.95–7.58 (m, integration not determined), 7.91–8.00 (m, integration not determined); ¹³C NMR (CDCl₃, 151 MHz, selected signals) δ 67.2, 68.4, 68.5, 68.6, 69.2, 69.4, 69.5, 70.1, 72.11, 72.14, 72.4, 72.5, 72.6, 73.3, 73.4, 73.46, 73.50, 73.6, 74.17, 74.19, 74.24, 74.5, 74.8, 74.9, 75.35, 75.37, 75.41, 75.58, 75.60, 75.8, 75.9, 76.0, 76.06, 76.12, 76.2, 77.1, 77.5, 77.6, 77.8, 78.4, 78.6, 80.1, 80.6, 80.7, 90.4, 96.1, 101.05, 101.14, 101.2, 101.62, 101.64, 102.2, 126.1, 126.9, 126.95, 127.05, 127.11, 127.13, 127.17, 127.20, 127.27, 127.31, 127.34, 127.36, 127.38, 127.43, 127.48, 127.54, 127.6, 127.67, 127.71, 127.81, 127.83, 127.88, 127.93, 127.95, 128.01, 128.05, 128.08, 128.10, 128.16, 128.21, 128.24, 128.25, 128.28, 128.33, 128.47, 128.49, 128.7, 129.3, 129.6, 129.8, 129.9, 133.1, 133.4, 137.57, 137.64, 138.25, 138.33, 138.4, 138.5, 138.6, 138.7, 138.9, 139.0, 139.25, 139.27, 165.8, 167.1; MS (MALDI, DHB) found *m/z* 1781.7 [M + Na]⁺, calcd for C₁₀₈H₁₁₀NaO₂₂ 1781.7.

2,3-Di-O-benzyl-4,6-O-benzylidene-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2-O-benzoyl-3,6-di-O-benzyl-α/β-D-glucopyranosyl trichloroacetimidate (34). According to the representative procedure for trichloroacetimidation, compound 44 (113 mg, 64.2 μmol) was treated with trichloroacetimidate (65 μL, 648.2 μmol) in the presence of DBU at 0 °C for 1 h to give compound 34 (68 mg, 35.7 μmol, 56%) after chromatography (hexane/EtOAc/Et₃N, 75:25:1): colorless oil; ¹H NMR (C₆D₆, 600 MHz) δ 3.04 (td, *J* = 9.6, 4.9 Hz, integration not determined), 3.23–3.32 (m, integration not determined), 3.33–3.39 (m, integration not determined), 3.41–3.48 (m, integration not determined), 3.55–3.73 (m, integration not determined), 3.76 (d, *J* = 3.1 Hz, integration not determined), 3.80–3.86 (m, integration not determined), 3.92 (d, *J* = 2.9 Hz, integration not determined), 3.98 (dd, *J* = 10.2, 4.8 Hz, integration not determined), 4.16–4.24 (m, integration not determined), 4.25–4.33 (m, integration not determined), 4.34–4.39 (m, integration not determined), 4.44–4.59 (m, integration not determined), 4.59–4.77 (m, integration not determined), 4.87–5.09 (m, integration not determined), 5.17–5.21 (m, integration not determined), 5.22–5.28 (m, integration not determined), 5.32 (d, *J* = 12.6 Hz, integration not determined), 5.43 (d, *J* = 12.3 Hz, integration

not determined), 5.67–5.75 (m, integration not determined), 6.01 (t, $J = 8.5$ Hz, integration not determined), 6.26 (d, $J = 7.9$ Hz, integration not determined), 6.91–7.39 (m, integration not determined), 7.45–7.50 (m, integration not determined), 7.52–7.60 (m, integration not determined), 8.06–8.17 (m, integration not determined), 8.37 (s, integration not determined), 8.58 (s, integration not determined); ^{13}C NMR (C_6D_6 , 151 MHz, selected signals) δ 68.0, 69.0, 69.2, 70.0, 70.1, 72.9, 73.2, 73.3, 73.9, 74.15, 74.20, 74.22, 74.4, 75.2, 75.3, 75.4, 76.0, 76.46, 76.53, 76.8, 77.7, 77.76, 77.83, 78.2, 78.9, 79.6, 79.7, 81.4, 81.8, 92.0, 95.0, 101.8, 102.0, 102.3, 102.8, 127.0, 127.6, 127.75, 127.80, 127.88, 127.93, 127.95, 128.00, 128.1, 128.79, 128.81, 128.88, 128.94, 128.97, 129.03, 129.2, 130.5, 130.6, 133.5, 138.7, 139.1, 139.35, 139.41, 139.9, 139.97, 140.03, 140.1, 140.5, 140.6, 161.0, 166.0; MS (MALDI, ATT) found m/z 1924.6 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{110}\text{H}_{110}\text{Cl}_3\text{NNaO}_{22}$ 1924.6.

Alllyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-benzoyl-3,6-di-O-benzyl- β -D-glucopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl-2-O-*p*-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (45). According to the representative procedure for glycosylation, the reaction of acceptor 35 (63 mg, 45.5 μmol) and trichloroacetimidate 34 (130 mg, 68.3 μmol) was carried out by the promotion of TMSOTf to give compound 45 (92 mg, 29.4 μmol , 65%) after column chromatography on silica gel (toluene/EtOAc, 90:10): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 2.86–2.92 (m, 1H), 3.03 (td, $J = 9.7, 4.9$ Hz, 1H), 3.13–3.23 (m, 5H), 3.23–3.30 (m, 4H), 3.35–3.40 (m, 2H), 3.42–3.66 (m, 14H), 3.68–3.76 (m, 8H), 3.89–3.96 (m, 3H), 3.97–4.09 (m, 4H), 4.10–4.28 (m, 9H), 4.32 (s, 1H), 4.35–4.59 (m, 18H), 4.61–4.86 (m, 19H), 4.87 (d, $J = 2.0$ Hz, 1H), 4.99 (d, $J = 12.2$ Hz, 1H), 5.14–5.24 (m, 3H), 5.47 (s, 1H), 5.82–5.89 (m, 1H), 6.65–6.69 (m, 2H), 6.85–6.89 (m, 2H), 6.94–6.98 (m, 3H), 7.00–7.40 (m, integration not determined), 7.42–7.45 (m, 2H), 7.51–7.57 (m, 1H), 7.84–7.89 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz, selected signals) δ 55.2, 67.2, 67.9, 68.3, 68.49, 68.52, 69.2, 69.28, 69.34, 69.4, 71.3, 72.1, 72.3, 72.37, 72.42, 72.6, 72.7, 73.28, 73.30, 73.3, 73.4, 73.7, 74.0, 74.1, 74.2, 74.3, 74.9, 75.1, 75.36, 75.40, 75.41, 75.45, 75.54, 75.6, 75.71, 75.74, 75.9, 76.0, 76.1, 77.1, 78.0, 78.3, 78.4, 78.6, 80.4, 80.5, 80.76, 80.79, 80.9, 97.5, 100.5, 101.26, 101.32, 101.6, 102.2, 113.4, 117.1, 126.1, 126.8, 126.87, 126.91, 126.97, 127.02, 127.1, 127.16, 127.18, 127.22, 127.3, 127.35, 127.42, 127.49, 127.54, 127.56, 127.60, 127.65, 127.70, 127.73, 127.8, 127.9, 128.00, 128.02, 128.06, 128.08, 128.11, 128.17, 128.20, 128.24, 128.28, 128.31, 128.7, 129.3, 129.8, 129.9, 131.3, 132.9, 133.9, 137.7, 138.1, 138.28, 138.31, 138.37, 138.42, 138.5, 138.7, 138.77, 138.82, 139.0, 139.06, 139.09, 139.3, 139.4, 158.8, 165.0; MS (MALDI, DHB) found m/z 3148.4 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{193}\text{H}_{200}\text{NaO}_{38}$ 3148.4.

Alllyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl- β -D-glucopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl-2-O-*p*-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (46). According to the representative procedure for debenzoylation, compound 45 (56 mg, 17.9 μmol) was treated with LiAlH_4 (2.7 mg, 71.1 μmol) in Et_2O to give compound 46 (43 mg, 14.2 μmol , 80%) after chromatography on silica gel (hexane/EtOAc, 70:30): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 3.02 (td, $J = 9.6, 4.8$ Hz, 1H), 3.08–3.17 (m, 3H), 3.18–3.28 (m, 6H), 3.29–3.42 (m, 5H), 3.42–3.82 (m, 23H), 3.90–3.98 (m, 3H), 4.02 (t, $J = 9.6$ Hz, 1H), 4.09–4.25 (m, 9H), 4.29 (d, $J = 12.0$ Hz, 1H), 4.31–4.43 (m, 7H), 4.44–4.54 (m, 12H), 4.58 (d, $J = 12.5$ Hz, 1H), 4.59–4.80 (m, 18H), 4.85–4.90 (2 overlapping d, 2H), 5.00 (d, $J = 11.9$ Hz, 1H), 5.14–5.19 (m, 1H), 5.21–5.26 (m, 1H), 5.47 (s, 1H), 5.82–5.92 (m, 1H), 6.68–6.73 (m, 2H), 7.09–7.39 (m, integration not determined), 7.41–7.46 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz, selected signals) δ 55.2, 67.2, 67.9, 68.5, 68.6, 69.2, 69.3, 69.4, 69.6, 71.3, 71.4, 72.1, 72.26, 72.31, 72.4, 72.6, 72.7, 73.2, 73.3, 73.4, 73.6, 73.8, 74.09, 74.14, 74.2, 74.66, 74.69, 74.8, 74.9, 75.0, 75.2, 75.28, 75.31, 75.46, 75.48, 75.53, 75.6, 75.7, 75.9, 76.06, 76.13, 77.0, 77.1, 78.3, 78.4, 78.6, 80.7, 80.8, 81.8, 82.4, 97.5,

101.1, 101.2, 101.4, 101.7, 101.9, 102.2, 103.7, 113.5, 117.2, 126.1, 126.7, 126.8, 126.9, 127.00, 127.01, 127.11, 127.13, 127.16, 127.18, 127.3, 127.37, 127.41, 127.5, 127.55, 127.61, 127.62, 127.67, 127.71, 127.73, 127.8, 127.97, 127.98, 128.05, 128.08, 128.13, 128.15, 128.19, 128.21, 128.23, 128.27, 128.31, 128.7, 129.3, 131.0, 133.8, 137.7, 137.9, 138.0, 138.3, 138.4, 138.5, 138.7, 138.8, 139.00, 139.04, 139.08, 139.09, 139.12, 139.27, 139.30, 139.5, 158.9; MS (MALDI, DHB) found m/z 3044.3 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{186}\text{H}_{196}\text{NaO}_{37}$ 3044.3.

Alllyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl-2-O-*p*-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (33). According to the representative procedure for *gluco* to *manno* epimerization, alcohol 46 (43 mg, 14.2 μmol) was subject to oxidation with DMSO (0.48 mL) and acetic anhydride (0.24 mL) for 16 h at room temperature, and the resulting carbonyl compound was reduced by NaBH_4 (7 mg, 185.0 μmol) to give compound 33 (26 mg, 8.6 μmol , 60%) after column chromatography on silica gel (hexane/EtOAc, 65:35): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 2.69 (br s, 1H), 3.03 (td, $J = 9.7, 4.9$ Hz, 1H), 3.12–3.25 (m, 5H), 3.26–3.40 (m, 6H), 3.42–3.60 (m, 10H), 3.60–3.80 (m, 13H), 3.90 (br s, 1H), 3.91–3.98 (m, 3H), 4.00–4.07 (2 overlapping t, 2H), 4.10–4.33 (m, 11H), 4.34–4.54 (m, 18H), 4.56–4.80 (m, 20H), 4.85–4.91 (2 overlapping d, 2H), 5.14–5.19 (m, 1H), 5.21–5.26 (m, 1H), 5.47 (s, 1H), 5.80–5.93 (m, 1H), 6.70–6.76 (m, 2H), 7.08–7.38 (m, integration not determined), 7.41–7.47 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz, selected signals) δ 55.2, 67.2, 67.9, 68.1, 68.5, 69.1, 69.2, 69.35, 69.36, 69.43, 69.5, 71.4, 71.7, 71.8, 72.1, 72.26, 72.34, 72.4, 72.6, 72.7, 73.2, 73.3, 73.35, 73.37, 73.41, 73.44, 73.7, 74.0, 74.12, 74.14, 74.7, 74.89, 74.92, 75.1, 75.2, 75.29, 75.33, 75.4, 75.52, 75.54, 75.6, 75.7, 75.9, 76.0, 76.2, 77.1, 78.3, 78.4, 78.6, 79.4, 80.7, 80.76, 80.84, 81.4, 97.5 ($^1J_{\text{CH}} = 169.3$ Hz), 100.2 ($^1J_{\text{CH}} = 159.3$ Hz), 101.3, 101.4 ($^1J_{\text{CH}} = 155.7$ Hz), 101.56 ($^1J_{\text{CH}} = 157.9$ Hz), 101.61 ($^1J_{\text{CH}} = 156.4$ Hz), 101.8 ($^1J_{\text{CH}} = 155.7$ Hz), 102.2 ($^1J_{\text{CH}} = 157.1$ Hz), 113.5, 117.2, 126.1, 126.90, 126.94, 127.0, 127.1, 127.2, 127.25, 127.29, 127.35, 127.37, 127.41, 127.44, 127.5, 127.55, 127.62, 127.64, 127.67, 127.70, 127.72, 127.73, 127.8, 127.96, 127.98, 128.05, 128.09, 128.11, 128.13, 128.15, 128.19, 128.22, 128.23, 128.3, 128.7, 129.3, 131.1, 133.9, 137.7, 138.29, 138.30, 138.33, 138.4, 138.6, 138.7, 138.8, 139.05, 139.11, 139.15, 139.21, 139.3, 158.9; MS (MALDI, DHB) found m/z 3044.3 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{186}\text{H}_{196}\text{NaO}_{37}$ 3044.3.

Alllyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl-3,6-di-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl-2-O-*p*-methoxybenzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (47). According to the representative procedure for acetylation, alcohol 33 (94 mg, 31.1 μmol) was treated with acetic anhydride (0.4 mL) in pyridine in the presence of DMAP for 3 h at room temperature to give acetate 47 (87 mg, 28.4 μmol , 92%) after column chromatography on silica gel (hexane/EtOAc, 70:30): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 2.01 (s, 3H), 3.02 (td, $J = 9.7, 4.9$ Hz, 1H), 3.11–3.20 (m, 4H), 3.22 (dt, $J = 9.4, 3.2$ Hz, 1H), 3.26–3.35 (m, 5H), 3.35–3.60 (m, 12H), 3.60–3.73 (m, 10H), 3.74–3.79 (m, 2H), 3.89–3.99 (m, 4H), 4.02 (t, $J = 9.5$ Hz, 1H), 4.07–4.24 (m, 8H), 4.27–4.32 (2 overlapping d, 2H), 4.34 (d, $J = 12.2$ Hz, 1H), 4.36–4.81 (m, 38H), 4.85–4.90 (2 overlapping d, 2H), 5.13–5.19 (m, 1H), 5.20–5.26 (m, 1H), 5.39 (d, $J = 3.4$ Hz, 1H), 5.47 (s, 1H), 5.78–5.96 (m, 1H), 6.65–6.71 (m, 2H), 7.09–7.39 (m, integration not determined), 7.41–7.46 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz, selected signals) δ 21.0, 55.2, 67.2, 67.9, 68.5, 68.8, 69.0, 69.1, 69.2, 69.36, 69.44, 71.36, 71.41, 71.9, 72.1, 72.2, 72.3, 72.4, 72.6, 72.7, 73.1, 73.3, 73.36, 73.38, 73.40, 73.7, 74.1, 74.2, 74.5, 74.9, 75.1, 75.25, 75.29, 75.40, 75.42, 75.5, 75.6, 75.7, 75.8, 76.09, 76.13, 76.2, 77.1, 78.1, 78.3, 78.4, 78.6, 80.7, 80.8, 81.3, 97.5, 99.7, 101.1, 101.3, 101.4, 101.68, 101.70, 102.2, 113.4, 117.1, 126.1, 126.8, 126.85, 126.90, 127.01, 127.04, 127.09, 127.13, 127.15, 127.21, 127.27, 127.34, 127.37, 127.41, 127.44, 127.5, 127.59, 127.64, 127.65, 127.70, 127.72, 127.8, 127.96,

127.99, 128.05, 128.07, 128.09, 128.12, 128.16, 128.21, 128.3, 128.7, 129.2, 131.2, 133.9, 137.7, 138.3, 138.35, 138.38, 138.43, 138.45, 138.48, 138.54, 138.77, 138.80, 139.07, 139.10, 139.15, 139.20, 139.23, 139.3, 158.8, 170.3; MS (MALDI, DHB) found m/z 3086.1 [M + Na]⁺, calcd for C₁₈₈H₁₉₈NaO₃₈ 3086.3.

2,3-Di-O-benzyl-4,6-benzylidene-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2-O-acetyl-3,6-di-O-benzyl-β-D-mannopyranosyl-(1→4)-3,6-di-O-benzyl-2-O-p-methoxybenzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-α-D-mannopyranose (48). According to the representative procedure for deallylation, compound 47 (83 mg, 27.1 μmol) was treated with a catalytic amount of PdCl₂ in MeOH/CH₂Cl₂ (1:1, v/v, 4.0 mL) at room temperature for 3 h to give compound 48 (49 mg, 16.2 μmol, 60%) after column chromatography on silica gel (hexane/EtOAc, 60:40): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 2.01 (s, integration not determined), 2.70 (br s, integration not determined), 3.02 (td, $J = 9.7, 4.9$ Hz, integration not determined), 3.11–3.20 (m, integration not determined), 3.22–3.74 (m, integration not determined), 3.75 (t, $J = 2.8$ Hz, integration not determined), 3.77–3.79 (m, integration not determined), 3.90–4.26 (m, integration not determined), 4.28–4.84 (m, integration not determined), 4.92–4.97 (m, integration not determined), 5.22 (d, $J = 2.1$ Hz, integration not determined), 5.39 (d, $J = 3.6$ Hz, 1H, integration not determined), 5.47 (s, integration not determined), 6.66–6.71 (m, integration not determined), 7.09–7.39 (m, integration not determined), 7.41–7.46 (m, integration not determined); ¹³C NMR (CDCl₃, 151 MHz, selected signals) δ 21.0, 55.2, 67.2, 68.5, 68.8, 69.0, 69.1, 69.2, 69.45, 69.52, 69.6, 71.4, 71.5, 71.91, 71.94, 72.1, 72.2, 72.28, 72.33, 72.4, 72.5, 72.7, 73.1, 73.4, 73.5, 73.6, 73.71, 73.74, 74.0, 74.1, 74.2, 74.3, 74.5, 74.8, 74.9, 75.1, 75.3, 75.40, 75.42, 75.5, 75.7, 75.8, 75.9, 76.0, 76.07, 76.11, 76.14, 76.2, 77.15, 77.20, 78.0, 78.4, 78.6, 80.4, 80.5, 80.69, 80.73, 81.3, 93.0, 93.4, 99.7, 101.09, 101.14, 101.3, 101.58, 101.61, 101.7, 102.2, 113.4, 126.1, 126.8, 126.86, 126.90, 127.1, 127.16, 127.19, 127.23, 127.35, 127.37, 127.43, 127.45, 127.52, 127.56, 127.60, 127.65, 127.70, 127.74, 127.77, 127.81, 127.9, 128.0, 128.06, 128.08, 128.10, 128.12, 128.15, 128.19, 128.24, 128.29, 128.34, 128.4, 128.5, 128.7, 129.3, 131.12, 137.7, 138.1, 138.3, 138.37, 138.43, 138.5, 138.75, 138.77, 138.9, 139.0, 139.06, 139.09, 139.12, 139.2, 158.8, 170.3; MS (MALDI, DHB) found m/z 3046.4 [M + Na]⁺, calcd for C₁₈₅H₁₉₄NaO₃₈ 3046.3.

β-D-Mannopyranosyl-(1→4)-β-D-mannopyranosyl-(1→4)-β-D-mannopyranosyl-(1→4)-2-O-acetyl-β-D-mannopyranosyl-(1→4)-β-D-mannopyranosyl-(1→4)-β-D-mannopyranosyl-(1→4)-D-mannopyranose (5). According to the representative procedure for global deprotection of benzyl, benzylidene, and *p*-methoxybenzyl groups, compound 48 (10 mg, 3.31 μmol) was treated with 10% Pd/C (10 mg) in THF/MeOH/AcOH (2.0:1.0:0.3, v/v/v, 1.2 mL) under an atmosphere of hydrogen at room temperature to give compound 5 (2.7 mg, 2.26 μmol, 69%) after reversed-phase chromatography on a C18 column (1% AcOH in H₂O). Heptasaccharide 5 was isolated as a mixture of regioisomers resulting from the migration of the acetyl group: white hygroscopic solid; ¹H NMR (1% CD₃CO₂D in D₂O, 600 MHz) δ 2.18 (s, integration not determined), 2.19 (s, integration not determined), 3.40–3.60 (m, integration not determined), 3.60–3.67 (m, integration not determined), 3.69–4.08 (m, integration not determined), 4.08–4.15 (m, integration not determined), 4.19 (d, $J = 3.3$ Hz), 4.73 (s, integration not determined), 4.75 (s, integration not determined), 4.83 (s, integration not determined), 4.90 (s, integration not determined), 4.94 (s, integration not determined), 5.11 (dd, $J = 9.8, 3.1$ Hz, integration not determined), 5.17 (s, integration not determined), 5.51 (d, $J = 3.6$ Hz, integration not determined); ¹³C NMR (1% CD₃CO₂D in D₂O, 151 MHz, selected signals) δ 63.19, 63.23, 63.3, 63.7, 69.4, 71.4, 71.6, 72.6, 72.65, 72.69, 72.78, 72.84, 72.9, 73.2, 73.4, 73.6, 74.2, 74.26, 74.28, 74.4, 75.5, 76.0, 76.1, 77.50, 77.53, 77.7, 77.9, 78.0, 79.1, 79.19, 79.22, 79.28, 79.33, 79.5, 80.1, 96.4, 96.5, 101.8, 102.3, 102.8, 102.86, 102.91, 103.0, 175.7, 176.1; ESI-HRMS found m/z 1217.3722 [M + Na]⁺, calcd for C₄₄H₇₄NaO₃₇ 1217.3807.

Allyl 2,3-Di-O-benzyl-4,6-benzylidene-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-3,6-di-O-benzyl-β-D-

mannopyranosyl-(1→4)-3,6-di-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-α-D-mannopyranoside (49). According to the representative procedure for selective deprotection of *p*-methoxybenzyl group, compound 33 (123 mg, 40.7 μmol) was treated with CAN (112 mg, 204.3 μmol) in CH₃CN/toluene/H₂O (90:5:5, v/v/v, 8.8 mL) at room temperature for 2 h to give compound 49 (88 mg, 30.3 μmol, 75%) after column chromatography on silica gel (hexane/EtOAc, 60:40): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 2.58 (d, $J = 2.7$ Hz, 1H), 2.76 (d, $J = 2.3$ Hz, 1H), 3.03 (td, $J = 9.7, 4.9$ Hz, 1H), 3.13–3.21 (m, 4H), 3.24 (ddd, $J = 9.5, 3.7, 2.5$ Hz, 1H), 3.26–3.32 (m, 3H), 3.35–3.40 (m, 3H), 3.42–3.60 (m, 9H), 3.61–3.75 (m, 8H), 3.75–3.80 (m, 2H), 3.88 (br t, 1H), 3.91–3.98 (m, 4H), 4.00–4.25 (m, 9H), 4.25–4.29 (2 overlapping d, 2H), 4.33–4.55 (m, 20H), 4.57 (d, $J = 12.4$ Hz, 1H), 4.60–4.80 (m, 17H), 4.84 (d, $J = 12.3$ Hz, 1H), 4.89 (d, $J = 2.3$ Hz, 1H), 5.15–5.19 (m, 1H), 5.21–5.26 (m, 1H), 5.47 (s, 1H), 5.82–5.91 (m, 1H), 7.10–7.39 (m, integration not determined), 7.42–7.46 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz, selected signals) δ 67.2, 67.5, 68.0, 68.1, 68.5, 69.0, 69.16, 69.24, 69.4, 69.5, 71.2, 71.4, 71.8, 71.9, 72.1, 72.3, 72.4, 72.5, 72.68, 72.73, 73.0, 73.35, 73.37, 73.42, 73.5, 74.0, 74.1, 74.7, 74.86, 74.89, 75.0, 75.2, 75.29, 75.34, 75.4, 75.7, 75.9, 76.05, 76.06, 77.12, 78.2, 78.4, 78.6, 79.4, 78.0, 80.8, 80.9, 81.5, 97.5, 100.3, 101.2, 101.5, 101.57, 101.62, 102.2, 117.2, 126.1, 126.9, 127.0, 127.06, 127.11, 127.15, 127.17, 127.3, 127.35, 127.43, 127.46, 127.49, 127.53, 127.55, 127.59, 127.62, 127.64, 127.66, 127.69, 127.73, 128.0, 128.05, 128.09, 128.13, 128.20, 128.24, 128.3, 128.4, 128.7, 133.8, 137.7, 138.1, 138.15, 138.21, 138.22, 138.3, 138.37, 138.40, 138.41, 138.6, 138.7, 138.8, 138.9, 139.07, 139.09, 139.13, 139.2; MS (MALDI, DHB) found m/z 2924.4 [M + Na]⁺, calcd for C₁₇₈H₁₈₈NaO₃₆ 2924.3.

Allyl 2,3-Di-O-benzyl-4,6-benzylidene-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2-O-acetyl-3,6-di-O-benzyl-β-D-mannopyranosyl-(1→4)-2-O-acetyl-3,6-di-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-α-D-mannopyranoside (50). According to the representative procedure for acetylation, alcohol 49 (89 mg, 30.7 μmol) was treated with acetic anhydride (0.4 mL) in pyridine (1.5 mL) in the presence of DMAP for 3 h at room temperature to give acetate 50 (84 mg, 28.1 μmol, 91%) after column chromatography on silica gel (hexane/EtOAc, 65:35): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 1.94–2.10 (2 overlapping s, 6H), 3.02 (td, $J = 9.7, 4.9$ Hz, 1H), 3.10–3.22 (m, 5H), 3.27–3.32 (m, 3H), 3.34–3.56 (m, 13H), 3.58–3.67 (m, 5H), 3.68–3.72 (2 overlapping d, 2H), 3.74–3.79 (m, 2H), 3.90–3.97 (m, 4H), 3.97–4.05 (2 overlapping t, 2H), 4.07–4.16 (m, 3H), 4.16–4.23 (m, 3H), 4.25–4.31 (2 overlapping d, 2H), 4.36–4.61 (m, 23H), 4.61–4.77 (m, 15H), 4.83 (d, $J = 12.2$ Hz, 1H), 4.88 (d, $J = 2.4$ Hz, 1H), 5.13–5.19 (m, 1H), 5.19–5.26 (m, 1H), 5.35 (d, $J = 3.6$ Hz, 1H), 5.38 (d, $J = 3.6$ Hz, 1H), 5.47 (s, 1H), 5.81–5.91 (m, 1H), 7.09–7.39 (m, integration not determined), 7.41–7.46 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz, selected signals) δ 21.0, 67.2, 67.9, 68.5, 68.6, 68.7, 68.8, 68.9, 69.15, 69.22, 69.4, 69.5, 71.3, 71.4, 72.11, 72.13, 72.2, 72.4, 72.5, 72.7, 73.1, 73.3, 73.35, 73.38, 73.40, 74.1, 74.2, 74.5, 74.6, 74.7, 74.9, 75.1, 75.29, 75.34, 75.4, 75.6, 75.7, 75.80, 75.83, 76.1, 76.2, 77.2, 78.0, 78.1, 78.4, 78.6, 78.8, 80.7, 81.3, 97.5, 99.5, 99.7, 101.2, 101.3, 101.4, 101.7, 102.1, 117.1, 126.1, 126.86, 126.90, 127.07, 127.10, 127.16, 127.19, 127.23, 127.27, 127.30, 127.32, 127.35, 127.42, 127.45, 127.52, 127.54, 127.59, 127.62, 127.63, 127.66, 127.72, 127.73, 127.96, 128.00, 128.02, 128.05, 128.07, 128.10, 128.13, 128.16, 128.21, 128.24, 128.27, 128.32, 128.7, 133.9, 137.7, 138.26, 138.30, 138.37, 138.41, 138.5, 138.66, 138.73, 138.8, 138.99, 139.04, 139.06, 139.08, 139.16, 139.20, 170.3; MS (MALDI, DHB) found m/z 3008.4 [M + Na]⁺, calcd for C₁₈₂H₁₉₂NaO₃₈ 3008.3.

2,3-Di-O-benzyl-4,6-benzylidene-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2-O-acetyl-3,6-di-O-benzyl-β-D-mannopyranosyl-(1→4)-2-O-acetyl-3,6-di-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranosyl-(1→4)-2,3,6-tri-O-benzyl-β-D-mannopyranose (51). According to the representative procedure for deallylation, compound 50 (71

mg, 23.8 μmol) was treated with a catalytic amount of PdCl_2 in $\text{MeOH}/\text{CH}_2\text{Cl}_2$ (1:1, v/v, 3.6 mL) at room temperature for 4 h to give compound **51** (43 mg, 14.6 μmol , 61%) after column chromatography on silica gel (hexane/EtOAc, 60:40): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 2.01 (s, integration not determined), 2.02 (s, integration not determined), 2.03 (s, integration not determined), 2.65 (br s, integration not determined), 3.02 (td, $J = 9.6$, 4.8 Hz, integration not determined), 3.10–3.22 (m, integration not determined), 3.22–3.58 (m, integration not determined), 3.58–3.72 (m, integration not determined), 3.74 (t, $J = 2.9$ Hz, integration not determined), 3.76–3.78 (m, integration not determined), 3.88–3.95 (m, integration not determined), 3.96–4.05 (m, integration not determined), 4.07–4.23 (m, integration not determined), 4.24–4.32 (2 overlapping d, integration not determined), 4.36–4.82 (m, integration not determined), 4.90–4.96 (m, integration not determined), 5.22 (d, $J = 2.8$ Hz, integration not determined), 5.34–5.37 (m, integration not determined), 5.39 (d, $J = 3.5$ Hz, integration not determined), 5.41 (d, $J = 3.5$ Hz, integration not determined), 5.47 (s, integration not determined), 7.09–7.38 (m, integration not determined), 7.41–7.45 (m, integration not determined); ^{13}C NMR (CDCl_3 , 151 MHz, selected signals) δ 21.0, 67.2, 68.5, 68.6, 68.7, 68.8, 68.9, 69.20, 69.22, 69.3, 69.48, 69.51, 69.6, 71.3, 71.4, 71.5, 72.06, 72.13, 72.2, 72.4, 72.5, 72.7, 73.05, 73.08, 73.13, 73.35, 73.39, 73.40, 73.5, 73.6, 73.9, 74.12, 74.14, 74.19, 74.22, 74.5, 74.6, 74.66, 74.70, 74.89, 74.91, 75.0, 75.07, 75.11, 75.3, 75.4, 75.67, 75.74, 75.8, 75.9, 76.0, 76.1, 76.15, 77.16, 78.0, 78.4, 78.6, 78.8, 80.3, 80.7, 81.2, 93.0, 93.4, 99.5, 99.7, 101.16, 101.22, 101.26, 101.29, 101.7, 102.1, 126.1, 126.86, 126.90, 126.93, 127.07, 127.11, 127.16, 127.19, 127.24, 127.26, 127.32, 127.34, 127.43, 127.45, 127.50, 127.52, 127.54, 127.59, 127.61, 127.63, 127.67, 127.70, 127.73, 127.76, 127.77, 127.80, 127.9, 128.00, 128.02, 128.06, 128.07, 128.10, 128.13, 128.18, 128.20, 128.21, 128.25, 128.27, 128.30, 128.35, 128.40, 128.5, 128.8, 137.7, 138.07, 138.11, 138.25, 138.34, 138.36, 138.42, 138.5, 138.63, 138.64, 138.7, 138.8, 138.89, 138.92, 138.98, 139.00, 139.03, 139.06, 139.09, 139.2, 170.3; MS (MALDI, DHB) found m/z 2968.3 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{175}\text{H}_{188}\text{NaO}_{38}$ 2968.3.

β -D-Mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4)-D-mannopyranose (6**).** According to the representative procedure for global deprotection of benzyl, benzylidene, and *p*-methoxybenzyl groups, compound **51** (12 mg, 4.07 μmol) was subjected to catalytic hydrogenation with 10% Pd/C (12 mg) in THF/MeOH/AcOH (2.0:1.0:0.3, v/v/v, 1.65 mL) at room temperature for 23 h to give compound **6** (3.0 mg, 2.43 μmol , 60%) after reversed-phase chromatography on a C18 column (1% AcOH in H_2O). Heptasaccharide **6** was isolated as a mixture of regioisomers resulting from the migration of the acetyl groups: white hygroscopic solid; ^1H NMR (1% $\text{CD}_3\text{CO}_2\text{D}$ in D_2O , 600 MHz) δ 2.16 (s, integration not determined), 2.181 (s, integration not determined), 2.183 (s, integration not determined), 2.194 (s, integration not determined), 2.198 (s, integration not determined), 2.204 (s, integration not determined), 3.40–3.68 (m, integration not determined), 3.69–4.07 (m, integration not determined), 4.08–4.14 (m, integration not determined), 4.16–4.21 (m, integration not determined), 4.72 (s, integration not determined), 4.75 (s, integration not determined), 4.82 (s, integration not determined), 4.83 (s, integration not determined), 4.893 (s, integration not determined), 4.899 (s, integration not determined), 4.903 (s, integration not determined), 4.909 (s, integration not determined), 4.93 (s, integration not determined), 4.94 (s, integration not determined), 5.04 (dd, $J = 9.7$, 3.0 Hz, integration not determined), 5.06–5.13 (m, integration not determined), 5.17 (s, integration not determined), 5.41 (d, $J = 3.2$ Hz, integration not determined), 5.48 (d, $J = 3.5$ Hz, integration not determined), 5.49–5.52 (m, integration not determined); ^{13}C NMR (1% $\text{CD}_3\text{CO}_2\text{D}$ in D_2O , 150 MHz, selected signals) δ 63.1, 63.19, 63.24, 63.3, 63.7, 69.4, 71.3, 71.36, 71.40, 71.5, 71.6, 72.62, 72.64, 72.7, 72.75, 72.80, 72.88, 72.93, 73.2, 73.4, 73.6, 74.1, 74.2, 74.3, 74.4, 74.5, 75.5, 75.7, 75.8, 76.0, 76.08, 76.11, 77.5, 77.66, 77.69, 77.74, 77.86, 77.91, 78.0, 79.1, 79.18, 79.23, 79.3, 79.5, 79.9, 80.1, 96.4, 96.5, 101.3, 101.80, 101.84, 102.3, 102.86, 102.91, 103.0, 103.1, 175.5, 175.67,

175.70, 175.74, 176.1, 176.2; HRMS found m/z 1259.3822 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{46}\text{H}_{76}\text{NaO}_{38}$ 1259.3912.

β -D-Mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)-D-mannopyranose (4**).** According to the representative procedure for saponification, heptasaccharide **6** (7 mg, 5.66 μmol) was treated with NaOH (1.0 M, 100 μL) at room temperature for 2 h to give compound **4** (6.2 mg, 5.38 μmol , 95%) after lyophilization: white hygroscopic solid; ^1H NMR (D_2O , 600 MHz) δ 3.42–3.47 (m, integration not determined), 3.49–3.61 (m, integration not determined), 3.66 (dd, $J = 9.6$, 3.3 Hz, integration not determined), 3.71–4.02 (m), 4.07 (d, $J = 3.4$ Hz, integration not determined), 4.12–4.16 (m, integration not determined), 4.74 (s, integration not determined), 4.76 (s, integration not determined), 4.92 (s, integration not determined), 5.19 (s, integration not determined); ^{13}C NMR (D_2O , 151 MHz, selected signals) δ 63.2, 63.7, 69.4, 71.7, 72.7, 72.9, 73.2, 73.4, 73.6, 74.1, 74.2, 74.4, 75.5, 77.5, 77.8, 79.1, 79.2, 79.27, 79.34, 79.5, 96.4, 96.6, 102.87, 102.93; HRMS found m/z 1175.3623 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{42}\text{H}_{72}\text{NaO}_{36}$ 1175.3701.

Allyl 2,3-Di-O-Benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (55**).** According to the representative procedure for β -selective mannosylation, the reaction of compounds **38** (883 mg, 0.96 mmol) and **14** (663 mg, 1.23 mmol) was performed, followed by quench with Et_3N and $(\text{EtO})_3\text{P}$, to give compound **55** (784 mg, 0.58 mmol, 61%) after chromatography on silica gel (toluene/EtOAc, 95.5:4.5): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 3.05 (td, $J = 9.7$, 4.9, 1H), 3.19 (dt, $J = 9.5$, 3.1 Hz, 1H), 3.30 (dd, $J = 9.2$, 3.0 Hz, 1H), 3.40 (dd, $J = 9.9$, 3.0 Hz, 1H), 3.47–3.58 (m, 3H), 3.62–3.72 (m, 3H), 3.73 (d, $J = 3.0$ Hz, 1H), 3.76–3.81 (m, 2H), 3.91–3.99 (m, 3H), 4.04 (t, $J = 9.6$ Hz, 1H), 4.12–4.19 (m, 2H), 4.21–4.30 (t overlapped with d, 2H), 4.41 (d, $J = 12.1$ Hz, 1H), 4.45 (d, $J = 11.9$ Hz, 1H), 4.48–4.55 (m, 4H), 4.59 (d, $J = 12.3$ Hz, 1H), 4.62–4.81 (m, 9H), 4.85–4.92 (2 overlapping d, 2H), 5.13–5.21 (m, 1H), 5.21–5.28 (m, 1H), 5.48 (s, 1H), 5.79–5.94 (m, 1H), 7.09–7.41 (m, integration not determined), 7.42–7.47 (m, 2H); ^{13}C NMR (CDCl_3 , 151 MHz) δ 67.2, 67.9, 68.5, 69.25, 69.32, 71.3, 72.2, 72.4, 72.5, 72.7, 73.3, 73.5, 74.2, 74.9, 75.4, 75.6, 75.7, 76.0, 76.1, 77.1, 78.37, 78.42, 78.6, 80.7, 97.5, 101.3, 101.5, 102.2, 117.8, 126.1, 126.9, 127.10, 127.13, 127.2, 127.35, 127.37, 127.42, 127.5, 127.57, 127.63, 127.70, 127.73, 127.8, 128.0, 128.07, 128.09, 128.12, 128.21, 128.24, 128.3, 128.7, 133.8, 137.7, 138.3, 138.37, 138.42, 138.5, 138.8, 139.07, 139.10, 139.3; MS (MALDI, DHB) found m/z 1375.6 $[\text{M} + \text{Na}]^+$; calcd for $\text{C}_{84}\text{H}_{88}\text{NaO}_{16}$: 1375.6.

Allyl 2,3,6-Tri-O-Benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (56**).** According to the representative procedure for reductive ring-opening of benzylidene acetal, trisaccharide **55** (317 mg, 234 μmol) was reduced with Et_3SiH (115 μL , 712 μmol) in the presence of TfOH (70 μL , 791 μmol) at -78°C for 15 h, followed by treatment with TBAF (2.2 mL, 1 M solution in THF) at room temperature for 2 h, to give compound **56** (138 mg, 102 μmol , 44%) after column chromatography on silica gel (toluene/EtOAc, 92:8): colorless oil; ^1H NMR (CDCl_3 , 600 MHz) δ 2.68 (d, $J = 1.0$ Hz, 1H), 3.14 (dd, $J = 9.4$, 2.8 Hz, 1H), 3.20 (dt, $J = 9.6$, 4.8, 1H), 3.25 (ddd, $J = 9.5$, 4.0, 2.4, 1H), 3.33 (dd, $J = 9.2$, 2.8, 1H), 3.50–3.61 (m, 4H), 3.61–3.67 (2 overlapping dd, 2H), 3.69 (d, $J = 2.9$ Hz, 1H), 3.72–3.79 (m, 3H), 3.89–3.99 (m, 3H), 4.13–4.24 (m, 3H), 4.30 (d, $J = 12.0$, 1H), 4.34 (d, $J = 11.7$, 1H), 4.37–4.48 (m, 5H), 4.49–4.55 (d overlapped with 2 s, 3H), 4.56–4.80 (m, 9H), 4.86–4.91 (m, 2H), 5.13–5.20 (m, 1H), 5.20–5.28 (m, 1H), 5.80–5.93 (m, 1H), 7.12–7.39 (m, integration not determined); ^{13}C NMR (CDCl_3 , 151 MHz) δ 67.9, 68.8, 69.3, 69.5, 71.1, 71.35, 71.38, 72.3, 72.6, 72.7, 73.3, 73.5, 73.7, 74.1, 74.2, 74.6, 74.7, 75.3, 75.5, 75.6, 75.7, 76.1, 78.3, 80.7, 81.8, 97.5, 101.4, 101.6, 117.2, 127.10, 127.12, 127.2, 127.3, 127.38, 127.44, 127.5, 127.55, 127.62, 127.7, 127.75, 127.77, 127.9, 128.08, 128.12, 128.13, 128.21, 128.22, 128.25, 128.31, 128.4, 133.9, 138.10, 138.11, 138.3, 138.4, 138.5, 139.0, 139.1, 139.2, 139.3; MS (MALDI, DHB) found m/z 1377.6 $[\text{M} + \text{Na}]^+$, calcd for $\text{C}_{84}\text{H}_{90}\text{NaO}_{16}$ 1377.6.

Allyl 2,3-Di-O-Benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (57). According to the representative procedure for β -selective mannosylation, the reaction of compounds 56 (308 mg, 227 μ mol) and 14 (160 mg, 296 μ mol) was performed, followed by quench with Et₃N and (EtO)₃P, to give compound 57 (262 mg, 147 μ mol, 65%) after chromatography on silica gel (toluene/EtOAc, 94:6): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 3.03 (td, *J* = 9.7, 4.9 Hz, 1H), 3.13–3.19 (m, 1H), 3.21 (dt, *J* = 9.4, 3.0 Hz, 1H), 3.27–3.35 (2 overlapping dd, 2H), 3.38 (dd, *J* = 9.9, 2.9 Hz, 1H), 3.43–3.51 (2 overlapping dd, 2H), 3.51–3.58 (m, 3H), 3.61–3.68 (2 overlapping dd, 2H), 3.69–3.74 (m, 3H), 3.75–3.80 (m, 2H), 3.91–3.98 (m, 3H), 4.03 (t, *J* = 9.6 Hz, 1H), 4.10–4.26 (m, 5H), 4.28 (d, *J* = 12.1 Hz, 1H), 4.37–4.42 (2 overlapping d, 2H), 4.46–4.54 (m, 7H), 4.56–4.83 (m, 13H), 4.86–4.91 (2 overlapping d, 2H), 5.14–5.19 (m, 1H), 5.20–5.26 (m, 1H), 5.47 (s, 1H), 5.80–5.92 (m, 1H), 7.09–7.39 (m, integration not determined), 7.42–7.46 (m, 2H); ¹³C NMR (CDCl₃, 151 Hz) δ 67.2, 67.9, 68.5, 69.2, 69.3, 69.4, 71.4, 72.1, 72.3, 72.4, 72.6, 72.7, 73.3, 73.4, 74.1, 74.2, 74.9, 75.4, 75.5, 75.6, 75.7, 75.9, 76.1, 76.2, 77.1, 78.3, 78.4, 78.6, 80.8, 80.9, 97.5, 101.3, 101.4, 101.7, 102.2, 117.2, 126.1, 126.90, 126.93, 127.07, 127.12, 127.15, 127.18, 127.35, 127.37, 127.42, 127.45, 127.54, 127.6, 127.66, 127.70, 127.73, 127.96, 127.99, 128.05, 128.09, 128.14, 128.20, 128.21, 128.24, 128.3, 128.7, 133.9, 137.7, 138.3, 138.4, 138.47, 138.54, 138.8, 139.09, 139.11, 139.14, 139.3; MS (MALDI, DHB) found *m/z* 1807.8 [M + Na]⁺, calcd for C₁₁₁H₁₁₆NaO₂₁ 1807.8.

Allyl 2,3-Di-O-Benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (58). According to the representative procedure for trans-acetalation, compound 57 (1.502 g, 0.84 mmol) was treated with neopentyl glycol (438 mg, 4.21 mmol) in the presence of camphorsulfonic acid (88 mg, 0.38 mmol) for 24 h at room temperature to give compound 58 (1.119 g, 0.66 mmol, 78%) after chromatography on silica gel (toluene/EtOAc, 74:26): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 2.25 (d, *J* = 2.0 Hz, 1H), 2.36 (br s, 1H), 3.03–3.09 (m, 2H), 3.19–3.23 (m, 2H), 3.26 (dd, *J* = 9.5, 2.8 Hz, 1H), 3.32 (dd, *J* = 9.2, 2.8 Hz, 1H), 3.40 (dd, *J* = 11.5, 6.5 Hz, 1H), 3.51–3.59 (m, 4H), 3.61–3.69 (m, 3H), 3.69–3.73 (m, 2H), 3.73–3.80 (m, 4H), 3.91–3.98 (m, 2H), 4.10–4.25 (m, 5H), 4.26–4.30 (2 overlapping d, 2H), 4.35 (d, *J* = 11.8 Hz, 1H), 4.37–4.43 (2 overlapping d, 2H), 4.46 (s, 1H), 4.48–4.54 (m, 5H), 4.56–4.81 (m, 11H), 4.84 (d, *J* = 12.4 Hz, 1H), 4.86–4.91 (2 overlapping d, 2H), 5.14–5.19 (m, 1H), 5.21–5.28 (m, 1H), 5.80–5.92 (m, 1H), 7.09–7.41 (m, integration not determined); ¹³C NMR (CDCl₃, 151 MHz) δ 62.7, 67.2, 67.9, 69.3, 69.4, 71.0, 71.3, 72.4, 72.45, 72.52, 72.7, 73.3, 73.4, 73.5, 73.8, 74.08, 74.10, 74.4, 75.39, 75.41, 75.43, 75.5, 75.6, 75.7, 76.3, 76.5, 78.2, 80.3, 80.7, 82.0, 97.5, 101.2, 101.4, 101.5, 117.1, 126.9, 127.0, 127.08, 127.11, 127.14, 127.27, 127.33, 127.37, 127.43, 127.52, 127.54, 127.6, 127.67, 127.71, 127.76, 127.80, 127.82, 127.88, 127.92, 128.07, 128.11, 128.13, 128.18, 128.19, 128.23, 128.3, 128.4, 133.8, 137.7, 138.25, 138.32, 138.38, 138.40, 138.65, 138.68, 138.9, 139.1, 139.2, 139.3; MS (MALDI, DHB) found *m/z* 1719.7 [M + Na]⁺, calcd for C₁₀₄H₁₁₂NaO₂₁ 1719.8.

Allyl 2,3,6-Tri-O-Benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (54). According to the representative procedure for selective monobenzylation, diol 58 (1.047 g, 0.62 mmol) was heated with Bu₂SnO (307 mg, 1.23 mmol) under reflux in toluene, followed by treatment with BnBr (295 μ L, 2.47 mmol, 4.0 equiv) and CsF (422 mg, 2.78 mmol) under reflux for 6 h, to give compound 54 (896 mg, 0.50 mmol, 81%) after column chromatography on silica gel (toluene/EtOAc, 88:12): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 2.69 (d, *J* = 1.4 Hz, 1H), 3.13 (dd, *J* = 9.5, 2.8 Hz, 1H), 3.16–3.25 (m, 3H), 3.28–3.35 (2 overlapping dd, 2H), 3.49–3.59 (m, 6H), 3.61–3.67 (2 overlapping dd, 2H), 3.68–3.74 (3 overlapping d, 3H), 3.74–3.79 (m, 2H), 3.89–3.97 (m, 3H), 4.13–4.19 (m, 3H), 4.22 (t, *J* = 9.1 Hz, 1H), 4.25–4.29 (2 overlapping d, 2H), 4.33 (d, *J* = 11.9 Hz, 1H), 4.37–4.43 (m, 5H), 4.46–4.54 (m, 6H), 4.56–4.63 (2 overlapping d, 2H), 4.63–

4.80 (m, 9H), 4.82 (d, *J* = 12.4 Hz, 1H), 4.85–4.91 (2 overlapping d, 2H), 5.13–5.19 (m, 1H), 5.21–5.27 (m, 1H), 5.82–5.91 (m, 1H), 7.10–7.38 (m, integration not determined); ¹³C NMR (CDCl₃, 151 MHz) δ 67.9, 68.7, 69.3, 69.35, 69.40, 71.1, 71.3, 72.2, 72.4, 72.6, 72.7, 73.3, 73.37, 73.38, 73.6, 74.05, 74.10, 74.12, 74.5, 74.6, 75.3, 75.38, 75.44, 75.5, 75.6, 75.7, 76.1, 76.2, 78.2, 80.75, 80.77, 81.8, 97.5, 101.4, 101.5, 101.6, 117.1, 127.0, 127.05, 127.09, 127.18, 127.22, 127.3, 127.4, 127.5, 127.6, 127.65, 127.68, 127.72, 127.8, 127.9, 127.96, 128.04, 128.06, 128.11, 128.17, 128.19, 128.21, 128.3, 128.4, 133.8, 138.06, 138.10, 138.3, 138.40, 138.44, 138.5, 138.9, 139.08, 139.10, 139.13, 139.3; MS (MALDI, DHB) found *m/z* 1809.7 [M + Na]⁺, calcd for C₁₁₁H₁₁₈NaO₂₁ 1809.8.

Allyl 2,3-Di-O-Benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-benzoyl-3,6-di-O-benzyl- β -D-glucopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (59). According to the representative procedure for glycosylation, acceptor 54 (454 mg, 254 μ mol) was treated with trichloroacetimidate 34 (725 mg, 381 μ mol) in the presence of TMSOTf (6.8 μ L, 38 μ mol) to give compound 59 (696 mg, 197 μ mol, 78%) after column chromatography on silica gel (hexane/EtOAc, 70:30): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 2.85–2.91 (m, 1H), 3.03 (td, *J* = 9.6, 5.0 Hz, 2H), 3.11–3.23 (m, 6H), 3.23–3.32 (m, 5H), 3.36–3.41 (m, 2H), 3.41–3.68 (m, 17H), 3.68–3.73 (3 overlapping d, 3H), 3.73–3.78 (m, 2H), 3.90–3.97 (m, 3H), 3.97–4.09 (m, 4H), 4.09–4.27 (m, 11H), 4.32–4.36 (d overlapped with s, 2H), 4.36–4.54 (m, 17H), 4.54–4.61 (3 overlapping d, 3H), 4.61–4.84 (m, 21H), 4.84–4.89 (2 overlapping d, 2H), 5.0 (d, *J* = 12.1 Hz, 1H), 5.14–5.25 (m, 3H), 5.47 (s, 1H), 5.81–5.91 (m, 1H), 6.85–6.90 (m, 2H), 6.93–7.40 (m, integration not determined), 7.42–7.47 (m, 2H), 7.50–7.58 (m, 1H), 7.82–7.89 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz, selected signals) δ 67.2, 67.9, 68.2, 68.49, 68.51, 69.18, 69.24, 69.3, 69.35, 69.42, 71.3, 72.1, 72.2, 72.35, 72.40, 72.6, 72.7, 73.3, 73.35, 73.42, 73.7, 74.0, 74.05, 74.07, 74.2, 74.3, 74.9, 75.0, 75.36, 75.39, 75.43, 75.46, 75.51, 75.6, 75.7, 75.9, 76.0, 76.08, 76.14, 76.2, 76.3, 77.1, 78.0, 78.2, 78.4, 78.6, 80.56, 80.63, 80.7, 80.76, 80.80, 80.9, 97.5, 100.4, 101.2, 101.30, 101.34, 101.5, 101.6, 102.2, 117.1, 126.1, 126.8, 126.9, 126.95, 126.97, 127.05, 127.08, 127.16, 127.20, 127.22, 127.30, 127.34, 127.40, 127.42, 127.48, 127.52, 127.58, 127.60, 127.63, 127.7, 127.8, 127.91, 127.93, 127.96, 127.98, 128.04, 128.1, 128.2, 128.3, 128.7, 129.8, 129.9, 132.9, 133.9, 137.7, 138.1, 138.3, 138.36, 138.38, 138.42, 138.44, 138.5, 138.7, 138.76, 138.82, 139.0, 139.06, 139.09, 139.13, 139.2, 139.26, 139.29, 139.32, 164.9; MS (MALDI, DHB) found *m/z* 3551.1 [M + Na]⁺, calcd for C₂₁₉H₂₂₆NaO₂₄ 3550.5.

Allyl 2,3-Di-O-Benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl- β -D-glucopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (60). According to the representative procedure for debenzoylation, compound 59 (93 mg, 26.3 μ mol) was treated with LiAlH₄ (8 mg, 210.8 μ mol) in Et₂O to give compound 60 (60 mg, 17.5 μ mol, 67%) after chromatography on silica gel (hexane/EtOAc, 70:30): colorless oil; ¹H NMR (CDCl₃, 600 MHz) δ 3.02 (td, *J* = 9.6, 5.0 Hz, 1H), 3.08–3.42 (m, 16H), 3.42–3.58 (m, 10H), 3.58–3.67 (m, 3H), 3.67–3.82 (m, 10H), 3.90–3.98 (m, 3H), 4.02 (t, *J* = 9.5 Hz, 1H), 4.09–4.19 (m, 6H), 4.19–4.28 (m, 6H), 4.31–4.43 (m, 8H), 4.44–4.54 (m, 14H), 4.57 (d, *J* = 12.3 Hz, 1H), 4.59–4.82 (m, 21H), 4.85–4.91 (2 overlapping d, 2H), 4.99 (d, *J* = 12.0 Hz, 1H), 5.15–5.18 (m, 1H), 5.20–5.26 (m, 1H), 5.47 (s, 1H), 5.83–5.91 (m, 1H), 7.08–7.39 (m, integration not determined), 7.42–7.45 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz, selected signals) δ 67.2, 67.9, 68.5, 68.6, 69.2, 69.3, 69.4, 69.45, 69.52, 71.36, 71.38, 72.1, 72.2, 72.3, 72.4, 72.6, 72.7, 73.2, 73.3, 73.37, 73.40, 73.41, 73.6, 74.08, 74.12, 74.16, 74.19, 74.7, 74.8, 74.9, 75.0, 75.2, 75.29, 75.33, 75.39, 75.41, 75.48, 75.52, 75.6, 75.7, 75.9, 76.06, 76.12, 76.13, 76.2, 77.0, 77.1, 78.3, 78.4, 78.6, 80.75, 80.80, 80.84, 81.9, 82.4, 97.5, 101.0, 101.3, 101.4, 101.6, 101.7, 101.8, 102.2, 103.7, 117.2, 126.1, 126.7, 126.8, 126.9, 126.96, 126.99,

127.01, 127.06, 127.11, 127.16, 127.19, 127.3, 127.4, 127.46, 127.54, 127.6, 127.66, 127.70, 127.96, 127.98, 128.01, 128.03, 128.05, 128.08, 128.13, 128.19, 128.21, 128.23, 128.26, 128.30, 128.7, 133.9, 137.7, 137.8, 138.0, 138.3, 138.42, 138.44, 138.48, 138.54, 138.7, 138.8, 138.99, 139.01, 139.09, 139.10, 139.2, 139.27, 139.31, 139.5; MS (MALDI, DHB) found m/z 3446.9 $[M + Na]^+$, calcd for $C_{212}H_{222}NaO_{41}$ 3446.5.

Allyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-3,6-di-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (61). According to the representative procedure for gluco to manno epimerization, compound **60** (88 mg, 25.7 μ mol, 1.0 equiv) was subjected to oxidation with DMSO (1.0 mL) and acetic anhydride (0.5 mL) at room temperature for 16 h, and subsequent reduction with L-Selectride (1 M in THF, 77 μ L, 77 μ mol) at -78 °C for 30 min, to give compound **61** (51 mg, 14.9 μ mol, 58%) after column chromatography on silica gel (hexane/EtOAc, 65:35): colorless oil; 1H NMR ($CDCl_3$, 600 MHz) δ 2.68 (br s, 1H), 3.03 (td, J = 9.6, 4.9 Hz, 1H), 3.10–3.23 (m, 6H), 3.26–3.40 (m, 7H), 3.41–3.59 (m, 12H), 3.60–3.78 (m, 11H), 3.89 (br s, 1H), 3.91–3.97 (m, 3H), 4.00–4.07 (2 overlapping t, 2H), 4.10–4.32 (m, 13H), 4.35–4.54 (m, 21H), 4.55–4.82 (m, 23H), 4.85–4.90 (2 overlapping d, 2H), 5.14–5.19 (m, 1H), 5.19–5.28 (m, 1H), 5.47 (s, 1H), 5.75–5.95 (m, 1H), 7.03–7.39 (m, integration not determined), 7.41–7.47 (m, 2H); ^{13}C NMR ($CDCl_3$, 151 MHz, selected signals) δ 67.2, 67.9, 68.0, 68.5, 69.1, 69.2, 69.3, 69.36, 69.42, 71.4, 71.8, 72.1, 72.2, 72.3, 72.36, 72.38, 72.6, 72.7, 73.2, 73.3, 73.35, 73.38, 73.39, 73.41, 74.0, 74.12, 74.13, 74.7, 74.9, 75.1, 75.2, 75.28, 75.33, 75.36, 75.41, 75.43, 75.5, 75.6, 75.7, 75.9, 76.0, 76.1, 76.2, 77.1, 78.3, 78.4, 78.6, 79.3, 80.75, 80.77, 80.84, 81.4, 97.5, 100.2, 101.3, 101.4, 101.57, 101.61, 101.7, 102.2, 117.2, 126.1, 126.90, 126.94, 126.98, 127.0, 127.07, 127.11, 127.16, 127.20, 127.3, 127.35, 127.41, 127.44, 127.45, 127.54, 127.60, 127.62, 127.70, 127.73, 127.95, 127.98, 128.01, 128.05, 128.09, 128.11, 128.18, 128.20, 128.23, 128.3, 128.7, 133.9, 137.7, 138.3, 138.4, 138.45, 138.48, 138.51, 138.6, 138.7, 138.8, 139.05, 139.11, 139.2, 139.25, 139.27; MS (MALDI, DHB) found m/z 3446.1 $[M + Na]^+$, calcd for $C_{212}H_{222}NaO_{41}$ 3446.5.

Allyl 2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl-3,6-di-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranoside (53). According to the representative procedure for acetylation, alcohol **61** (92 mg, 26.9 μ mol) was treated with acetic anhydride (0.35 mL) in pyridine in the presence of DMAP (catalytic amount) for 3 h at room temperature to give acetate **53** (85 mg, 24.5 μ mol, 91%) after column chromatography on silica gel (hexane/EtOAc, 67:33): colorless oil; 1H NMR ($CDCl_3$, 600 MHz) δ 2.00 (s, 3H), 3.02 (td, J = 9.6, 4.9 Hz, 1H), 3.10–3.23 (m, 6H), 3.25–3.33 (m, 6H), 3.35–3.59 (m, 14H), 3.60–3.73 (m, 8H), 3.74–3.79 (m, 2H), 3.90–3.98 (m, 4H), 4.02 (t, J = 9.5 Hz, 1H), 4.07–4.31 (m, 12H), 4.34–4.54 (m, 22H), 4.55–4.82 (m, 23H), 4.84–4.91 (2 overlapping d, 2H), 5.13–5.20 (m, 1H), 5.20–5.26 (m, 1H), 5.38 (d, J = 3.3 Hz, 1H), 5.47 (s, 1H), 5.80–5.92 (m, 1H), 7.07–7.38 (m), 7.41–7.46 (m, 2H); ^{13}C NMR ($CDCl_3$, 151 MHz, selected signals) δ 21.0, 67.2, 67.9, 68.5, 68.8, 69.0, 69.1, 69.2, 69.4, 69.5, 71.36, 71.40, 72.0, 72.1, 72.15, 72.24, 72.4, 72.6, 72.7, 73.0, 73.3, 73.35, 73.36, 73.38, 73.39, 74.05, 74.10, 74.14, 74.2, 74.5, 74.9, 75.0, 75.3, 75.4, 75.5, 75.6, 75.7, 75.82, 75.84, 76.08, 76.12, 76.17, 76.21, 77.1, 78.1, 78.3, 78.4, 78.6, 80.7, 80.76, 80.82, 81.4, 97.5 ($^1J_{CH}$ = 167.5 Hz), 99.8 ($^1J_{CH}$ = 159.5 Hz), 101.1 ($^1J_{CH}$ = 155.4 Hz), 101.3, 101.4 ($^1J_{CH}$ = 157.3 Hz), 101.5 ($^1J_{CH}$ = 158.2 Hz), 101.6 ($^1J_{CH}$ = 157.0 Hz), 101.7 ($^1J_{CH}$ = 157.6 Hz), 102.2 ($^1J_{CH}$ = 157.0 Hz), 117.1, 126.1, 126.8, 126.85, 126.89, 126.96, 126.98, 127.04, 127.06, 127.10, 127.14, 127.2, 127.26, 127.30, 127.34, 127.37, 127.42, 127.44, 127.50, 127.54, 127.57, 127.58, 127.65, 127.68, 127.70, 127.73, 127.74, 127.95, 127.97, 127.99, 128.05, 128.08, 128.09, 128.13, 128.18, 128.21, 128.24, 128.3, 128.7, 133.9, 137.7,

138.3, 138.35, 138.39, 138.43, 138.45, 138.53, 138.54, 138.78, 138.81, 139.08, 139.11, 139.14, 139.17, 139.23, 139.25, 139.27, 170.3; MS (MALDI, DHB) found m/z 3448.1 $[M + Na]^+$, calcd for $C_{214}H_{224}NaO_{42}$ 3448.5.

2,3-Di-O-benzyl-4,6-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl-3,6-di-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-benzyl- α -D-mannopyranose (62). According to the representative procedure for deallylation, compound **53** (104 mg, 30.0 μ mol) was treated with a catalytic amount of $PdCl_2$ in $MeOH/CH_2Cl_2$ (1:1, v/v, 4.6 mL) at room temperature for 3 h to give compound **62** (61 mg, 17.8 μ mol, 59%) after column chromatography on silica gel (hexane/EtOAc, 62:38): colorless oil; 1H NMR ($CDCl_3$, 600 MHz) δ 2.00 (s, integration not determined), 2.64 (d, J = 3.5 Hz, integration not determined), 3.02 (td, J = 9.7, 4.9 Hz, integration not determined), 3.10–3.25 (m, integration not determined), 3.25–3.60 (m, integration not determined), 3.60–3.73 (m, integration not determined), 3.75 (t, J = 2.8 Hz, integration not determined), 3.77–3.79 (m, integration not determined), 3.90–4.05 (m, integration not determined), 4.06–4.32 (m, integration not determined), 4.34–4.84 (m, integration not determined), 4.92–4.98 (2 overlapping d, integration not determined), 5.23 (t, J = 2.9 Hz, integration not determined), 5.39 (d, J = 3.4 Hz, integration not determined), 5.47 (s, integration not determined), 7.09–7.40 (m, integration not determined), 7.41–7.46 (m, integration not determined); ^{13}C NMR ($CDCl_3$, 151 Hz, selected signals) δ 21.0, 67.2, 68.5, 68.8, 68.9, 69.1, 69.2, 69.36, 69.42, 69.5, 69.6, 71.4, 71.5, 72.0, 72.1, 72.2, 72.26, 72.30, 72.4, 72.5, 72.7, 73.0, 73.4, 73.5, 73.6, 74.0, 74.06, 74.11, 74.14, 74.16, 74.23, 74.5, 74.8, 74.9, 75.0, 75.30, 75.32, 75.39, 75.41, 75.5, 75.7, 75.8, 75.9, 76.08, 76.11, 76.2, 77.1, 77.2, 78.1, 78.4, 78.6, 80.4, 80.55, 80.59, 80.7, 80.8, 81.4, 93.0, 93.4, 99.7, 101.1, 101.2, 101.3, 101.5, 101.6, 101.7, 102.2, 126.1, 126.8, 126.85, 126.89, 126.98, 127.01, 127.07, 127.11, 127.15, 127.22, 127.27, 127.33, 127.34, 127.37, 127.42, 127.5, 127.58, 127.63, 127.69, 127.73, 127.8, 128.0, 128.05, 128.09, 128.15, 128.18, 128.21, 128.3, 128.4, 128.5, 128.7, 128.8, 130.9, 137.7, 138.1, 138.35, 138.39, 138.42, 138.46, 138.51, 138.54, 138.78, 139.0, 139.1, 139.17, 139.23, 170.3; MS (MALDI, DHB) found m/z 3448.5 $[M + Na]^+$, calcd for $C_{211}H_{220}NaO_{42}$ 3448.5.

β -D-Mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranosyl-(1 \rightarrow 4)- β -D-mannopyranose (8). According to the representative procedure for global deprotection of benzyl, benzylidene and *p*-methoxybenzyl groups, compound **62** (25 mg, 7.29 μ mol) was subject to hydrogenation by catalysis of 10% Pd/C in THF/MeOH/AcOH (2.0:1.0:0.3, v/v/v, 2.13 mL) at room temperature to give compound **8** (6.2 mg, 1.81 μ mol, 63%) after reversed-phase chromatography ($CH_3CN/H_2O/AcOH$, 2.5/97.5/0.1). Octasaccharide **8** was isolated as a mixture of regioisomers resulting from the migration of the acetyl group: white hygroscopic solid; 1H NMR (1% CD_3CO_2D in D_2O , 600 MHz) δ 2.19 (s, integration not determined), 2.20 (s, integration not determined), 3.41–3.60 (m, integration not determined), 3.60–3.68 (m, integration not determined), 3.71–4.08 (m, integration not determined), 4.09–4.15 (m, integration not determined), 4.20 (d, J = 3.6 Hz, integration not determined), 4.72 (s, integration not determined), 4.73 (s, integration not determined), 4.75 (s, integration not determined), 4.83 (s, integration not determined), 4.91 (s, integration not determined), 4.95 (s, integration not determined), 5.12 (dd, J = 9.8, 3.1 Hz, integration not determined), 5.18 (d, J = 1.5 Hz, integration not determined), 5.51 (d, J = 3.8 Hz, integration not determined); ^{13}C NMR (1% CD_3CO_2D in D_2O , 151 MHz, selected signals) δ 23.0, 23.3, 63.2, 63.3, 63.7, 69.4, 71.4, 71.6, 72.6, 72.66, 72.70, 72.79, 72.84, 72.9, 73.2, 73.4, 73.6, 74.1, 74.2, 74.26, 74.28, 74.4, 75.5, 76.0, 76.1, 77.50, 77.53, 77.7, 77.9, 78.0, 79.10, 79.13, 79.22, 79.23, 79.27, 79.32, 79.5, 80.1, 96.4, 96.6, 101.8, 102.3, 102.8, 102.86, 102.91, 103.0, 175.7, 176.1; HRMS found m/z 1357.4569 $[M + H]^+$, calcd for $C_{50}H_{85}O_{42}$ 1357.4515.

β -D-Mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)-D-mannopyranose (7). According to the representative procedure for saponification, octasaccharide **8** (6.1 mg, 4.49 μ mol) was treated with 1.0 M NaOH in water at room temperature for 1.5 h to give compound **7** (5.0 mg, 3.80 μ mol, 85%) after lyophilization: white hygroscopic solid; ^1H NMR (D_2O , 600 MHz) δ 3.45 (ddd, $J = 9.6, 7.1, 2.4$ Hz, integration not determined), 3.49–3.53 (m, integration not determined), 3.54–3.62 (m, integration not determined), 3.67 (dd, $J = 9.7, 3.2$ Hz, integration not determined), 3.71–4.03 (m, integration not determined), 4.08 (d, $J = 3.3$ Hz, integration not determined), 4.12–4.16 (m, integration not determined), 4.75 (s, integration not determined), 4.77 (s, integration not determined), 4.93 (s, integration not determined), 5.19 (d, $J = 1.2$ Hz, integration not determined); ^{13}C NMR (D_2O , 125 MHz) δ 63.2, 63.7, 69.4, 71.7, 72.6, 72.67, 72.71, 72.9, 73.2, 73.4, 73.6, 74.1, 74.2, 74.4, 75.5, 77.5, 77.8, 79.15, 79.21, 79.27, 79.34, 79.5, 96.4, 96.6, 102.87, 102.93; HRMS found m/z 1315.4443 [$\text{M} + \text{H}$] $^+$, calcd for $\text{C}_{48}\text{H}_{83}\text{O}_{41}$ 1315.4410.

Independent Synthesis of β -D-Mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)-D-mannitol (52). To a solution of compound **51** (36 mg, 12.2 μ mol) in THF/EtOH (1:2, v/v, 1.5 mL) was added NaBH_4 (10 mg) in one portion. The mixture was stirred at room temperature for 17 h, and water was added at 0 °C. The mixture was diluted with CH_2Cl_2 , washed with water and brine, dried over MgSO_4 , and concentrated in vacuo. The residue was purified by column chromatography on silica gel (toluene/EtOAc, 75:25) to give a mannitol compound **64** (22 mg, 7.5 μ mol, 61%).

The above-prepared mannitol compound was treated with 10% Pd/C (22 mg) in THF/MeOH/AcOH (2.0:1.0:0.3, v/v/v, 2.97 mL) under an atmosphere of hydrogen at room temperature for 23 h. The mixture was passed through a pad of Celite to remove the catalyst, and the filtrate was concentrated under reduced pressure. The residue was dissolved in $\text{H}_2\text{O}/\text{MeOH}/\text{AcOH}$ (3:6:1, v/v/v, 3.00 mL) and treated with 10% Pd/C (22 mg), and the suspension was stirred under an atmosphere of hydrogen at room temperature for 17 h. The suspension was filtered through a pad of Celite and concentrated under reduced pressure. Reversed-phase chromatography ($\text{CH}_3\text{CN}/\text{H}_2\text{O}/\text{AcOH}$, 2:98:1) of the residue gave a debenzylated compound (5.3 mg).

The above-prepared debenzylated compound (5.3 mg) was treated in 0.1 M NaOH (0.7 mL) at room temperature for 1.0 h. The solution was passed through a short cation-exchange column (Dowex 50W-X8 H^+ form) and washed with water. The eluent was concentrated under reduced pressure and lyophilized to give compound **52** (4.7 mg, 3.46 μ mol, 47%): white hydroscopic solid; ^1H NMR (D_2O , 600 MHz) δ 3.46 (ddd, $J = 9.7, 7.1, 2.5$ Hz, 1H), 3.55–3.62 (m, 6H), 3.66–3.72 (m, 3H), 3.73–3.88 (m, 18H), 3.90–3.99 (m, 9H), 4.06–4.10 (m, 2H), 4.12–4.19 (m, 5H), 4.75 (s, integration not determined), 4.76 (s, integration not determined), 4.78 (s, integration not determined), 4.83 (s, integration not determined); ^{13}C NMR (D_2O , 151 MHz) δ 63.2, 63.5, 63.7, 65.1, 65.8, 69.4, 72.2, 72.6, 72.7, 72.9, 73.2, 73.3, 74.1, 74.2, 74.3, 75.5, 77.5, 77.7, 79.1, 79.25, 79.33, 79.7, 102.7, 102.86, 102.92; HRMS found m/z 1155.4041 [$\text{M} + \text{H}$] $^+$, calcd for $\text{C}_{42}\text{H}_{75}\text{O}_{36}$ 1155.4038.

Independent Synthesis of β -D-Mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)- β -D-mannopyranosyl-(1→4)-D-mannitol (63). To a solution of compound **61** (93 mg, 27.1 μ mol, 1.0 equiv) in MeOH/ CH_2Cl_2 (1:1, v/v, 4.0 mL) was added a catalytic amount of PdCl_2 . The mixture was stirred at room temperature for 3 h, diluted with EtOAc, and filtered through a small pad of silica gel. The filtrate was washed with saturated aqueous NaHCO_3 , water and brine, dried over MgSO_4 , and concentrated in vacuo. The residue was purified by column chromatography on silica gel (hexane/EtOAc, 55:45) to give a deallylation product (54 mg).

The above-prepared deallylated product (54 mg) was dissolved in THF/EtOH (1:2, v/v, 2.1 mL), and NaBH_4 (10 mg) was added in one portion. The mixture was stirred at room temperature for 15 h, and water was added at 0 °C. The mixture was diluted with EtOAc, washed with water and brine, dried over MgSO_4 , and concentrated in vacuo. The residue was purified by column chromatography on silica gel (toluene/EtOAc, 70:30) to give a mannitol product **65** (40 mg, 11.8 μ mol, 43%).

The above-prepared mannitol product (31 mg, 9.15 μ mol, 1.0 equiv) was treated with acetic anhydride (0.25 mL) and DMAP (catalytic amount) in pyridine (1.0 mL) for 3.0 h at room temperature and then quenched by addition of MeOH at 0 °C. After removal of the solvent by evaporation in vacuo, the residue was dissolved in Et₂O. The mixture was washed with water and brine, dried over MgSO_4 , and concentrated in vacuo. The residue was purified by column chromatography on silica gel (hexane/EtOAc, 65:35) to give the acetylation product **66** (26 mg, 7.40 μ mol, 81%).

The above-prepared product (11 mg, 3.13 μ mol) was treated with 10% Pd/C (11 mg) in THF/MeOH/AcOH (2.0:1.0:0.3, v/v/v, 1.32 mL) under an atmosphere of hydrogen at room temperature for 23 h. The mixture was passed through a pad of Celite to remove the catalyst and then concentrated in vacuo. The residue was dissolved in $\text{H}_2\text{O}/\text{MeOH}/\text{AcOH}$ (3:6:1, v/v/v, 1.73 mL) and treated with 10% Pd/C (11 mg) under an atmosphere of hydrogen at room temperature for 17 h. The suspension was filtered through a pad of Celite and concentrated under reduced pressure. Reversed-phase chromatography ($\text{CH}_3\text{CN}/\text{H}_2\text{O}/\text{AcOH}$, 5:95:1) of the residue gave the debenzylation product (1.9 mg).

The above-prepared product was treated in 0.1 M NaOH (0.4 mL) at room temperature for 1.5 h. The solution was passed through a short cation-exchange column (Dowex 50W-X8 H^+ form) and washed with water. The eluent was concentrated under reduced pressure and lyophilized to give compound **63** (1.6 mg, 1.21 μ mol, 39%): white hydroscopic solid; ^1H NMR (D_2O , 600 MHz) δ 3.45 (ddd, $J = 9.6, 7.1, 2.4$ Hz, 1H), 3.54–3.62 (m, 7H), 3.65–3.71 (m, 3H), 3.72–3.88 (m, 21H), 3.89–3.99 (m, 10H), 4.05–4.10 (m, 2H), 4.11–4.19 (m, 6H), 4.75 (s, integration not determined), 4.76 (s, integration not determined), 4.77 (s, integration not determined), 4.82 (s, integration not determined); ^{13}C NMR (D_2O , 151 MHz) δ 63.2, 63.5, 63.7, 65.1, 65.8, 69.4, 72.2, 72.6, 72.7, 72.9, 73.2, 73.3, 74.15, 74.17, 74.3, 75.5, 77.5, 77.8, 79.2, 79.3, 79.4, 79.72, 79.73, 102.7, 102.87, 102.93; HRMS found m/z 1317.4609 [$\text{M} + \text{H}$] $^+$, calcd for $\text{C}_{48}\text{H}_{85}\text{O}_{41}$ 1317.4566.

■ ASSOCIATED CONTENT

📁 Supporting Information

NMR spectra of synthetic compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

■ AUTHOR INFORMATION

Corresponding Author

*E-mail: (J.-M.F.) jmfang@ntu.edu.tw, (C.-H.W.) chwong@gate.sinica.edu.tw.

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

We thank Academia Sinica and Yuen-Foong-Yu Co., Ltd., for financial support. We also thank Dr. Che-Hsiung Hsu, Dr. Chi-Fon Chang, and Ms. Yi-Ping Huang for technical support of NMR measurements.

■ REFERENCES

- (1) (a) Tang, Z.-Z.; Cheng, S.-J. *Bull. Bot. Res.* **1984**, *4*, 141–146. (b) Bao, X.-S.; Shun, Q.-S.; Chen, L.-Z. *Chinese Medicinal Dendrobium*; Fudan University Press and Shanghai Medical University Press: Shanghai, 2001.

- (2) Lu, S.-F.; Guo, G.-J.; Cai, Y.-P. *Chin. Trad. Herbal Drugs* **2005**, *5*, 790–793.
- (3) (a) Yang, L.; Wang, Z.; Xu, L. *J. Chromatogr. A* **2006**, *1104*, 230–237. (b) Lee, Y. H.; Park, J. D.; Baek, N. I.; Kim, S. I.; Ahn, B. Z. *Planta Med.* **1995**, *61*, 178–180. (c) Honda, C.; Yamaki, M. *Phytochemistry* **2000**, *53*, 987–990. (d) Yang, H.; Chou, G. X.; Wang, Z. T.; Hu, Z. B.; Xu, L. S. *Asian Nat. Prod. Res.* **2004**, *53*, 35–38. (e) Wrigley, T. C. *Nature* **1960**, *188*, 1108. (f) Morita, H.; Fujiwara, M.; Yoshida, N.; Kobayashi, J. *Tetrahedron* **2000**, *56*, 5801–5805. (g) Zhao, W. M.; Ye, Q. H.; Tan, X. J.; Jiang, H. L.; Li, X. Y.; Chen, K. X.; Kinghorn, A. D. *J. Nat. Prod.* **2001**, *64*, 1196–1200. (h) Ye, Q. H.; Qin, G. W.; Zhao, W. M. *Phytochemistry* **2002**, *61*, 885–890. (i) Ye, Q. H.; Zhao, W. M. *Planta Med.* **2002**, *68*, 723–729.
- (4) Hsieh, Y. S.-Y.; Chien, C.; Liao, S. K.-S.; Liao, S.-F.; Hung, W.-T.; Yang, W.-B.; Lin, C.-C.; Cheng, T.-J. R.; Chang, C.-C.; Fang, J.-M.; Wong, C.-H. *Bioorg. Med. Chem.* **2008**, *16*, 6054–6068.
- (5) (a) Nicola, N. A.; Metcalf, D.; Matsumoto, M.; Johnson, G. R. *J. Biol. Chem.* **1983**, *258*, 9017–9023. (b) Metcalf, D. *Science* **1985**, *229*, 16–22. (c) Welte, K.; Platzer, E.; Lu, L.; Gabrilove, J. L.; Levi, E.; Mertelsmann, R.; Moore, M. A. *Proc. Natl. Acad. Sci. U.S.A.* **1985**, *82*, 1526–1530. (d) Nomura, H.; Imazeki, I.; Oheda, M.; Kubota, N.; Tamura, M.; Ono, M.; Ueyama, Y.; Asano, S. *EMBO J.* **1986**, *5*, 871–876. (e) Nagata, S.; Tsuchiya, M.; Asano, S.; Kaziro, Y.; Yamazaki, T.; Yamamoto, O.; Hirata, Y.; Kubota, N.; Oheda, M.; Nomura, H.; Ono, M. *Nature* **1986**, *319*, 415–418. (f) Souza, L. M.; Boone, T. C.; Gabrilove, J.; Lai, P. H.; Zsebo, K. M.; Murdock, D. C.; Chazin, V. R.; Bruszewski, J.; Lu, H.; Chen, K. K.; Barendt, J.; Platzer, E.; Moore, M. A. S.; Mertelsmann, R.; Welte, K. *Science* **1986**, *232*, 61–65. (g) Welte, K.; Gabrilove, J.; Bronchud, M. H.; Platzer, E.; Morstyn, G. *Blood* **1996**, *88*, 1907–1929.
- (6) (a) Viret, F.; Goncalves, A.; Tarpin, C.; Chabannon, C.; Viens, P. *Bull. Cancer* **2006**, *93*, 463–471. (b) Franzke, A. *Cytokine Growth Factor Rev.* **2006**, *17*, 235–244. (c) Panopoulos, A. D.; Watowich, S. S. *Cytokine* **2008**, *42*, 277–288.
- (7) Xiao, B. G.; Lu, C. Z.; Link, H. *J. Cell. Molec. Med.* **2007**, *11*, 1272–1290.
- (8) Wong, C.-H.; Yang, W.-B.; Cheng, T.-J.; Hsieh, Y. S.-Y.; Chien, C.; Lin, C.-C.; Wen, H.-Y.; Fang, J.-M. Structure and Bioactivity of the Polysaccharides and Oligomers in Medicinal Plant *Dendrobium huoshanense*. US Patent 8,354,127 B2, 2013.
- (9) Crich, D.; Banerjee, A.; Yao, Q. *J. Am. Chem. Soc.* **2004**, *126*, 14930–14934.
- (10) (a) Crich, D.; Sun, S. X. *J. Am. Chem. Soc.* **1998**, *120*, 435–436. (b) Crich, D.; Sun, S. X. *Tetrahedron* **1998**, *54*, 8321–8348. (c) Crich, D.; Smith, M. *J. Am. Chem. Soc.* **2001**, *123*, 9015–9020. (d) Crich, D. *J. Carbohydr. Chem.* **2002**, *21*, 667–690.
- (11) Crich, D.; Li, W. J.; Li, H. M. *J. Am. Chem. Soc.* **2004**, *126*, 15081–15086.
- (12) Crich, D. *Acc. Chem. Res.* **2010**, *43*, 1144–1153.
- (13) Koto, S.; Asami, K.; Hirooka, M.; Nagura, K.; Takizawa, M.; Yamamoto, S.; Okamoto, N.; Sato, M.; Tajima, H.; Yoshida, T.; Nonaka, N.; Sato, T.; Zen, S.; Yago, K.; Tomonaga, F. *Bull. Chem. Soc. Jpn.* **1999**, *72*, 765–777.
- (14) Oshitari, T.; Shibasaki, M.; Yoshizawa, T.; Tomita, M.; Takao, K.; Kobayashi, S. *Tetrahedron* **1997**, *53*, 10993–11006.
- (15) Schmidt, R. R.; Kinzy, W. *Adv. Carbohydr. Chem. Biochem.* **1994**, *50*, 21–123.
- (16) (a) Danishefsky, S. J.; Hu, S.; Cirillo, P. F.; Eckhardt, M.; Seeberger, P. H. *Chem.—Eur. J.* **1997**, *3*, 1617–1628. (b) Wang, Z. G.; Warren, J. D.; Dudkin, V. Y.; Zhang, X. F.; Iserloh, U.; Visser, M.; Eckhardt, M.; Seeberger, P. H.; Danishefsky, S. J. *Tetrahedron* **2006**, *62*, 4954–4978. (c) Ghosh, S.; Misra, A. K. *Tetrahedron: Asymmetry* **2010**, *21*, 725–730. (d) Ghosh, S.; Misra, A. K. *Tetrahedron: Asymmetry* **2010**, *21*, 2755–2761.
- (17) Crich, D.; Smith, M.; Yao, Q. J.; Picione, J. *Synthesis* **2001**, 323–326.
- (18) Debenham, S. D.; Toone, E. J. *Tetrahedron: Asymmetry* **2000**, *11*, 385–387.
- (19) (a) Sliedregt, L. A. J. M.; Vandermaarel, G. A.; Vanboom, J. H. *Tetrahedron Lett.* **1994**, *35*, 4015–4018. (b) Codee, J. D. C.; van den Bos, L. J.; Litjens, R. E. J. N.; Overkleeft, H. S.; van Boeckel, C. A. A.; van Boom, J. H.; van der Marel, G. A. *Tetrahedron* **2004**, *60*, 1057–1064.
- (20) Jayaprakash, K. N.; Radhakrishnan, K. V.; Fraser-Reid, B. *Tetrahedron Lett.* **2002**, *43*, 6953–6955.
- (21) Andrews, M. A.; Gould, G. L. *Carbohydr. Res.* **1992**, *229*, 141–147.
- (22) Nagashima, N.; Ohno, M. *Chem. Lett.* **1987**, 141–144.
- (23) Csuk, R.; Prell, E.; Korb, C.; Kluge, R.; Strohl, D. *Tetrahedron* **2010**, *66*, 467–472.
- (24) Deninno, M. P.; Etienne, J. B.; Duplantier, K. C. *Tetrahedron Lett.* **1995**, *36*, 669–672.
- (25) Ogawa, T.; Kaburagi, T. *Carbohydr. Res.* **1982**, *110*, C12–C15.
- (26) Albright, J. D.; Goldman, L. *J. Am. Chem. Soc.* **1965**, *87*, 4214–4216.
- (27) Liu, K. K. C.; Danishefsky, S. J. *J. Org. Chem.* **1994**, *59*, 1892–1894.
- (28) Bock, K.; Pedersen, C. *J. Chem. Soc., Perkin Trans. 2* **1974**, 293–299.
- (29) Hannuksela, T.; du Penhoat, C. H. *Carbohydr. Res.* **2004**, *339*, 301–312.
- (30) (a) Willför, S.; Sundberg, K.; Tenkanen, M.; Holmbom, B. *Carbohydr. Polym.* **2008**, *72*, 197–210. (b) Ekholm, F. S.; Ardá, A.; Eklund, P.; André, S.; Gabius, H. J.; Jiménez-Barbero, J.; Leino, R. *Chem.—Eur. J.* **2012**, *18*, 14392–14405.
- (31) Pekari, K.; Tailler, D.; Weingart, R.; Schmidt, R. R. *J. Org. Chem.* **2001**, *66*, 7432–7442.
- (32) Sakagami, M.; Hamana, H. *Tetrahedron Lett.* **2000**, *41*, 5547–5551.
- (33) (a) Brown, H. C.; Krishnam, S. *J. Am. Chem. Soc.* **1972**, *94*, 7159–7161. (b) Lichtenthaler, F. W.; Lergenmuller, M.; Peters, S.; Varga, Z. *Tetrahedron: Asymmetry* **2003**, *14*, 727–736.